

Nephroprotective Effect of Aqueous Extract of Parsley against Nephrotoxicity Induced by Carbon Tetrachloride in the Male Rats

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ABSTRACT

Background: CCl_4 causes disorders in different body organs especially kidneys by generating free radicals. High exposure to CCl_4 can cause kidney damage. It produces renal diseases in human. Metabolism of CCl_4 involves in the production of free radicals through its activation by drug metabolizing enzymes located in the endoplasmic reticulum. Parsley has antioxidant, anti-inflammatory, anticancer activities, and probable immune boosting properties make it relevant in the traditional treatment of urinary tract infection, nephritis, and cystitis.

Objectives: The present study aimed to investigate the protective effects of oral administration of aqueous extract of parsley against the histopathological and biochemical alterations induced in the kidney by CCl_4 of male rats.

Materials and Methods: 24 male rats were used for this study and divided into 4 groups. The first group was control group injected intraperitoneally with olive oil (1ml/kg of body weight) three time/week, the 2nd was injected intraperitoneally with CCl_4 (1ml/kg of body weight) three time/week, the 3rd was injected intraperitoneally with CCl_4 (1ml/kg of body weight) three time/week concurrently with receiving aqueous Parsley leaves extract (5g/kg body wt/day) by gastric intubation's, the 4th was injected intraperitoneally with CCl_4 (1ml/kg of body weight) three time/week concurrently with receiving aqueous Parsley leaves extract (20g/kg body wt/day) by gastric intubation's, respectively for 4 weeks. The Kidneys were dissected out and specimens were taken and processed for light microscopic examinations. Blood samples were obtained for assessment of serum urea, creatinine, and uric acid.

Results: The results showed a significant increase in serum urea, creatinine, uric acid, K^+ , and Cl^- concentrations and a significant decrease in serum Na^+ concentration in rats treated with CCl_4 compared with controls. Histopathological investigation of rats treated with CCl_4 showed degeneration of epithelial lining and disruption of brush borders of the proximal convoluted tubules and presence of epithelial debris inside their lumens. The renal corpuscle appeared with degeneration of the glomerulus and disrupted Bowman's capsule. Extensive perivascular infiltration of inflammatory cells and massive interstitial hemorrhage were seen. Co-administration of Parsley with CCl_4 significantly dose dependent improvement in the structural changes in the kidney and the serum urea, creatinine, uric acid, K^+ , and Cl^- concentrations were significantly declined and serum Na^+ concentration was significantly increased compared with CCl_4 treated group.

Conclusion: It can be concluded that, carbon tetrachloride has adverse effects on the kidney, it caused a serious pathophysiological changes in the kidney. Parsley aqueous extract was able to protect the kidney against these effects. So, the persons expose to CCl_4 should be advised to take parsley aqueous extract.

Keywords: CCl_4 , Parsley, Nephrotoxicity, Nephroprotective, Renal pathophysiological changes, Histopathology

INTRODUCTION

CCl_4 does not occur naturally, it is a clear liquid with sweet smell that can be detected at low levels [1, 2]. Volatile organic compounds such as CCl_4 are a class of solvents to which many

people are exposed occupationally and environmentally [2]. CCl_4 was formerly used for metal degreasing and as dry cleaning, fabric-spotting, and fire extinguisher fluids, grain fumigant and reaction medium. Because of its

harmful effects, these uses are now banned and it is only used in some industrial applications [2, 3]. The primary routes of potential human exposure to CCl₄ are inhalation, ingestion, and dermal contact [2, 4].

A number of reports clearly demonstrated that CCl₄ causes disorders in kidneys, liver, lungs, testis as well as in blood by generating free radicals [2, 5, 6]. In addition, report on various documented case studies established that CCl₄ produces renal diseases in human [7, 8]. A number of endogenous and exogenous nephropathy risk factors generate oxygen free radicals *in vivo*. Therefore the role of oxygen derived free radicals and lipid peroxidation has attracted considerable attention [2, 9, 10]. Metabolism of CCl₄ involves in the production of free radicals through its activation by drug metabolizing enzymes located in the endoplasmic reticulum [2, 11]. High exposure to CCl₄ can cause kidney damage [2, 4]. CCl₄ induces oxidative stress in many settings [12]; therefore, it might be expected to contribute to nephrotoxicity [2]. CCl₄ is one of such widely used environmental toxicant to experimentally induce animal models of acute nephrotoxicity and hepatic damages [2].

Medicinal plants and herbs play an important role in the prevention and treatment of kidney diseases. Parsley, a bright green biennial shrub is widely used traditionally as a food additive and herbal remedies for many ailments [13]. Parsley (*Petroselinum crispum*) is a member of Apiaceous family that has been employed in the food, pharmaceutical, perfume, and cosmetic industries [14]. Parsley leaves are used as culinary, garnishing and medicinal herb in the Mediterranean region of Southern Europe. Parsley extract was reported to produce a diuretic effect and good antioxidant activity [15, 16]. It has been reported that parsley has anti-inflammatory and probable immune boosting properties make it relevant in the traditional treatment of urinary tract infection, nephritis, cystitis and prevention of renal stones formation. Parsley increases diuresis by inhibiting the Na⁺/K⁺ - ATPase pump in the kidney, thereby enhancing sodium and water excretion while increasing potassium resorption [16, 17]. It is one of the most used medicinal plants to treat renal diseases [18], diabetes, cardiac diseases, and arterial hypertension [19]. laxative activities [20], antioxidant [21], antihepatotoxic, anti-hyperlipidemic, anti-hyperglycemic [22], anticoagulant [23],

antianemic, menorrhagic, and anti-microbial [24-26]. In popular medicine, parsley is used to treat various illnesses such as Alzheimer's disease, thrombosis and strokes. Parsley is widely employed against cardiovascular diseases [27, 28]. Also, parsley leaves were used for treatment of constipation, jaundice, colic, flatulence edema, rheumatism. It was used to treat lumbago, as a blood pressure regulator, impotence and nose bleed [29], eczema, knee, ache, impotence and bleeding [26, 29], nose bleeding, hematoma, skin blemishes due to its bleaching properties, halitosis, ear ache, otitis, and as an emenagogue favouring menstruation and alleviating its pains. It is also widely used as a galactofuge by lactating mothers to stop excessive milk production. Large amount can have uterotonic effect and therefore its use is contraindicated in pregnancy and ingestion of more than 10 drops a day of the oil may cause abortion [30, 31].

Parsley is a power house of nutrition, rich in tocopherol and vitamin A [22, 28, 32]. It contains starch, vitamins B, C, β-carotene and zinc. It is a good source of iron, calcium, phosphorous and antioxidants like luteolin, vitamin C, vitamin A and zinc and these may likely account for its hepato protective effect [33, 34]. Phytochemically, the leaves and seeds of *P. crispum* has been shown to contain high levels of essential oil known as apiole, while the tender buds contain psoralen and related compounds that can induce photosensitivity and these include xanthotoxin, flicusin, bergapten, majudin, heraclin and antimicrobial furocoumarins namely 8-methoxypsoralen, 5-methoxypsoralen, oxypuecedanin, isopimpinellin, 6'-acetyllopin and a new monoterpene glycoside [29, 35, 36]. Besides having significant nutritional value, parsley also exhibits antioxidant and neutralizing properties [28, 37]. The constituents of parsley which include ascorbic acid, carotenoids, flavo-noids, coumarins, apiole, various terpenoic compounds, phenyl propanoids, phthalides, furano coumarins, and tocopherol, have been chemically investigated. Phytochemical screening of parsley has revealed the presence of several classes of flavonoids [38]. Flavonols (kaempferol and quercetin) and flavones (apigenin and luteolin), which occur as glycosidic form in nature, are major flavonoids found in parsley (*Petroselinum crispum*) and other apiaceous vegetables [39]. Kaempferol and quercetin, which belong to flavonol group, possess a wide range of biochemical and pharmacological effects and have been recommended as chemopreventive agents or nutritional supplements [40, 41]. The predominant

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mechanism of their biological actions is thought to result from antioxidant activity, enzyme inhibition, and the capacity to scavenge free radicals [40, 42]. Therefore, it is speculated that the health promoting effect of parsley (*Petroselinum crispum*) may be due to its flavonol constituents and the content of flavonoid compounds in parsley is about 100 mg/100 g fresh weight [43].

Parsley is rich with an antioxidant arsenal that includes luteolin, flavonoid that searches out and eradicates free radicals in the body that cause oxidative stress in cells [28, 44]. Components of fresh parsley leaf scavenge superoxide anion in vitro [45], and methanol extracts of parsley scavenge hydroxyl radical [38]. Supplementation of diets with fresh leaf can increase antioxidant capacity of rat plasma [46] and decrease the oxidative stress in humans [21]. It has been reported that parsley alcoholic extract has a protective effect against toxicity induced by sodium valproate in male rats [47]. Parsley leaves are rich in Apigenin and its glucosidal flavonoids that were found to possess anti-inflammatory especially for renal inflammation; antioxidant and anticancer activities [48, 49]. Aqueous extract of parsley reduced the number of calcium oxalate deposits and therefore parsley can be used for kidney and bladder stones [50].

Antioxidants such as melatonin, ascorbate, α -tocopherol, silibinin, lazaroid, propionyl carnitine and superoxide dismutase/catalase, have been shown to ameliorate CCl₄-induced renal toxicity [2, 51]. However, according to our knowledge, the reported literatures on the protective effect of parsley against CCl₄ nephrotoxicity are still limited. Therefore, it has become a task to prevent nephropathy damage induced by CCl₄ by eliminating free radicals and prevent lipid peroxidation with a natural antioxidant rich plant like parsley. So, the aim of this study was to investigate the protective effects of oral administration of aqueous extract of parsley against the histopathological and biochemical alterations induced by CCl₄ in kidney of male F-344/NHsd Fischer rats.

MATERIALS AND METHODS

Materials

Chemicals

Carbon tetrachloride (CCl₄) was purchased from Sigma Aldrich, St Louis, MO. It was applied as 1ml/kg body weight of CCl₄ suspended in olive oil (1:9 v/v) i.p three a week

for 4 weeks as a modified method to the method of Marsillach et al., [52] and El-Baz et al., [53]. Other chemicals were xylene, formalin 10%, ethanol (absolute alcohol), chloroform, purified water, and paraffin wax. All these chemicals were obtained from the Chemistry department of the Faculty of Science – Zawia.

Plant Materials

The plain leaf parsley type daily collected from a vegetable market in Zawia (Libya), carefully washed under tap water and left to dry at room temperature. Then they were weighted and completely blended in distilled water (1:1w/v). Animals were received Parsley juice orally (Haidari *et al.*, [54] by gastric intubation's at two doses of 5g/Kg parsley or 20g/ kg/day for six weeks

Animals

24 male F-344/NHsd Fischer rats (14-15 weeks, 200±25gm) were used in this study. Rats were purchased from Animal Welfare House of Libyan National Medical Research Centre, Zawia, Libya. They were housed in plastic cages, 6 rats per cage, and left one week for acclimation, before commencing the experiment. The rats were kept in a room under standard conditions of ventilation, temperature (25 ± 4°C), humidity (65 ± 5 %) with light/dark cycle. The animals were provided with tap water *ad libitum* and feed with the standard commercial chow. The animal procedures were performed by Guidelines for Ethical Conduct in the Care and Use of Animals.

Methods

Experimental Design

After one week of acclimation, the animals were randomized and divided into four groups (6 rats for each) as follow:

- Group I (control group): This group included 6 animals that was provided with tap water and fed with a normal diet and injected intraperitoneal with 1 ml/kg body weight of olive oil for 4 weeks.
- Group II (CCl₄ group): The animals were injected intraperitoneal with 1 ml/kg body weight of CCl₄ three a week for 4 weeks.
- Group III (CCl₄/ (5g/kg) Parsley leaves extract co-administration): The animals were injected intraperitoneal with 1 ml/kg body weight of CCl₄ three a week concurrently with receiving aqueous Parsley leaves extract (5g/kg body wt/day) by gastric intubation's for 4 weeks.
- Group IV (CCl₄/ (20g/kg) Parsley leaves extract co-administration): The animals were injected

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intraperitoneal with 1 ml/kg body weight of CCl₄ three a week concurrently with receiving aqueous Parsley leaves extract (20g/kg body wt/day) by gastric intubation's for 4 weeks.

Blood Sampling

At the end of the experiment and 24 hours after the last dose, all animals were anaesthetized with ether and blood samples were collected by heart puncture. The samples were collected in a clean dry tube and centrifuged at 3000 rpm for 15 minutes then, the serum was separated and kept in a deep freezer at -20°C until biochemical measurements were carried out.

Biochemical Analysis

Serum urea measurement was based upon the cleavage of urea with urease [55]. Serum uric acid was determined according to Fossatti *et al.*, [56]. Serum creatinine was measured without protein precipitation [57]. Sodium and potassium ions levels were estimated spectro photometrically using commercial kits by the modified method described by Maruna and Trinder [58].

Histological Preparation

At the end of the experimentation and 24 hours after the last dose, all animals were anaesthetized with ether. The animals were immediately dissected, and small pieces of the kidney were quickly removed and fixed in formalin fluid. After fixation specimens were

dehydrated in an ascending series of alcohol, then were kept in terpineol for three days to ensure complete dehydration and clearing purposes. Cleared specimens were rinsed in three changes of xylol before embedding in paraffin wax (m.p.56-58°C). Three sections of 5 microns thick were taken from each kidney sample, each being at a distance of at least 500 microns from the proceeding one and mounted on clean slides without an adhesive medium. The sections were stained with Hematoxylin and Eosin and examined under a binocular light microscope [59].

Statistical Analysis

Results were expressed as mean \pm SD, Data were analyzed by one way ANOVA. The difference between means \pm SD was tested at $P < 0.05$ using Duncan's multiple range test. In all statistical tests, the probability level of $P < 0.05$ was considered significant.

RESULTS

The results showed a significant ($P < 0.01$) increase in serum urea, creatinine, uric acid, K⁺, Cl⁻ and concentrations and a significant decrease Na⁺ concentration in rats treated with CCl₄ compared with control group. Co-administration of Parsley with CCl₄, a dose dependent, significantly ($P < 0.01$) decreased serum urea, creatinine, uric acid, K⁺, and Cl⁻ concentrations and increased serum Na⁺ concentration compared to CCl₄ group (Table.1& Figure.1-6).

Table1. Effect of treatment of male rats with CCl₄ and aqueous extract of parsley on serum urea, creatinine, uric acid, Na⁺, K⁺, and Cl⁻ concentrations

Parameters	Groups			
	Control	CCl ₄	CCl ₄ + 5g/kg Parsley (CCl ₄ +5P)	CCl ₄ + 20g/kg Parsley (CCl ₄ +20P)
	Mean \pm SD	Mean \pm SD	Mean \pm SD	Mean \pm SD
Urea (mg/dl)	37.67 \pm 1.03	50.67 \pm 5.47**	42.83 \pm 0.75**##	38.33 \pm 1.37##
Creatinine (mg/dl)	0.37 \pm 0.01	0.71 \pm 0.09**	0.45 \pm 0.02**##	0.40 \pm 0.01**##
Uric Acid (mg/dl)	1.85 \pm 0.10	3.35 \pm 0.26**	2.56 \pm 0.14**##	1.96 \pm 0.13**##
Na ⁺ (mmol/L)	140.27 \pm 1.20	128.47 \pm 6.78**	140.00 \pm 0.91##	142.17 \pm 0.42##
K ⁺ (mmol/L)	4.80 \pm 0.40	8.01 \pm 0.79**	6.11 \pm 0.45**##	5.41 \pm 0.22**##
Cl ⁻ (mmol/L)	101.72 \pm 2.26	106.30 \pm 0.92**	106.23 \pm 0.46**	105.13 \pm 1.01**

All data are mean of 6 individuals. *: Significant differences as compared with control group ($P < 0.05$); **: Significant differences as compared with control group ($P < 0.01$); ##: Significant differences as compared with control group ($P < 0.01$).

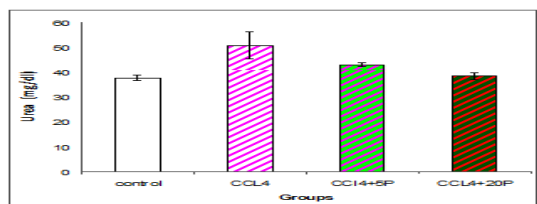


Figure1. Effect of treatment of male rats with CCl₄ and aqueous extract of parsley on serum urea concentration

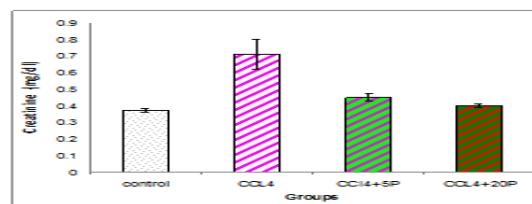


Figure2. Effect of treatment of male rats with CCl₄ and aqueous extract of parsley on serum creatinine concentration

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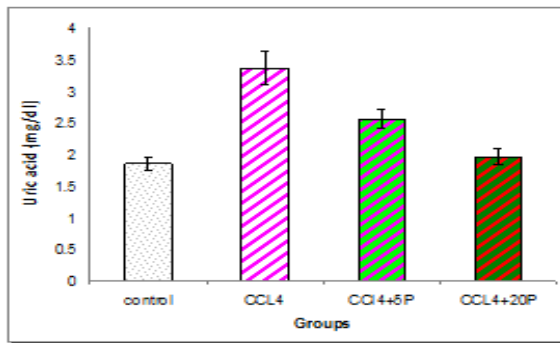


Figure3. Effect of treatment of male rats with CCl₄ and aqueous extract of parsley on serum uric acid concentration

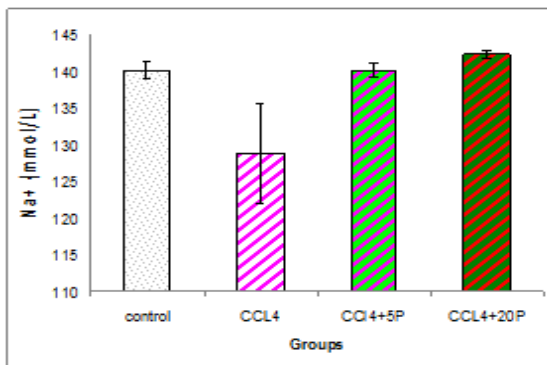


Figure4. Effect of treatment of male rats with CCl₄ and aqueous extract of parsley on serum Na⁺ ions concentration

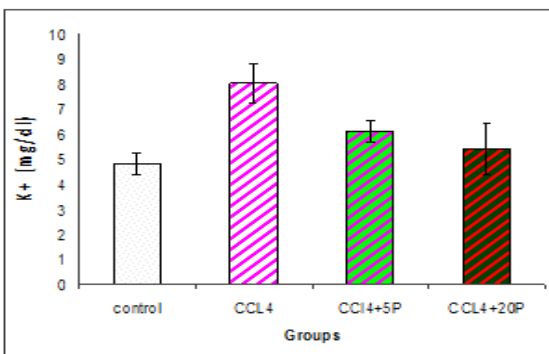


Figure5. Effect of treatment of male rats with CCl₄ and aqueous extract of parsley on serum K⁺ ions concentration

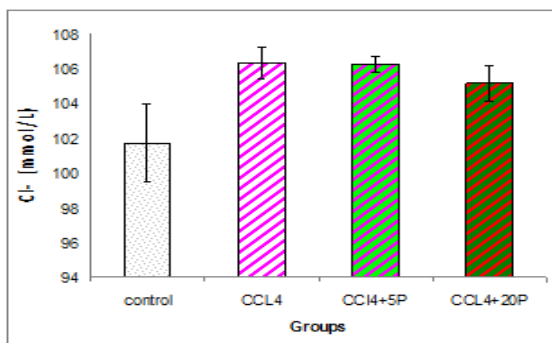


Figure6. Effect of treatment of male rats with CCl₄ and aqueous extract of parsley on serum Cl⁻ ions concentration

Histologically, by light microscopic examination, the kidney appeared with normal structures in normal control animals. The glomerulus appeared normal with intact Bowman's capsule and Bowman's space. The proximal convoluted tubules appeared with intact limiting basement membrane and their lumens showed intact brush borders. The distal convoluted tubules showed wide lumens with low cuboidal lining cells. An afferent arteriole appeared near the glomerulus (Fig. 7).

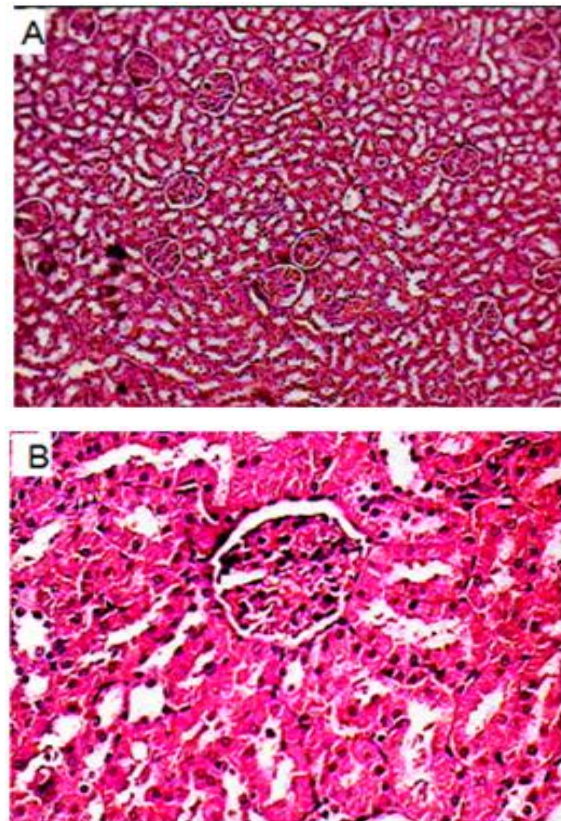


Figure7. Light micrograph of sections in the kidney of control rats (administered olive oil) showed the glomerulus appeared normal with intact Bowman's capsule and Bowman's space. The proximal convoluted tubules appeared with intact limiting basement membrane and their lumens showed intact brush borders. The distal convoluted tubules showed wide lumens with low cuboidal lining cells. An afferent arteriole appeared near the glomerulus. (Haematoxylin & Eosin; A: $\times 100$ & B $\times 400$).

In CCl₄ treated rats, the proximal convoluted tubules showed degenerated epithelial lining with disruption of their brush borders and presence of epithelial debris inside their lumens. The renal corpuscle appeared with degeneration of the glomerulus and disrupted Bowman's capsule. The afferent arteriole appeared near the glomerulus with thickening in its wall and degeneration of endothelial lining. Extensive perivascular infiltration of inflammatory cells

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and massive interstitial hemorrhage were seen (Fig. 8).

In animals co-administered with CCl₄ and 5g/kg body weight of Parsley aqueous extract, most of the proximal convoluted tubules appeared with normal epithelial lining with their brush borders, but some tubules appeared regenerating with disrupted brush borders. The glomerulus appeared more or less normal with intact

Bowman's capsule. Massive interstitial hemorrhage was seen (Fig. 9).

In animals co-administered with CCl₄ and 20g/kg body weight of Parsley aqueous extract, most of the proximal convoluted tubules appeared with normal epithelial lining and regained their brush borders. Some tubules still have diminished brush borders. The glomerulus appeared normal with intact Bowman's capsule (Fig. 10).

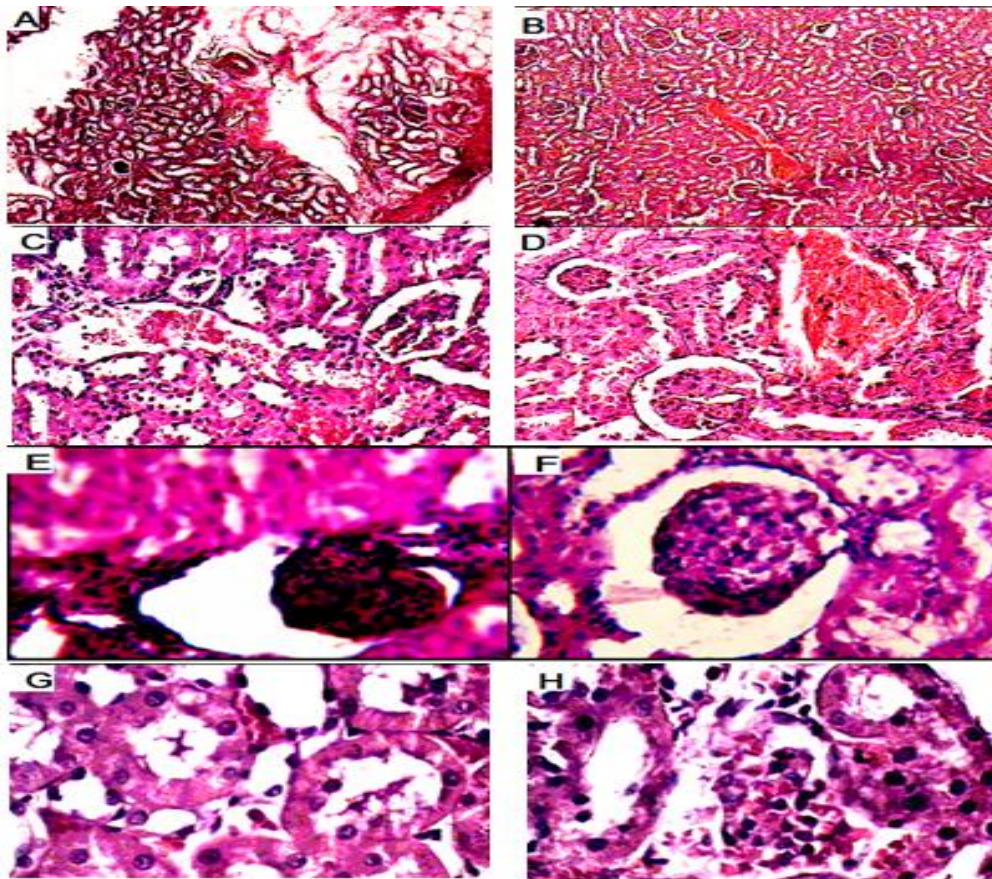


Figure 8. Light micrograph of sections in the kidney of CCl₄ treated rats. The proximal convoluted tubules showed degenerated epithelial lining with disruption of their brush borders and presence of epithelial debris inside their lumens. The renal corpuscle appeared with degeneration of the glomerulus and disrupted Bowman's capsule. Extensive perivascular infiltration of inflammatory cells and massive interstitial hemorrhage were seen. (Haematoxylin & Eosin; A&B × 100; C&D × 400; E-H × 1000)

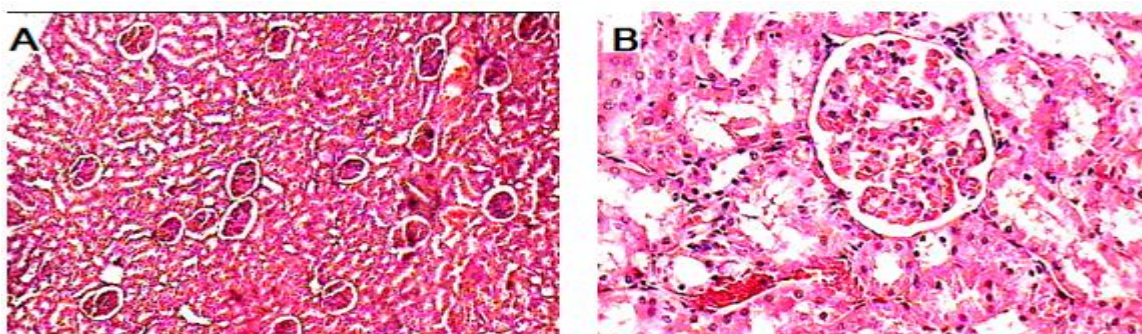


Figure 9. Light micrograph of a section in the kidney of CCl₄ treated rats co-administered 5g/kg body weight Parsley aqueous extract. Most of the proximal convoluted tubules appeared with normal epithelial lining with their brush borders, but some tubules appeared regenerating with disrupted brush borders. The glomerulus appeared more or less normal with intact Bowman's capsule. massive interstitial hemorrhage were seen (Haematoxylin & Eosin; A×100 & B×400).

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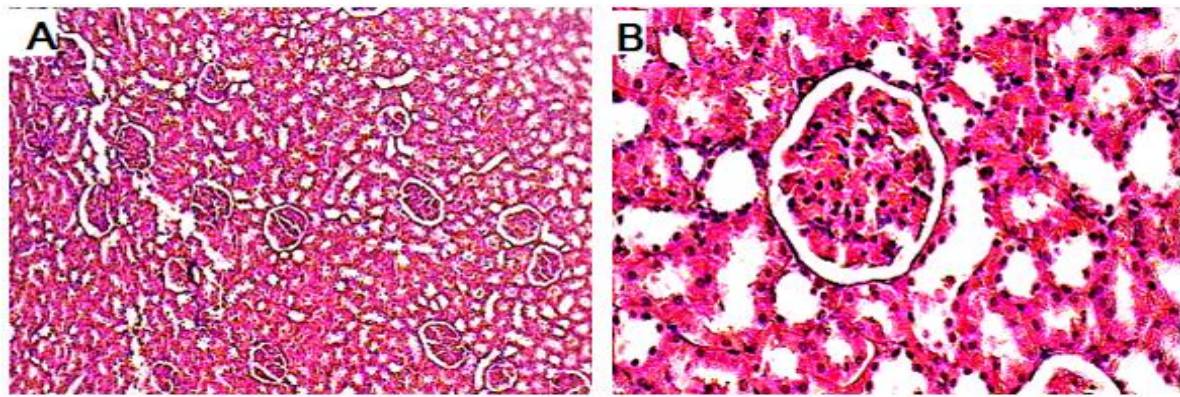


Figure 9. Light micrograph of a section in the kidney of CCl₄ treated rats co-administered 5g/kg body weight Parsley aqueous extract. Most of the proximal convoluted tubules appeared with normal epithelial lining and regained their brush borders. Some tubules still have diminished brush borders. The glomerulus appeared normal with intact Bowman's capsule (Haematoxylin & Eosin; A×100 & B×400).

DISCUSSION

The results of the present study showed a significant increase in the level of urea, creatinine, and uric acid in the group treated with 1 ml/kg of body weight of carbon tetrachloride. These results were consistent with the results of Makni *et al.*, [60] who reported that intraperitoneally injection of rats with 1ml/kg body weight of CCl₄ caused a significant increase in plasma creatinine, urea and uric acid levels. Kidney histological sections showed glomerular hypertrophy and tubular dilatation in CCl₄-treated rats. Adewole *et al.*, [2] demonstrated that chronic administration of CCl₄ caused marked impairment in renal function. Serum creatinine and BUN concentrations were significantly higher in CCl₄- treated rats which are consistent with lower creatinine and BUN clearance. CCl₄ is metabolized by cytochrome P450 2E1 to trichloromethyl radical (CCl₃·). CCl₃· and its highly reactive derivative, the trichloromethylperoxyl radical (Cl₃COO·), are assumed to initiate free radical-mediated lipid peroxidation leading to accumulation of lipid peroxidation products that causes renal injuries.

These radicals are capable of initiating a chain of lipid peroxidation reactions by abstracting hydrogen from polyunsaturated fatty acids. Peroxidation of lipids, particularly those containing polyunsaturated fatty acids, can dramatically change the properties of biological membranes, resulting in severe cell damage and play a significant role in pathogenesis of diseases [2, 61]. This phenomenon results in the generation of ROS, like superoxide anion O⁻, H₂O₂ and hydroxyl radical OH·. Evidence suggests that various enzymatic and non-enzymatic systems have been developed by mammalian cells to cope with ROS and other

free radicals [2, 62]. However, when a condition of oxidative stress establishes, the defense capacities against ROS becomes insufficient [2, 63]. Oxidative stress can promote the formation of a variety of vasoactive mediators that can affect renal function directly by initiating renal vasoconstriction or decreasing the glomerular capillary ultra filtration coefficient; and thus reducing glomerular filtration rate [2, 64]. In vitro and in vivo studies indicate that CCl₄ enhances lipid peroxidation, reduces renal microsomal NADPH cytochrome P450, and renal reduced/oxidized glutathione ratio (GSH/GSSG) in kidney cortex as well as renal microsomes and mitochondria [65].

In the present study, treatment of rats with CCl₄ caused a serious histopathological changes in the kidney. The proximal convoluted tubules showed degenerated epithelial lining with disruption of their brush borders and presence of epithelial debris inside their lumens. The renal corpuscle appeared with degeneration of the glomerulus and disrupted Bowman's capsule. The afferent arteriole appeared near the glomerulus with thickening in its wall and degeneration of endothelial lining. Extensive perivascular infiltration of inflammatory cells and massive interstitial hemorrhage were seen. These results similar to those described in the study of Adewole *et al.*, [2] found that the histopathological alterations induced by CCl₄ in rats were glomerular hypercellularity, moderate to severe necrosis and tubule-interstitial alterations. Authors suggested that the capacity for tubular absorption may have been altered, thus bringing about functional overload of nephrons with subsequent renal dysfunctions. Also, Makni *et al.*, [60] reported that the Kidney of rats injected intraperitoneally with 1ml/kg

body weight of CCl₄ showed glomerular hypertrophy and tubular dilatation.

In the current study, co-administration of Parsley with CCl₄, a dose dependent, significantly ($P < 0.01$) decreased serum urea, creatinine, uric acid, K⁺, and Cl⁻ concentrations and increased serum Na⁺ concentration and most of the proximal convoluted tubules appeared with normal epithelial lining and regained their brush borders. Some tubules still have diminished brush borders. The glomerulus appeared normal with intact Bowman's capsule, these results similar to those of the previous studies [66-69]. Shalaby and Hammoda, [66] reported that treatment of rats with 80 mg/kg body weight (i.p) gentamicin for 10 days caused a significant increase in serum urea, creatinine, sodium and potassium, marked tubular necrosis in the kidney. But, rats pre-treated orally with parsley extract (100 and 200 mg/kg) for 6 weeks and intoxicated with gentamicin (80 mg/kg) during last 10 days of the experiment showed that a significant decreases in serum urea, creatinine, sodium and potassium, and increased activity of antioxidant enzymes. Also, the extracts increased urine volume and urinary excretion of Na⁺ and K⁺ electrolytes, denoting a diuretic effect. But, the small dose (100mg/kg) of parsley leaves extract caused a mild necrosis in renal tubules with protein casts in their lumen and the large dose (200 mg/kg) of parsley extract, caused only mild congestion of intertubular blood vessels. The mechanisms of nephroprotective effect may be due to inhibition of tissue lipid peroxidation and enhancement of antioxidant activity. The nephroprotective effect of parsley was attributed to the antioxidant activity due to its high content of flavonoids [66, 70]. Authors concluded that oral pre-treatments with parsley in gentamicin-nephrotoxic rats caused nephroprotective, diuretic and antioxidant effects as they reversed serum biochemical parameters and mitigated histopathological alterations in kidney induced by gentamicin in rats. These results affirm the traditional use of parsley for the prevention of kidney diseases. Elkhamisy, [67] evaluated that the nephroprotective, diuretic, and antioxidant effects of parsley leaves extract on gentamicin induced nephrotoxic rats. The results showed that gentamicin induced nephrotoxicity manifested by alterations in serum and urine biochemical parameters, increased tissue lipid peroxidation, decreased antioxidant activity and presence of renal tubular necrosis upon histopathology. Oral pretreatments with parsley leaves extract caused a

nephroprotective effect evident by significant decreases in serum levels of urea, and creatinine in nephrotoxic rats. The extracts decreased serum sodium and potassium levels, and increased activity of antioxidant enzymes. Parsley leaves extract also increased urine volume and urinary excretion of Na⁺ and K⁺ electrolytes, denoting a diuretic activity and mitigated renal tubular necrosis induced by gentamicin.

The nephroprotective mechanisms of parsley leaves extract could be attributed to inhibition of lipid peroxidation and enhancement of antioxidant enzymes activity. Afzal *et al.*, [71] found that a polyherbal formulation containing parsley produced nephroprotective and diuretic effects in rats. Also, Mahmoud *et al.*, [69] investigated that the nephroprotective and antioxidant effects of parsley plant as watery extract of fresh leaves (decoction), fresh leaves, seeds and seeds oil against gentamicin-induced nephrotoxicity in male albino rats. The results showed that serum urea, uric acid, and creatinine were reduced significantly in rats groups received parsley products as compared to gentamicin treated group. These data suggests that parsley herb is effective in reduction of blood urea, creatinine and uric acid levels in rats with nephrotoxicity induced by gentamicin because parsley might has a diuretics effect. In addition, Khalil *et al.*, [68] confirmed that the protective effect of parsley (*Petroselinum crispum*) leaves oils against toxicity induced by carbon tetrachloride (CCl₄) in experimental rats. Results indicated that the main components in parsley oil were a-Pinene (26.6%) and Myristicin (20.3%). Kidney function tests for serum urea nitrogen, creatinine, and uric acid were found to be increased rats treated with CCl₄. Administration of 0.5 ml of parsley oil attenuated the adverse effects and biochemical alterations caused by CCl₄. Parsley oil have strong radical scavenging activity and antioxidant activity. Therefore, the results of this study show that parsley oil led to the protective effect against CCl₄ toxicity. In this study pretreatment with parsley leaves oil showed an increase in the activity of antioxidant enzymes compared to CCl₄ treated animals indicating the potentiality of parsley leaves oil to act as an antioxidant by preventing the peroxidative damage caused by CCl₄.

CONCLUSION

It can be concluded that, carbon tetrachloride has adverse effects on the kidney, it caused a serious pathophysiological changes in the kidney. Parsley aqueous extract was able to protect the kidney against these effects. So, the persons expose to

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CCl₄ should be advised to take parsley aqueous extract. Our study recommended that the consumption of parsley plant parts may be useful for patients who suffer from renal diseases.

REFERENCES

- [1] Doherty RE. (2000). A history of the production and use of Carbon Tetrachloride Tetrachloroethylene and 1,1, 1-Trichloroethane in the US: 1(1): 69-81.
- [2] Adewole SO, Salako AA, Doherty OW, and Naicker T. (2007). Effect of melatonin on carbon tetrachloride induced kidney injury in Wistar rats. *African J Biomed Res*, 10; 153-164.
- [3] DeShon ND. (1979). Carbon tetrachloride. *Kirk-Othmer Encyclopedia of Chemical Technology*, 3rd ed. Grayson M, and Eckroth D, eds. New York: John Wiley and Sons.
- [4] Sakata T., Watanabe N., Hobara N., and Nagashima H. (1987). Chronic liver injury in rats by CCl₄ inhalation. *Bull Environ Contam Toxicol*, 38: 959-961.
- [5] Ahmad F. F., Cowan D. L., Sun A. Y. (1987). Detection of free radical formation in various tissues after acute carbon tetrachloride administration in gerbil. *Life Sci*, 41: 2469-2475.
- [6] Ozturk F., Ucar M., Ozturk I. C., Vardi N., Batcioglu K. (2003): Carbon tetrachloride –induced nephrotoxicity and protective effect of betaine in Sprague-Dawley rats. *Urology* 62: 353-356.
- [7] Ruprah H, Mant TGK, and Flanagan RJ. (1985). Acute carbon tetrachloride poisoning in 19 patients: implications for diagnosis and treatment. *Lancet*, 1: 1027- 1029.
- [8] Gosselin RE, Smith RP, and Hodge HC. (1984). *Clinical toxicology of commercial products*. 5th edition. Williams and Wilkins and Wilkins, Baltimore.
- [9] Gebhardt R. (2002): Inhibition of cholesterol biosynthesis in HepG2 cells by artichoke extracts is reinforced by glucosidase pretreatment. *Phytother Res*, 16: 368-372.
- [10] Das S, Santra A, Lahiri S, Guha Mazumder DN. (2005). Implications of oxidative stress and hepatic cytokine (TNF- α and IL-6) response in the pathogenesis of hepatic collagenesis in chronic arsenic toxicity. *Toxicol Appl Pharmacol*, 204: 18-26.
- [11] Slater TF, Sawyer BC. (1971). The stimulatory effects of carbon tetrachloride and other halogenoalkanes on peroxidative reaction in rat liver fractions in vitro. Inhibitory effects of free radical scavengers and other agents. *J Biochem*, 123: 823-828.
- [12] Abraham P., Wilfred G., and Catherine S. P. (1999): Oxidative damage to the lipids and protein in the lungs, testis and kidney of rats during carbon tetrachloride intoxication. *Clin Chim Acta.*, 289: 177-179.
- [13] Awe EO, and Banjoko SO. (2013). Biochemical and haematological assessment of toxic effects of the leaf ethanol extract of *Petroselinum crispum* (Mill) Nyman ex A.W. Hill (Parsley) in rats. *BMC Complementary and Alternative Medicine* 13:75
- [14] Lopez MG, Sanchez-Mendoza IR and Ochoa-Alejo N. (1999). Comparative study of volatile components and fatty acids of plants and in-vitro cultures of parsley *Petroselinum crispum* (Mill) nym ex hill. *J Agric Food Chem*, 47: 3292–3296.
- [15] Darias V, Martin-Herrera D, Abdalla S, and Fuente D. (2001). Plant used in urinary pathologies in the Canary island. *Pharm Biol*, 39:170–180.
- [16] Kreydiyyeh SI and Usta J. (2002). Diuretic effect and mechanism of action of parsley. *J Ethnopharmacol*, 79(3): 353-359.
- [17] Parsley effectiveness, how it works and drug interaction. www.emedicine.health.com. Last assessed 07-02-2013.
- [18] Jouad H, Haloui M, Rhiouani H, El Hilaly J, and Eddouks M. (2001). Ethnobotanical survey of medicinal plants used for the treatment of diabetes, cardiac and renal diseases in the North centre region of Morocco (Fez-Boulemane). *J Ethnopharmacol*, 77:175–182.
- [19] Eddouks M, Maghrani M, Lemhadri A, Ouahidi ML, Jouad H. (2002). Ethnopharmacological survey of medicinal plants used for the treatment of diabetes mellitus, hypertension and cardiac diseases in the south-east region of Morocco (Tafilalet). *J Ethnopharmacol*, 82: 97–103.
- [20] Kreydiyyeh SI, Usta J, Kaouk I, and Al-Sadi R. (2001). The mechanism underlying the laxative properties of parsley extract. *Phytomedicine*, 8: 382–388.
- [21] Nielsen SE, Young JF, Daneshvar B, Lauridsen ST, Knuthsen P, Sandström B, Dragsted LO. (1999). Effect of parsley (*Petroselinum crispum*) intake on urinary apigenin excretion, blood antioxidant enzymes and biomarkers for oxidative stress in human subjects. *British J Nutrition*, 81(6): 447-455.
- [22] Ozsoy-Sacan O, Yanardag R, Orak H, Ozgey Y, Yarat A, and Tunali T. (2006). Effects of parsley (*Petroselinum crispum*) extract versus glibornuride on the liver of streptozotocin induced diabetic rats. *J Ethnopharmacol*, 104: 175-181.
- [23] Yazicioglu A, and Tuzlaci E. (1996). Folk medicinal plants of Trabzon (Turkey). *Fitoterapia*, 67: 307–318.
- [24] Ojala T, Remes S, Haansuu P, Vuorela H, Hiltunen R, Haahtela K, Vuorela P.(2000). Antimicrobial activity of some coumarin containing herbal plants growing in Finland. *J Ethnopharmacol*, 73: 299–305.

Nephroprotective Effect of Aqueous Extract of Parsley against Nephrotoxicity Induced By Ccl4 in the Male Rats

- [25] Wong, P.Y. and Kitts, D.D. (2006). Studies on the dual antioxidant and antibacterial properties of parsley and cilantro extracts. *Food chemistry*, 97(3): 505-515.
- [26] Maooda SN, Allam AA, Ajarem J, Abdel-Maksoud MA, Al-Basher GI, and Wang ZY. (2016). Effect of parsley (*Petroselinum crispum*, Apiaceae) juice against cadmium neurotoxicity in albino mice (*Mus Musculus*). *Behavioral and Brain Functions*, 12(1): 6
- [27] Al-Daraji HJ, Al-Mashadani HA, Al-Hassani AS, Mirza HA, Al-Hayani WK. (2012). The Influence of parsley (*Petroselinum crispum*) as feed additive on hematological traits of local Iraqi geese. *Advanc Nutrition Res*, 1(1):1-5.
- [28] Soliman HA, Eltablawy NA, and Hamed MS. (2015). The ameliorative effect of *Petroselinum crispum* (parsley) on some diabetes complications. *J Med Plants Stud*, 3(4): 92-100
- [29] Manderfeld MM, Schafer HW, Davidson PM, Zottola EA. (1997). Isolation and identification of antimicrobial furocoumarins from parsley. *J Food Prot*, 60:72-77.
- [30] Parsley and informed farmers. www.informedfarmers.com. Last assessed 01-02-2013.
- [31] Properties of Parsley. (2013). www.botanical-online.com/medicinals. Last assessed 07-02-2013.
- [32] Vora SR, Patil RB, and Pillai M. (2012). Oxidative stress associated alterations in lysosomal enzymes and modulatory effect of *Petroselinum crispum* (Mill) Nyman Ex. A.W. Hill leaf extract on mouse brain. *American-Eurasian Journal of Scientific Res*, 7(2): 64-68.
- [33] Yoshikawa M, Uemura T, Shimoda H, Kishi A, Kawahara Y, and Mastuda H (2000). Medicinal food stuff XVIII. Phytoestrogens from aerial part of *Petroselinum crispum* Mill, and structures of 6''-acetylopin and a new monoterpene glycoside. *Chem Pharm Bull*, 48: 1039-1044.
- [34] Popovic D, Kaurinovic B, Jakovljvic V, Mimica-Dukic N, Bursac M. (2007). Effect of *Petroselinum crispum* extracts on some biochemical parameters of oxidase stress in mice treated with carbon tetrachloride. *Phytothera Res*, 7: 531-537.
- [35] Zaynoun S, Abi-Ali L, Tenekjian K, and Karban A. (1985). The bergapten content of garden parsley and its significance in causing cutaneous photosensitization. *Clin Exp Dermatol*, 10: 328-331.
- [36] Zhang H, Chen F, Wang X, and Yao HY. (2006). Evaluation of antioxidant activity of parsley (*Petroselinum crispum*) essential oil and identification of its antioxidant constituents. *Food Res Int*, 39: 833-839.
- [37] Mahmood S, Hussain S, and Malik F. (2014). Critique of medicinal conspicuousness of parsley (*Petroselinum crispum*): A culinary herb of Mediterranean region. *Pak. J. Pharm. Sci*, 27(1):193-202
- [38] Fejes S, Blazovics A, Lemberkovics E, Petri G, Szoke É, Kery A. (2000). Free radical scavenging and membrane protective effects of methanol extracted fractions of parsley. *Acta Alimentaria (Budapest)*. 29(1):81-87.
- [39] Peterson S, Lampe JW, Bammler TK, Gross-Steinmeyer K and Eaton DL. (2006). Apiaceous vegetable constituents inhibit human cytochrome P-450 1A2 (hCYP1A2) activity and hCYP1A2-mediated mutagenicity of aflatoxin B1. *Food Chem. Toxicol.* 44: 1474-1484.
- [40] Potapovich AI and Kostyuk VA. (2003). Comparative study of antioxidant properties and cytoprotective activity of flavonoids. *Biochem*, 68: 514-519.
- [41] Kinoshita T, Lepp Z, Kawai Y, Terao J and Chuman H. (2006). An integrated database of flavonoids. *Biofactors*, 26: 179-188.
- [42] Lin CM, Chen CS, Chen CT, Liang YC and Lin JK. (2002). Molecular modeling of flavonoids that inhibits XO. *Biochem Biophys Res Commun*, 294: 167-172.
- [43] Hall IH, Scoville JP, Reynolds DJ, Simlot R and Duncan P. (1990). Substituted cyclic imides as potential anti-gout agents. *Life Sci*, 46: 1923-1927.
- [44] Rashwan NM. (2012). Biological study on the effect of arginine and parsley on renal toxicity in rats. *World J Med Sci*, 7 (4): 264-269.
- [45] Campanella, L., Bonanni, A., Favero, G., Tomassetti, M. (2003). Determination of antioxidant properties of aromatic herbs, olives and fresh fruit using an enzymatic sensor. *Anal Bioanal Chem*, 375(8), 1011-1016.
- [46] Hempel J, Pforte H, Raab B, Engst W, Bohm H, Jacobasch G. (1999). Flavonols and flavones of parsley cell suspension culture change the antioxidative capacity of plasma in rats. *Food/Nahrung*, 43(3):201-204.
- [47] Jassim AM. (2013). Protective Effect of *Petroselinum crispum* (parsley) extract on histopathological changes in liver, kidney and pancreas induced by sodium valproate-in male rats. *Kufa J Vet Med Sci.* 4(1): 20 - 27.
- [48] Dorman HJ, Lantto TA, Raasmaja A and Hiltunen R. (2011). Antioxidant, pro-oxidant and cytotoxic properties of parsley. *Food Func*, 2(6): 328-337.
- [49] Papay ZF, Kosa A, Boldizsar I, Ruskai A, Balogh E and Antal I. (2012). Pharmaceutical and formulation aspects of *Petroselinum crispum* extract. *Acta Pharmaceut Hung*, 82(1): 3-14.
- [50] Saeidi J, Bozorgi H, Zendehdel A, and Mehrzad J. (2012). Therapeutic effects of aqueous extracts of *Petroselinum sativum* on

Nephroprotective Effect of Aqueous Extract of Parsley against Nephrotoxicity Induced By Ccl4 in the Male Rats

- ethylene glycol-induced kidney calculi in rats. *Urol J*, 9(1): 361-366.
- [51] Miller N. J., Rice-Evans C. A. (1997): The relative contribution of ascorbic acid and phenolic antioxidants to the total antioxidant activity of orange and apple fruit juices and blackcurrant drink. *Food Chem.*, 60: 331-337.
- [52] Marsillach J, Camps J, Ferré N, Beltran R, Rul A, Mackness B, Mackness M, and Joven J. (2009). Paraoxonase-1 is related to inflammation, fibrosis and PPAR delta in experimental liver disease. *BMC Gastroenterol*, 9(3): 1-13.
- [53] El-Baz FK, Khalil WKB, Aly HF, Shoman TM, Saad SA. (2015). The Ameliorating effect of *Jatropha curcas* extract against CCl₄ induced cardiac toxicity and genotoxicity in albino rats. *Int J Pharm Sci Rev Res*, 34(2): 223-233.
- [54] Haidari F, Keshavarz SA, Shahi MM, Mahboob SA and Rashidid MR. (2011). Effects of parsley (*Petroselinum crispum*) and its flavonol constituents, kaempferol and quercetin, on serum uric acid levels, biomarkers of oxidative stress and liver xanthine oxidoreductase activity in oxonate-induced hyperuricemic rats. *Iranian J Pharm Res*, 10 (4): 811-819.
- [55] Fawcett J.K. and Scott J.E.(1960). A rapid and precise method for the determination of urea. *J. Clin. Path.*, 13: 156 -159.
- [56] Fossatti P., Prencipe L. and Berti G.(1980). Use of 3, 5-dichloro-2-hydroxybenzene-sulfonic acid/4-aminophenazone chromogenic system indirect enzymic assay of uric acid in serum and urine. *Clin. Chem.*, 26: 227- 231.
- [57] Bartels H., Bohmer M. and Heierli C.(1972). Serum creatinine determination without protein precipitation. *Clin. Chem. Acta*, 37: 193-197.
- [58] Maruna, RFL and Trinder P: *Clin Chem Acta* 1958; 2: 581.
- [59] Ross MH, Reith EJ and Romrell LJ. (1989). *Histology: A Text Atlas* (2nd ed.). Baltimore. Williams &Wilkins, pp.51-84
- [60] Makni M, Chtourou Y, Garoui EM, Boudawara T, and Fetoui H. (2012). Carbon tetrachloride-induced nephrotoxicity and DNA damage in rats Protective role of vanillin. *Human Exper Toxicol*, 31 (8): 844-852.
- [61] Aleynik S. I., Leo M. A., Ma X., Aleynik M. K., Lieber C. S. (1997): Polyenylphosphatidylcholine prevents carbon tetrachloride-induced lipid peroxidation while it attenuates liver fibrosis. *J Hepatol* 27: 554-561
- [62] Recknagel RO, Glende EA, Dolak JA, Waller RL. (1989). Mechanisms of carbon tetrachloride toxicity. *Pharmacol Ther*, 43: 139-154.
- [63] Halliwell B, and Gutteridge JMC. (2000): *Free radicals in biology and medicine*. Oxford University Press, pp. 148-149.
- [64] Garcia-Cohen EC, Marin J, Diez-Picazo LD, Baena AB, Salaices M, Rodriguez-Martinez MA (2000). Oxidative stress induced by tertbutylhydroperoxide causes vasoconstriction in the aorta from hypertensive and aged rats: role of cyclooxygenase- 2 isoform *J Pharmacol Exp Ther*, 293(1): 75-81.
- [65] Rungby J., and Ernst E. (1992). Experimentally induced lipid peroxidation after exposure to chromium, mercury or silver: interactions with carbon tetrachloride. *Pharmacol Toxicol*, 70(3): 205-207.
- [66] Shalaby MA and Hammada AA. (2014). Evaluation of nephroprotective and diuretic effects of parsley and turmeric herbs on gentamicin nephrotoxic rats. *World J Pharm Pharmaceut Sci*, 3(12): 1729-1744.
- [67] Elkhamisy AE. (2015). Protective effect of parsley leaves and turmeric roots extracts against gentamicin induced nephrotoxicity in male rats. *World J Day Food Sci*, 10 (1): 01-08.
- [68] Khalil AF, Elkhatry HO, and El Mehairy HF. (2015). Protective effect of peppermint and parsley leaves oils against hepatotoxicity on experimental rats. *Annals Agr Sci*, 60(2): 353-359.
- [69] Mahmoud NY, Salem ZA and El-Moslemany AM. (2015). Nephroprotective and antioxidant effects of parsley plant parts against gentamicin-induced nephrotoxicity in rats. *Acad J Nutr*, 4(3): 113-122.
- [70] Fejes S, Kery A, Blazovics A, Lugasi A, Lemberkovics E and Petri G. (1989). Investigation of the in vitro antioxidant effect of *Petroselinum crispum*. *Acta Pharmaceut Hung*, 68(3): 150-156.
- [71] Afzal M, Khan NA. Ghufuran A, Iqbal, A. and Inamuddin M. (2004). Diuretic and nephroprotective effect of *Jawarish zarooni sada*, a polyherbal formulation. *Ethno Pharmacol*, 91: 219-23

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