

## Zinc Immunology and Zn<sup>2+</sup> Ions Binding Anti-Bacterial Vaccine Activity for Bacterial Cell Walls against Gram-Positive and Gram-Negative Bacteria

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### Abstract

Zinc homeostasis during acute phase response is the temporal transfer of serum zinc to the tissues, causing transient serum hypozincemia, which is rebalanced during resolution of the inflammatory response. Bacteria have to avoid recognition by the host immune system in order to establish a successful infection which bacterial autolysins enable the bacteriolyses of bacterial cell walls trim cell surface PGN to prevent detection by bacterial innate immune system.

The bacterial cell walls are remodeled by PGN synthesis and PGN autolysin. *S.aureus* amidase AmiA shed light on PGN binding and cleavage that amiA distinguishes PGN mostly by the peptide, and cleavage is facilitated by a zinc-activated water molecule, developing new therapeutics against MRSA. The autolytic activity of the recombinant amidase of the Aas (autolysin/adhesin of *Staphylococcus saprophyticus*) is inhibited and is necessary for the C-terminal GW repeats, not the N-terminal repeats. AmiB catalyzes the degradation of PGN in bacteria, resulting in a marked increases of sensitivity to oxidative stress and organic acids. Amidase activity of amiC controls cell separation and PGN fragments release. In these autolysins, zinc-dependent PGN autolysin of amidases may be enhanced and induced anti-bacterial vaccine activities.

Autolysin-mediated lysis-induced bacterial cell death can contribute to the bactericidal vaccine activities. Lytic amidase autolysin LytA associates with the cell wall via its zinc-binding motif. The LytB PGN hydrolase responsible for physical separation of daughter cells cleaves the GlcNAc- $\beta$ -(1,4)-MurNAc glycosidic bond of PGN building units. LytC, LytD, and LytF are expressed in the same subpopulation of cells and complete flagellar synthesis. Major Atl autolysin also have an essential role in the early events of the fibronectin-binding proteins (FnBPs)-dependent *S.aureus* biofilm phenotype.

Human peptidoglycan recognition proteins (PGLYRPs) are novel class of recognition and effector molecules with broad Zn<sup>2+</sup>-dependent bactericidal activity against both Gram-positive and Gram-negative bacteria. It has been clear that the D-glutamate is effective for community acquired MRSA, and the other, it is efficient for *P. aeruginosa* PA14.

Adoption of Zn<sup>2+</sup> ions to the bacterial cell surface increases cell wall cohesion and favors the projection of elongated *S. aureus* surface protein (SasG) away from the cell surface. Zinc is an essential nutrient for microbial growth, but can be toxic in excess. Zinc importer adcABC of the primary group A streptococcus (GAS) zinc uptake system is composed of a cell surface-exposed zinc-binding protein (adcA), an inner membrane permease (AdcB), and a cytosolic ATPase (AdcC) that provides the energy for zinc import by ATP hydrolysis.

Enterotoxigenic *E.coli* (ETEC) is the most common bacterial cause of children's diarrhea, in which antigen and antitoxin antibodies that neutralized both toxins that are associated with all cases of ETEC diarrhea, and polypeptide or subunit vaccines have the potential to effectively protect against ETEC diarrhea. Oral vaccines which are intended for global use do not necessarily induce the same immune responses in all children worldwide. Zinc has positive effect in children with complication of diarrhea that young children are immunized

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with oral inactivated whole cell cholera vaccine containing recombinant cholera toxin B subunit, in which the combination of zinc with cholera vaccine and oral rehydration solutions (ORSs) has a positive impact on cholera and diarrhea. Acute diarrhea remains a leading cause of childhood death despite the undeniable success of oral rehydration therapy (ORT) that vaccination is the most effective method of preventing infectious diseases.

Zinc uptake A (ZnuA) is a high affinity acquisition of Zn<sup>2+</sup> in *E. coli* was demonstrated and shown to occur via the ATP-binding cassette (ABC) permease, ZnuABC that the Znu permease comprises the solute-binding protein (SBP) ZnuA, and an ABC transporter. The acquisition of zinc by *P. aeruginosa* PAO<sub>1</sub> reveals a hitherto unrecognized complexity in zinc homeostasis that enables the bacterium to survive under zinc limitation that the mechanisms and pathways utilized by *P. aeruginosa* to survive and promulgate in environments of varying Zn<sup>2+</sup> abundance, with the findings widely applicable to other prokaryotic organisms. Recombinant flagella and pili targeting lipo-polysaccharides and O-antigens have shown some promise in preventing infection that outer membrane protein including OprF and OprI are newer representative of vaccine candidates which many of the aforementioned vaccine act on a single target, thus lacking a broad range of protection. Recombinant AfeA expresses abundant epitopes on the bacterial surface and induces protective responses in the mouse pulmonary clearance model following aerosol challenge with *Moraxella catarrhalis*.

Multivalent fusion DNA vaccine against *Brucella abortus* has been constructed that the expression of BAB antigens conjugated to SOD protein can polarize mice immunity to a Th1-type phenotype.

Zinc oxide (ZnO) nanoparticles (ZnO-NPs) are attractive antibacterial properties due to increased specific surface area as the reduced particle size leading to enhanced particle surface reactivity. Bacteriolytic activity of ZnO-NPs is associated with the generation of reactive oxygen species (ROS) including hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), hydroxyl radicals (OH<sup>•</sup>), and peroxide (O<sub>2</sub><sup>-2</sup>) that ROS have been cell wall damage due to ZnO-localized interaction, enhanced membrane permeability, internalization of Nps due to loss of proton motive force and uptake of toxic dissolved zinc ions. Released zinc ions from zinc oxide penetrate the bacterial cell wall via diffusion that ZnO-NPs disintegrate the cell membrane and accumulate in the cytoplasm where they interact with biomolecules causing cell apoptosis leading to cell death. ZnO-NPs caused significant up-regulation of biosynthesis and degradation. The antibacterial mechanism of ZnO-NPs is likely due to disruption of the cell membrane and oxidative stress such as *Campylobacter*.

Accordingly, Zn<sup>2+</sup> ions under the homeostasis region could be appreciable for anti-bacterial vaccine development.

**Keywords:** Zinc homeostasis, Zn<sup>2+</sup>-binding vaccine, PGN hydrolase and autolysin, Amidase, ZnO-NPs, ROS.

### ABBREVIATIONS

**Aas**=autolysin/adhesion of *Staphylococcus saprophyticus*, **ABC**=ATP-binding cassette, **APC**=antigen presenting cell, **A. stephensi**=*Anopheles stephensi*, **B.abortus**=*Brucella abortus*, *B. subtilis*=*Bacillus subtilis*, **CBPs**=choline binding proteins, **C. difficile**=*Clostridium difficile*, **E. coli**=*Escherichia coli*, **E. faecalis**=*Enterococcus faecalis*, **E. faecium**=*Enterococcus faecium*, **ETEC**=*Enterotoxigenic E.coli*, **Eps**=Zinc dependent endopeptidases, **FnBPs**=fibronectin-binding proteins, **Gas**=group A streptococcus, **GeIE**=zinc metalloprotease, gelatinase, **M.catarrhalis**=*Moraxella catarrhalis*, **MCPs**=Metallo carboxy peptidases, **MIBRs**=most probable immunoprotective B-cell epitope regions,

**MRB**=multidrug bacteria, **MRSA**=methicillin-resistant *Staphylococcus aureus*, **ORSs**=oral rehydration solutions, **ORT**=oral rehydration therapy, **P. aeruginosa**=*Pseudomonas aeruginosa*, **PBP2a**=penicillin-binding protein2a, **PGN**=peptidoglycan, **PGRPs**=peptidoglycan recognition proteins, **PSP**=plasmid stabilization protein, **ROS**=reactive oxygen species, **Sags**=superantigens, **SasG**=*S. aureus* surface protein, **S. aureus**=*Staphylococcus aureus*, **SBP**=solute-binding protein, **SEB**=staphylococcal enterotoxin serotype B, **SOD**=superoxide dismutase, **S. pneumoniae**=*Streptococcus pneumoniae*, **TBVs**=transmission-blocking vaccines, **VRE**=vancomycin-resistant *Enterococcus faecium*, **ZnO-NPs**=Zinc oxide (ZnO) nanoparticles, **ZBL**=zinc binding lipoprotein, **ZnuA**=Zinc uptake A.

### INTRODUCTION

Zinc plays an important role in human immunity that zinc deficiency in the host is linked to increased susceptibility to bacterial infection, zinc homeostasis must be essential to be maintained for bacteria, and zinc excess is highly toxic toward microorganism in an anti-bacterial role for zinc in innate immune defense against infected diseases [1]. Zinc deficiency causes severe impairment of immune function, comprising the adaptive as well as the innate immune system, adequate zinc homeostasis is essential for a well-functioning immune system, and high zinc excess provokes an impairment of the immune system comparable to zinc deficiency [2].

Zinc is the second most abundant trace metal with human body 2-3 g, 90% in muscle and bone, and 10% other organs include prostate, liver, the gastrointestinal tract, kidney, skin, lung brain, heart, and pancreas in humans which cellular zinc underlies an efficient homeostatic control that avoids accumulation of zinc in excess [3]. The role of zinc in cell death has apoptosis that the influence of zinc on apoptosis is tissue/cell type, zinc concentration, and expression of zinc transporters and zinc-binding proteins [3]. Host zinc homeostasis changes in response to bacterial infections, including production of metal sequestering proteins and bombardment of bacteria with toxic level of zinc at host-pathogen interface [4]. Apoptosis is defined as cell death activated by an internally controlled suicide program that bacteria are able to trigger apoptosis, including the secretion of compounds such as protein synthesis inhibitions, pore forming proteins, molecules responsible for the activation of the endogenous death in the infected cell, and super antigens [5]. Regulation of apoptosis is essential for normal embryonic development and for homeostasis in adult tissue.

Zinc has a rather low toxicity and influences apoptosis by acting on several molecular regulators of programmed cell death which can inhibit apoptosis there by either prolonging the survival of infected cells such that the production of progeny virus is maximized or facilitating the establishment of virus persistence. The influence of zinc on apoptosis is very complex that variables in this complex network are tissue and cell type, zinc concentration, expression of zinc transporters and zinc-binding proteins, oxidative or nitrosative stress, and the improvement of molecular

opposing functions. Zinc-dependent antibacterial vaccine principle has been not completely

understood, but novel research as targets for anti bacterial vaccines and therapies has been proceeding [6,7].

Zinc ion killing occurs chiefly by bacteriolyses of bacterial cell walls due to activated peptidoglycan (PGN) autolysins such as amidases, endopeptidases, and carboxypeptidase against bacteria [8]. These PGN autolysins induced anti-bacterial vaccine activity may be enhanced by activation of zinc dependent PGN autolysins. PGN autolysins are bacterial PGN degrading enzymes that these muropeptides can be produced or modified by the activity of bacterial glycolytic and peptidolytic enzymes referred to as PGN hydrolases and autolysins which specific bacterial pathogens use PGN degradation to subvert host innate immunity [9].

In this review, Zn<sup>2+</sup> ions binding anti-bacterial vaccine activity for PGN autolysins induced vaccine, zinc-dependent bacterial vaccine, and ZnO nanoparticle induced anti-bacterial vaccine are discussed, and then, the zinc dependent molecular vaccine mechanisms are clarified.

### ZINC IMMUNITY IN INFECTION

The innate immune system represents the defense first line against a pathogen before the adaptive system can develop the appropriate response. Many organs are affected by zinc deficiency, especially the immune system that is markedly susceptible to changes of zinc levels which the immune response involves in the regulation of the innate and adaptive immunity, and this zinc homeostasis is critical for sustaining proper immune function [10]. Thus, inflammation is a natural process required to protect the host from tissue damage and infections, which leads to the resolution of the inflammatory response and the restoration of homeostasis. Despite zinc deficiency can be treated by proper zinc intake, suboptimal zinc status cannot simply diagnosed by reason of the lack of clinical signs and reliable biochemical indicators of zinc status. High zinc concentration is that zinc binding to proteins can activate or inactivate their activity, or change characteristics important for substrate binding, while, zinc homeostasis is primarily controlled via the expression and action of 14 zinc transporters that decreasing cytoplasmic zinc can describe export via

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ZnTs, but also the transport of zinc into one of those organelles [11]. Zinc homeostasis during acute phase response is the temporal transfer of serum zinc to the tissues, causing transient serum hypozincemia, which is rebalanced during resolution of the inflammatory response that intracellularly increased zinc can intoxicate engulfed pathogens and acts cytoprotective by promotion of neutralizing reactive oxygen species (ROS) and nitrogen species (RNS) [11]. Bacteria have to avoid recognition by the host immune system in order to establish a successful infection which bacterial autolysins enable the bacteriolyses of bacterial cell walls trim cell surface PGN to prevent detection by bacterial innate immune system [12].

### ZINC-BINDING PGN AUTOLYSIN ENHANCED ANTI-BACTERIAL VACCINE ACTIVITY

Bacterial peptidoglycan (PGN) structure of both Gram-positive and Gram-negative bacteria comprises repeating disaccharide backbones of N-acetylglucosamine (NAG) and  $\beta$ -(1-4)-N-acetylmuramic acid (NAM) that are crosslinked by peptide stem chains attached to the NAM residues [13]. The action sites of bacterial autolysins are comprised that for *S.aureus* PGN layer cell wall, there are N-acetylmuramidase-L-alanine amidase and DD-endopeptidase, and the other, for *E. coli* cell wall, there are endopeptidase of degrading enzyme at lipoprotein of C- and N-terminals, and also amidase, peptidase, and carboxypeptidase at thin PGN layer in periplasmic space [14]. The bacterial cell walls are a strong flexible meshwork of PGN that gives a bacterium structural integrity, in which to accommodate a growing cell, the walls are remodeled by PGN synthesis and PGN autolysin. In these autolysins, zinc-dependent PGN autolysin of amidases may be enhanced and induced anti-bacterial vaccine activities.

PGN is the main constituent of bacterial cell walls and must be continuously synthesized and degraded to maintain the integrity and viability of the cells that bacterial cell wall hydrolases of amidase, glycosidase, and peptidase display a modular architecture combining multiple and different catalytic domains, including some lytic transglycosylases as well as cell wall binding domains [15].

*S.aureus* amidase AmiA shed light on PGN binding and cleavage that amiA distinguishes PGN mostly by the peptide, and cleavage is facilitated by a zinc-activated water molecule, in order to develop new therapeutics against MRSA [16].

The autolytic activity of the recombinant amidase of the Aas (autolysin/adhesin of *Staphylococcus saprophyticus*) is inhibited and is necessary for the C-terminal GW repeats, not the N-terminal repeats [17]. Amidase gene (AmiB) catalyzes the degradation of PGN in bacteria that the amiB gene was composed of 1,722 nucleotides and 573 amino acid which is involved in the separation of daughter cells after cell division and inactivation of the amiB gene, resulting in a marked increase of sensitivity to oxidative stress and organic acids [18]. Amidase activity of amiC controls cell separation and PGN fragments release [19].

Autolysin-mediated lysis-induced bacterial cell death can contribute to the bactericidal vaccine activities. Lytic amidase autolysin LytA which is released by bacterial lysis, associates with the cell wall via its zinc-binding motif that the amidase domain comprises a complex substrate-binding crevice and needs to interact with a large-motif epitope of PGN for catalysis [20]. Suicidal amidase autolysin LytA having both autolysis and capsule shedding depends on the cell wall hydrolytic activity of LytA that capsule shedding drastically increases invasion of epithelial cells and is the main pathway by which pneumococci reduce surface bound capsule during early acute lung infection of mice [21]. In the biofilms increase as zinc concentrations increase and biofilm formation effect as a negative regulator of LytA dependent autolysis, zinc availability contributes to the ability of pneumococci to form aggregates and subsequently, biofilms [22]. The LytB PGN hydrolase responsible for physical separation of daughter cells cleaves the GlcNAc- $\beta$ -(1,4)-MurNAc glycosidic bond of PGN building units that cell wall digestion products and solubilisation rates might indicate a tight control of LytB activity to prevent unrestrained breakdown of the cell wall [23]. The PGN-remodeling autolysins LytC, LytD, and LytF are expressed in the same subpopulation of cells and complete flagellar synthesis that LytC appears to be important for flagellar function, motility was restored to a LytC mutant by mutation of either lon A, and LytC, LytD, and LytF autolysins to population heterogeneity in *B. subtilis* [24].

Atl is the major autolysin in *S aureus* that the bifunctional major autolysin play a key role in staphylococcal cell separation which processing of Atl yield catalytically active amidase and glucosamidase

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domains [25]. The biochemical and structural staphylococcal Atl have successful cloning, high level over-expression, and purification Atl proteins [26]. Major Atl autolysin also have an essential role in the early events of the fibronectin-binding proteins (FnBPs)-dependent *S.aureus* biofilm phenotype [27].

For the contribution of autolysins of PGN hydrolases to bacterial killing, there are N-acetylglucosaminidase (AtlA), two N-acetyl-muraminases (AtlB and AtlC) [28]. AtlA is the major PGN hydrolases of *Enterococcus faecalis* involved in cell division and cellular autolysis and the zinc metalloprotease, gelatinase (GelE) of their interplay proposed to regulate AtlA function, which N-terminal cleavage was required for efficient AtlA-mediated cell division, and AtlA septum localization and subsequent cell separation can be modulated by a single GelE-mediated N-terminal cleavage event [29].

Zinc-dependent endopeptidases (Eps) are predicted to hydrolyze PGN to facilitate cell growth that zinc availability affects strong activity of cell wall hydrolases, and zinc-regulated endopeptidases are present in divergent Gram-negative bacteria [30]. Fusion protein consisting of most probable immunoprotective B-cell epitope regions (MIBRs) are both plasmid stabilization protein (PSP) and zinc binding lipoprotein (ZBL), PSP and ZBL respectively (APZs), in which the autolysin MIBRs show the highest probability for eliciting immunoprotection and pneumococcal conjugate vaccine against *Streptococcus pneumoniae* [31].

Carboxypeptidases are exopeptidases that remove a single amino acid residue from the C terminus of proteins or peptides that the carboxy-peptidase B1 of and its evaluation have been high molecular characterization for transmission-blocking vaccines (TBVs) against Malaria eradication [32]. Metallo-carboxypeptidases (MCPs) of the M32 family of peptidases exhibit a significant hydrolytic activity and different hydrolysis patterns against *Trypanosoma brucei* or *cruzi* [33]. Thus, zinc-dependent carboxypeptidase autolysin could adapt to be appreciable the anti-bacterial vaccines. Furthermore, it is worth noting as a novel recombinant vaccine candidate comprising penicillin-binding protein 2a (PBP2a) and r-autolysin that active vaccination with a mixture of r-PBP2a/r-autolysin and conjugate form vaccine reduced the mortality rate and protected mice against lethal MRSA [34].

## Zn<sup>2+</sup> IONS-DEPENDENT BACTERIAL CELL WALLS INDUCED ANTI-BACTERIAL VACCINE ACTIVITY

### Zn<sup>2+</sup> Ions Binding Vaccine Activity for Thick PGN Cell Wall Against Gram-Positive Bacteria

Antibody and vaccine development against *S.aureus* that produces cell envelope-associated proteins, secreted toxin, host cell lysis antibody function interference, are physiologically and pathologically considered that staphylococcal enterotoxin serotype B (SEB) and super antigenicity of superantigens (Sags) are largely achieved by the activated (APCs) and T cells, leading to a massive release of cytokines [35]. Human peptidoglycan recognition proteins (PGLYRPs) are novel class of recognition and effector molecules with broad Zn<sup>2+</sup>-dependent bactericidal activity against both Gram-positive and Gram-negative bacteria [36]. In order to generate effective bacterial whole-cell vaccines auxotrophic for D-glutamate, it has been clear that the D-glutamate is effective for community acquired MRSA, and the other, it is efficient for *P. aeruginosa* PA14 [37].

*Clostridium difficile* Residues are important in zinc binding and enzymatic activity that CD630 28300 (named Zmp1) destabilizes the fibronectin network produced by human fibroblast which a novel extracellular zinc metalloprotease may be important in key steps of clostridial pathogenesis [38]. Mice were immunized with the antibodies raised against recombinant lipoproteins, showing significant reduction of colony counts in mice livers and demonstrating the efficacy of these metal binding lipoproteins as promising vaccine candidates [39]. Zinc supplementation promotes the induction of T cell immunity to control infection and ameliorate immunopathology against Gram-positive pneumonia in children [40]. Adsorption of Zn<sup>2+</sup> ions to the bacterial cell surface increases cell wall cohesion and favors the projection of elongated *S. aureus* surface protein (SasG) away from the cell surface, thereby enabling zinc-dependent homophilic bonds between opposing cells that zinc-dependent cell surface dynamics may represent a general mechanism for activating adhesion in biofilm-forming species [41]. Zinc is an essential nutrient for microbial growth, but can be toxic in excess. zinc importer adcABC of the primary group A streptococcus (GAS) zinc uptake system is composed of a cell surface-exposed zinc-binding protein (adcA), an inner membrane permease (AdcB), and a cytosolic

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ATPase (AdcC) that provides the energy for zinc import by ATP hydrolysis [42]. Immunization of mice with the extracellular component of the zinc importer confers protection against system GAS, and a similar struggle for zinc may occur during streptococcal infections [42]. Extracellular vesicles (EVs) are immuno-genic in mice, elicit cytolysin-neutralizing antibodies, and protect the animals in a lethal sepsis model that mechanisms underlie *S.aureus* EV production and

highlights the usefulness of Evs as a *S.aureus* vaccine platform [43]. Pneumococcal choline binding proteins (CBPs) include cell wall hydrolases and play a dual role for the development of novel antipneumococcal drugs, both as targets for inhibitors of binding to the cell wall and as active cell lytic agents [44].

**Table 1** represents vaccine activities of Zn<sup>2+</sup> ions for thick PGN layer cell wall against Gram-positive bacteria.

**Table 1.** Zn<sup>2+</sup> ions binding vaccine activity for Gram-positive PGN layer cell wall

Zn <sup>2+</sup> Ions	Gram-Positive Bacterial Thick PGN Cell Wall
Zn <sup>2+</sup> ions	<p><b>Zinc-binding PGN autolysins, Zn<sup>2+</sup>-dependent bactericide</b></p> <ul style="list-style-type: none"> <li><b>Zinc Ions-Dependent Autolysins</b></li> <li><i>S.aureus</i>amidase <i>AmiA</i></li> <li>Recombinant amidase of the <i>Aas</i></li> <li>Lytic amidase <i>LytA</i> for <i>Streptococcus pneumoniae</i></li> <li>Pneumococcal autolysin <i>LytA</i></li> <li><i>LytC, D, F</i> of PGN remodeling for <i>Bacillus subtilis</i></li> <li>Endopeptidase <i>LytF</i> for <i>Bacillus subtilis</i></li> <li><i>AtlA</i> autolysin for <i>GeLE</i> against <i>E. faecalis</i></li> <li><i>AtlA, AtlB, AtlC</i> autolysins against <i>enterococcus faecalis</i></li> <li>Fusion protein autolysin, MIBRs against <i>S. pneumoniae</i></li> <li>Carboxypeptidase B1 against <i>Anopheles stephensi</i> and for malaria as transmission-blocking vaccines</li> <li>Metallo-carboxypeptidase M32 against <i>Trypanosoma brucei</i> or <i>cruzi</i></li> <li>PBP2a and autolysin mixture against MRSA</li> </ul> <ul style="list-style-type: none"> <li>MDR of Gram-positive strain as antibody and vaccine</li> <li>Human PGLYRPs against both Gram-positive and Gram-negative bacteria</li> <li>D-glutamate auxotrophy against MRSA</li> <li>Extracellular zinc metalloprotease against <i>Clostridium difficile</i></li> <li>Zinc binding lipoprotein against <i>Enterococcal</i> infections</li> <li>Zinc supplementation for <i>pneumonia</i> in children</li> <li>Zn<sup>2+</sup>-dependent <i>S.aureus</i> surface protein (<i>SasG</i>) formation</li> <li>Zinc importer <i>AdcABC</i> for streptococcal infections against GAS</li> <li>Pneumococcal CBP cell wall hydrolases</li> </ul> <ul style="list-style-type: none"> <li>ZnO-NPs have a very high anti-bacterial activity and ROS generation against MRSA (ROS; H<sub>2</sub>O<sub>2</sub>, OH<sup>-</sup>, O<sub>2</sub><sup>-2</sup>)</li> <li>ZnO-NPs caused up-regulation of pyrimidine biosynthesis and degradation against MRSA</li> </ul>

### Zn<sup>2+</sup> Ions Binding Vaccine Activity for Gram-Negative Bacterial Cell Wall Having Outer Membrane Lipoprotein and PGN Thin Layer in Periplasm Space

Antibody and vaccine activity against *E. coli* have been clarified that enterotoxigenic *E.coli* (ETEC) is the most

common bacterial cause of children's diarrhea, in which antigen preparation induced antitoxin antibodies that neutralized both toxins that are associated with all cases of ETEC diarrhea, and polypeptide or subunit vaccines have the potential to effectively protect against ETEC diarrhea [45]. Oral vaccines which are

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intended for global use do not necessarily induce the same immune responses in all children world wide that vaccine designed for oral administration will need to be adjusted to these potential problems in order to maximize benefits for all children [46]. Zinc has positive effect in children with complication of diarrhea that young children are immunized with oral inactivated whole cell cholera vaccine containing recombinant cholera toxin B subunit, in which the combination of zinc with cholera vaccine and oral rehydration solutions (ORSs) has a positive impact on cholera and diarrhea [47]. Acute diarrhea remains a leading cause of childhood death despite the undeniable success of oral rehydration therapy (ORT) that Vaccination is the most effective method of preventing infectious diseases [48]. There may be an influence of zinc on cholera vaccination and a suppression of antibody formation against cholera toxin.

Zinc uptake A (ZnuA) is a high affinity acquisition of Zn<sup>2+</sup> in *E. coli* was shown to occur via the ATP-binding cassette (ABC) permease and ZnuABC that the Znu permease comprises the solute-binding protein (SBP) ZnuA, and an ABC transporter. The acquisition of zinc by *P. aeruginosa* PAO<sub>1</sub> reveals a hitherto unrecognized complexity in zinc homeostasis that enables the bacterium to survive under zinc limitation that the mechanisms and pathways utilized by *P. aeruginosa*

to survive and promulgate in environments of varying Zn<sup>2+</sup> abundance, with the findings widely applicable to other prokaryotic organisms [49]. Recombinant flagella and pili to targeting lipo-polysaccharides and O-antigens have shown some promise in an preventing infection that outer membrane protein including OprF and OprI are newer representative of vaccine candidates which many of the aforementioned vaccine act on a single target, thus lacking a broad range of protection [50]. Recombinant AfeA expresses abundant epitopes on the bacterial surface and induces protective responses in the mouse pulmonary clearance model following aerosol challenge with *Moraxella catarrhalis*, in which AfeA is an excellent vaccine antigen to be included in a vaccine to prevent infections caused by *M. catarrhalis* [51]. Multivalent fusion DNA vaccine against *Brucella abortus* has been constructed that the expression of BAB antigens, encoded in *B. abortus* BAB1 0279 open reading frame (ORF) genomic island 3 (GI-3) and conjugated to SOD protein can polarize mice immunity to a Th1-type phenotype, conferring low levels of protection in animal model [52].

**Table 2** exhibits Zn<sup>2+</sup> ions binding vaccine activities for Gram-negative cell wall with outer membrane lipoprotein at C- and N-terminals and thin PGN layer in plasmic space.

**Table 2.** Zn<sup>2+</sup> ions binding vaccine activity of Zn<sup>2+</sup> ions for Gram-negative cell wall with outer membrane lipoprotein and PGN layer in periplasmic space

Zn <sup>2+</sup> Ions	Gram-Negative Bacterial Cell Wall
Zn <sup>2+</sup> ions	<ul style="list-style-type: none"> <li>Membrane Lipoprotein at C- and N-terminals</li> <li>Zn<sup>2+</sup> binding autolysins, Zn<sup>2+</sup> ions</li> <li>Amidase gene <i>amiB</i>/<i>LysM</i></li> <li>Endopeptidase regulation</li> <li>EPEC subunit vaccine</li> <li>Oral vaccine by ORT</li> <li>ZnuB against <i>P. aeruginosa</i>,</li> <li>Preventive vaccine by recombinant flagella against <i>P. aeruginosa</i></li> <li>AfeA excellent vaccine antigen preventing infection of <i>M. catarrhalis</i></li> <li>Fusion DNA vaccine against <i>Brucella abortus</i></li> <li>ZnO-NPs disrupt the cell membrane and oxidative stress against <i>Campylobacter</i></li> </ul>
	<ul style="list-style-type: none"> <li>PGN Thin Layer in Periplasmic Space</li> <li>Zn<sup>2+</sup> binding autolysin, Zn<sup>2+</sup> ions</li> <li>AmiC in PGN fragment release</li> <li>Carboxypeptidase by transmission blocking vaccines</li> <li>PGRPs or PGLYRPs</li> <li>D-glutamate auxotrophy against <i>P. aeruginosa</i> PA14</li> <li>ORT in infectious diarrhoea Combination of zinc and cholera vaccine and ORS</li> <li>ZnuA against <i>P. aeruginosa</i></li> <li>Recombinant flagella and pili against <i>P. aeruginosa</i></li> </ul>

## **ZNO NANOPARTICLES-DEPENDENT ANTI-BACTERIAL VACCINE**

Zinc oxide (ZnO) nanoparticles (ZnO-NPs) are attractive antibacterial properties with broad-spectrum antibiotics due to increased specific surface area as the reduced particle size leading to enhanced particle surface reactivity. Bactericidal and bacteriostatic activity of ZnO-NPs is associated with the generation of reactive oxygen species (ROS) including hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), hydroxyl radicals (OH<sup>•</sup>), and peroxide (O<sub>2</sub><sup>-2</sup>) that ROS have been a major factor for several mechanisms including cell wall damage due to ZnO-localized interaction, enhanced membrane permeability, internalization of Nps due to loss of proton motive force and uptake of toxic dissolved zinc ions [53]. Zinc oxide is an essential ingredient of many enzymes, sun screens, and ointments for pain and itch relief that released zinc ions from zinc oxide penetrate the bacterial cell wall via diffusion that ZnO-NPs disintegrate the cell membrane and accumulate in the cytoplasm where they interact with biomolecules causing cell apoptosis leading to cell death [54]. ZnO-NPs against MRSA are that exposure to ZnO-NPs resulted in over three-log reduction in colonies of MRSA with minimal increase in ROS or lipid peroxidation which ZnO-NPs caused significant up-regulation of pyrimidine biosynthesis and carbohydrate degradation [55]. The antibacterial mechanism of ZnO-NPs is likely due to disruption of the cell membrane and oxidative stress in *Campylobacter*.

## **CONCLUSIONS**

Zinc homeostasis during acute phase response is the temporal transfer of serum zinc to the tissues, causing transient serum hypozincemia, which is rebalanced during resolution of the inflammatory response. Bacteria have to avoid recognition by the host immune system in order to establish a successful infection which bacterial autolysins enable the bacteriolyses of bacterial cell walls trim cell surface PGN to prevent detection by bacterial innate immune system.

The bacterial cell walls are remodeled by PGN synthesis and PGN autolysin. In these autolysins, zinc-dependent PGN autolysin of amidases may be enhanced and induced anti-bacterial vaccine activities. *S.aureus* amidase AmiA shed light on PGN binding and cleavage that amiA distinguishes PGN mostly by the peptide, and cleavage is facilitated by a zinc-activated water molecule, developing new therapeutics against MRSA.

The autolytic activity of the recombinant amidase of the Aas is inhibited and is necessary for the C-terminal GW repeats, not the N-terminal repeats. Amidase gene (AmiB) catalyzes the degradation of PGN in bacteria. Amidase activity of amiC controls cell separation and PGN fragments release.

Autolysin-mediated lysis-induced bacterial cell death can contribute to the bactericidal vaccine activities. Lytic amidase autolysin LytA which is released by bacterial lysis, associates with the cell wall via its zinc-binding motif that the amidase domain comprises a complex substrate-binding crevice and needs to interact with a large-motif epitope of PGN for catalysis. The LytB PGN hydrolase responsible for physical separation of daughter cells cleaves the GlcNAc-β-(1,4)-MurNAc glycosidic bond of PGN building units. The PGN-remodeling autolysins LytC, LytD, and LytF are expressed in the same subpopulation of cells and complete flagellar synthesis. Major Atl autolysin also have an essential role in the early events of the FnBPs-dependent *S.aureus* biofilm phenotype.

Human PGLYRPs are novel class of recognition and effector molecules with broad Zn<sup>2+</sup>-dependent bactericidal activity against both Gram-positive and Gram-negative bacteria. The D-glutamate is effective for community acquired MRSA, and the other, it is efficient for *P. aeruginosa* PA14.

Adoption of Zn<sup>2+</sup> ions to the bacterial cell surface increases cell wall cohesion and favors the projection of elongated SasG away from the cell surface, thereby enabling zinc-dependent homophilic bonds between opposing cells that zinc-dependent cell surface dynamics may represent a general mechanism for activating adhesion in biofilm-forming species. Zinc is an essential nutrient for microbial growth, but can be toxic in excess. zinc importer adcABC of the primary GAS zinc uptake system is composed of a cell surface-exposed zinc-binding protein (adcA), an inner membrane permease (AdcB), and a cytosolic ATPase (AdcC) that provides the energy for zinc import by ATP hydrolysis. EVs are immuno-genic in mice, elicit cytolytic-neutralizing antibodies, and protect the animals in a lethal sepsis model that mechanisms underlie *S.aureus* EV production and highlights the usefulness of Evs as a *S.aureus* vaccine platform.

EPEC is the most common bacterial cause of children's diarrhea, in which antigen preparation induced antitoxin antibodies that neutralized both toxins that



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are associated with all cases of ETEC diarrhea, and polypeptide or subunit vaccines have the potential to effectively protect against ETEC diarrhea. Oral vaccines which are intended for global use do not necessarily induce the same immune responses in all children worldwide that vaccine designed for oral administration will need to be adjusted to these potential problems in order to maximize benefits for all children. Zinc has positive effect in children with complication of diarrhea that young children are immunized with oral inactivated whole cell cholera vaccine containing recombinant cholera toxin B subunit, in which the combination of zinc with cholera vaccine and ORSs has a positive impact on cholera and diarrhea. Acute diarrhea remains a leading cause of childhood death despite the undeniable success of ORT that vaccination is the most effective method of preventing infectious diseases. There may be an influence of zinc on cholera vaccination and a suppression of antibody formation against cholera toxin.

ZnuA is a high affinity acquisition of Zn<sup>2+</sup> in *E. coli* was shown to occur via the ABC permease that the Znu permease comprises the SBP ZnuA, and an ABC transporter. The acquisition of zinc by *P. aeruginosa* PAO<sub>1</sub> reveals a hitherto unrecognized complexity in zinc homeostasis that enables the bacterium to survive under zinc limitation that the mechanisms and pathways utilized by *P. aeruginosa* to survive and promulgate in environments of varying Zn<sup>2+</sup> abundance, with the findings widely applicable to other prokaryotic organisms. Recombinant flagella and pili to targeting lipo-polysaccharides and O-antigens have shown some promise in preventing infection that outer membrane protein including OprF and OprI are newer representative of vaccine candidates which many of the aforementioned vaccine act on a single target, thus lacking a broad range of protection [50]. Recombinant AfeA expresses abundant epitopes on the bacterial surface and induces protective responses in the mouse pulmonary clearance model following aerosol challenge with *Moraxella catarrhalis*, in which AfeA is an excellent vaccine antigen to be included in a vaccine to prevent infections caused by *M. catarrhalis*. Multivalent fusion DNA vaccine against *Brucella abortus* has been constructed that the expression of BAB antigens conjugated to superoxide dismutase (SOD) protein can polarize mice immunity to a Th1-type phenotype, conferring low levels of protection in animal model.

ZnO-NPs are attractive antibacterial properties with broad-spectrum antibiotics due to increased specific surface area as the reduced particle size leading to enhanced particle surface reactivity. Bactericidal and bacteriostatic activity of ZnO-NPs is associated with the generation of reactive oxygen species (ROS) including hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), hydroxyl radicals (OH<sup>•</sup>), and peroxide (O<sub>2</sub><sup>•2</sup>) that ROS have been a major factor for several mechanisms including cell wall damage due to ZnO-localized interaction, enhanced membrane permeability, internalization of NPs due to loss of proton motive force and uptake of toxic dissolved zinc ions.

Zinc oxide is an essential ingredient of many enzymes, sun screens, and ointments for pain and itch relief that released zinc ions from zinc oxide penetrate the bacterial cell wall via diffusion that ZnO-NPs disintegrate the cell membrane and accumulate in the cytoplasm where they interact with biomolecules causing cell apoptosis leading to cell death. ZnO-NPs caused significant up-regulation of pyrimidine biosynthesis and carbohydrate degradation. The antibacterial mechanism of ZnO-NPs is likely due to disruption of the cell membrane and oxidative stress in *Campylobacter*.

## REFERENCES

- [1] C-I. Y. Ong, C. M. Gillen, T. C. Barnett et al; An antimicrobial role for zinc in innate immune defense against group A *Streptococcus*. The Journal of Infectious Diseases. 2014; 209: 1500-1508.
- [2] I. Wessels, M. Maywald and L. Rink; Zinc as a gatekeeper of immune function, *Nutrients*, 2017; 9: 1-44.
- [3] L. M. Pium, L. Rink and Hajo Kaase (2010); The essential toxin: Impact of zinc on human health. *International Journal of environmental research and public health*. 2017; 7: 1342-1365.
- [4] L. D. Palmer and E. Skaar; Transition metals and virulence in bacteria, *Annu Rev Genet*, 2016; 50, No 23: 67-94.
- [5] M. Lancellotti, R. F. C. Pereira, G. G. Cury and L. M. de Hollanda; Pathogenic and opportunistic respiratory bacteria-induced apoptosis. *The Brazilian Journal of Infectious Diseases*, 2009; 13 (3): 226-231.

## Zinc Immunology and Zn<sup>2+</sup> Ions Binding Anti-Bacterial Vaccine Activity for Bacterial Cell Walls against Gram-Positive and Gram-Negative Bacteria

- [6] H. S. Garmory and R. W. Titball; ATP-binding cassette transporters are targets for the development of antibacterial vaccines and therapies, *Infection and Immunity*, 2004; 72 (12): 6757-6763.
- [7] A. M. Downey, P. Kaplonek, and P. H. Seeberger; MAIT cells as attractive vaccine targets, *FEBS Letters*, PRESS, 2019; 593: 1627-1640.
- [8] T. Ishida; Antibacterial mechanism of bacteriolyses of bacterial infection cell walls by zinc ion induced activations of PGN autolysins, and DNA damages, *Journal of Genes and Proteins*, 2017; 1: 1-7.
- [9] J. Humann and L. J. Humann and L. Lenz; Bacterial peptidoglycan degrading enzymes and their impact on host muropeptide detection, *J. Innate Immun*, 2009; 1: 88-97.
- [10] N. Z. Gammoh and Lothar Rink; Zinc in infection and inflammation, *Nutrient*, 2017; 9: 1-25.
- [11] M. Maywald, Inga Wessels and L. Rink; Zinc signals and immunity, *J. Mol. Sci.* 2017; 18: 1-34.
- [12] M. L. Atilano, P. M. Pereira, F. Vaz et al; Bacterial autolysins trim cell surface peptidoglycan to prevent detection by the *Drosophila* innate immune system. *ElifeSciences*. 2014; 3: 1-23.
- [13] J. Humann, LL. Lenz ; Bacterial peptidoglycan degrading enzymes and their impact on host muropeptide detection, *J. Innate Immun*. 2009; 1 (2): 88-97.
- [14] T. Ishida, Comparative bacteriolytic mechanism for Ag<sup>+</sup> and Zn<sup>2+</sup> ions against *S.aureus* and *E. coli*: A review. *Annals of Microbiology and Infectious Diseases*, 2019; 2, Issue 1: 1-12.
- [15] A. Vermassen, S. Leroy, Regine Talon et al; Cell wall hydrolases in bacteria: Insight on the diversity of cell wall amidases, glycosidases and peptidases toward peptidoglycan. *Frontiers in Microbiology*. 2019; 10, article 331: 1-27.
- [16] F. M. Buttner, S. Zoll, M. Nega et al; Structure-function analysis of *S.aureus* amidase reveals the determination of peptidoglycan recognition and cleavage. *Journal of Biological Chemistry*. 2014; 289 (16): 11083-11095.
- [17] W. Hell, S. Reichl, A. Agnes, S. Gatermann; The autolytic activity of the recombinant amidase of *Staphylococcus saprophyticus* is inhibited by its own recombinant GW repeats. *FEMS Microbiology Letters*. 2003; 227: 47-51.
- [18] Ahn, Sun-Hee, D-G Kim, S-H. Jeong et al; Isolation of N-acetylmuramoyl-L-alanine amidase gene (*amiB*) from *Vibrio anguillarum* and the effect of *amiB* gene deletion on stress responses. *J. Microbiol. Biotechnol.* 2006; 16 (9): 1416-1421.
- [19] J. D. Lenz, E. A. Stohl, R. M. Robertson, K. T. Hackett et al; Amidase activity of *AmiC* controls cell separation and stem peptide release and is enhanced by *NlpD* in *Neisseria genorrhoeae*. *Journal of Biological Chemistry*. 2016; 291, No. 20: 10916-10933.
- [20] P. Mellroth, T. Sandalova, A. Kikhney et al; Structural and functional insights into peptidoglycan access for the Lytic amidase *LytA* of *Streptococcus pneumoniae*. *Mbio. Asm. Org.* 2014; 5, Issue 1: 1-10.
- [21] C. C. Kietzman, G. Gao, B. Mann et al; Dynamic capsule restructuring by the main pneumococcal autolysin *LytA* in response to the epithelium. *Nature Communications*. 2016; 108: 1-9.
- [22] L.R.Brown, R. C. Caulkins, T. E. Schartel et al; Increased zinc availability enhances initial aggregation and biofilm formation of *Streptococcus pneumoniae*. *Frontiers in Cellular and Infection Microbiology*. 2017; 7, article 233:1-7
- [23] P. Rico-Lastres, R. Diez-Martinez, M. Iglesias-Bexiga et al; Substrate recognition and catalysis by *LytB*, a pneumococcal peptidoglycan hydrolase involved in virulence. *Scientific Reports*. 2015; 5:1-17.
- [24] R. Chen, S. B. Guttenplan, K. M. Blair, and D.B. Kearns; Role of the D-dependent autolysins in *Bacillus subtilis* population heterogeneity. *Journal of Bacteriology*. 2009; 191, No.18: 5775-5784.
- [25] S.Zoll, M. Schlag, A. V. Shkumatov, M. Rautenberg et al; Ligand-binding properties and conformational dynamics of autolysin repeat domains in *Staphylococcal* cell wall recognition. *Journal of Bacteriology*. 2012; 194, No. 11: 3789-3802.
- [26] Vineet K. Singh; High level-expression and purification of *Atl*, the major autolytic protein of *Staphylococcus aureus*. *International Journal of Microbiology*. 2014; 2014 (4):1-7.

## Zinc Immunology and Zn<sup>2+</sup> Ions Binding Anti-Bacterial Vaccine Activity for Bacterial Cell Walls against Gram-Positive and Gram-Negative Bacteria

- [27] P. Houston, S.E. Rowe, C. Pozzi, et al; Essential Role for the major autolysin in the fibronectin-binding protein-mediated *Staphylococcus aureus* biofilm phenotype. *Infection and Immunity*. 2011; 70, No. 5: 1153-1165.
- [28] V. Dubee, F. Chau, M. Arthur, et al; The *in vitro* contribution of autolysins to bacterial killing elicited by amoxicillin increases with inoculum size in *Enterococcus faecalis*. *Antimicrobial Agents and Chemotherapy*. 2011; 55, No. 2: 910-912.
- [29] E. K. Stinemetz, P. Gao, K. L. Pinkston et al; Processing of the major autolysin of *E. faecalis*, AtlA, by the zinc-metalloprotease, GelE, impacts AtlA septal localization and cell separation. *PLOS ONE*. 2017; 12 (10) :1-17.
- [30] S. G. Murphy, L. Alvarez, M. C. Adams et al; Endopeptidase regulation as a novel function of the Zur-dependent zinc starvation response. *mBio*. 2019; 10, issue 1: 1-15.
- [31] Shirin Tarahomjoo; Serotype independent vaccine design against *Streptococcus pneumoniae* based on B-cell epitopes of autolysin, zinc binding lipoprotein and plasmid stabilization protein. *American Journal of Medical and Biological Research*. 2016; 4, No. 5: 84-89.
- [32] A. Raz, N. D. Djadid, S. Zakeri; Molecular characterization of the carboxypeptidase B1 of *Anopheles stephensi* and its evaluation as a target for transmission-blocking vaccines. *Infection and Immunity*. 2013; 81, No. 6: 2206-2216.
- [33] Characterization of the M32 metallo carboxy peptidase of *Trypanosoma brucei*: differences and similarities with its orthologue in *Trypanosoma cruzi*. *Mol Biochem Parasitol*. 2012; 184 (2): 63-70.
- [34] A novel recombinant vaccine candidate comprising PBP2a and autolysin against methicillin resistant *Staphylococcus aureus* confers protection in the experimental mice. *Molecular Immunology*. 2017; 91:1-7.
- [35] Y. Zhang, J. Su and D. Wu; Physiology and pathology of multidrug-resistant bacteria: Antibodies- and vaccine-based pathogen-specific targeting. *Physiology and Pathology of Immunology*. Chapter 10. 2018, 199-232 pages, INTECH.
- [36] M. Wang, L-H. L. Liu, S. Wang et al; Human peptidoglycan recognition proteins require zinc to kill both Gram-positive and Gram-negative bacteria and are synergistic with antibacterial peptides, *The Journal of Immunology*. 2007; 178: 3116-3125.
- [37] M.P. Cabral, P. Garcia, A. Beceiro et al; Design of live attenuated bacterial vaccines based on D-gltamate auxotrophy, *Nature Communications*. 2017; 10: 1- 17.
- [38] V. Cafardi, M. Biagini, M. Martinelli, et al; Identification of a novel zinc metalloprotease through a global analysis of *Clostridium difficile* extracellular proteins. *PLOS ONE*. 2013; 8, issue 11: 1-14.
- [39] F. Romero-Saavedra, D. Laverde, A. Budin-Verneull et al; Characterization of two metal binding lipoproteins as vaccine candidates for enterococcal infections. *PLOS ONE*. 2015; :1-15.
- [40] Pa T. Ngom, S. Howie M. O. Ota, et al; The potential role and possible immunological mechanisms of zinc adjunctive therapy for severe Pneumonia in children. *The Open Immunology*. 2011; 4: 1-10.
- [41] C. Formosa-Dague, P. Speziale, T. J. Foster et al; Zinc-dependent mechanical properties of *Staphylococcus aureus* biofilm-forming surface protein SasG. *PNAS*. 2016; 113, No. 2: 1-6.
- [42] A critical role of zinc importer AdcABC in group A *Streptococcus*-host interactions during infection and its implications for vaccine development. *EbioMedicine*. 2017; 21: 131-141.
- [43] X. Wang, C. D. Thompson C. Weidenmaier and J. C. Lee; Release of *Staphylococcus aureus* extracellular vesicles and their application as a vaccine platform. *Nature Communications*. 2018; 9(1): 1379-1392.
- [44] Choline binding proteins from *Streptococcus pneumoniae*: A dual role as enzymatics and targets for the design of new antimicrobials. *Antibiotics*. 2016; 21: 1-33.
- [45] Weiping Zhang and David A. Sack; Current progress in developing subunit vaccines against enterotoxigenic *Escherichia coli*-associated diarrhea. *Clinical and Vaccine Immunology*. 2015; 22, No. 9: 983-991.

## Zinc Immunology and Zn<sup>2+</sup> Ions Binding Anti-Bacterial Vaccine Activity for Bacterial Cell Walls against Gram-Positive and Gram-Negative Bacteria

- [46] D. A. Sack, F. Qadn, A-M. Svennerholm; Determinants of responses to oral vaccines in developing countries. *Ann Nestle*. 2008; 66 :72-79.
- [47] M I Qadir, A. Arshad, B. Ahmad; Zinc: Role in the management of diarrhea. *World Journal of Clinical Cases*. 2013; 1(4): 140-142.
- [48] Abduwahab MA Telmesani; Oral rehydration salts, zinc supplement and rota virus vaccine in the management of childhood acute diarrhea. *Journal of Family and Community Medicine*. 2010;17,issue 2:79-82.
- [49] V. G. Pederick, B. A. Eijkelkamp, S. L. Begg; ZnuA and zinc homeostasis in *Pseudomonas aeruginosa*. *Scientific Reports*. 2015; 5:1-14.
- [50] Hoggarth A, Weaver A, Pu Q, et al; Mechanistic research holds promise for bacterial vaccines and phage therapies for *Pseudomonas aeruginosa*. 2019; 13: 909-924.
- [51] T. F. Murphy, A. L. Brauer, A. Johnson; A cation-binding surface protein as a vaccine antigen to prevent *Moraxella catarrhalis* otitis media and infections in chronic obstructive pulmonary disease. *Clinical and Vaccine immunology*. 2017;24,issue 9: 1-15.
- [52] L. Gomez, J. Llanos, E. Escalona et al; Multivalent fusion DNA vaccine against *Brucella abortus*. *BioMed Research International*. 2017; 2017: 1-8.
- [53] A. Sirelkhatim, S. Mahmud, A. Seeni; Review on zinc oxide nanoparticles: Antibacterial activity and toxicity mechanism. *Nano-Micro Letters*. 2015;7(3): 219-242.
- [54] K. Salahuddin Siddigi, A. ur Rahman, Tajuddin, and A. Husen; Properties of zinc oxide nanoparticles and their activity against microbes. *Nanoscale Res Lett*. 2018; 13 (1): 141-154.
- [55] U. Kadiyata, E. S. Turali, J. H. Bahng et al ; Unexpected insights into antibacterial activity of zinc oxide nanoparticles against *MRSA*. *Nanoscale*. 2019; 10(10) :1-21.

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