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Immune Regulation: The Possible Mechanism by which Dehydroepiandrosterone Improves Ovarian Reserve in Patients Having Diminished Ovarian Reserve (Dor)

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Abstract

Diminished ovarian reserve (DOR) or poor ovarian reserve (POR) is becoming a worldwide problem and becoming an important cause of infertility. There has been concentration on this subject by the laboratories of Gleicher et al and to some extent Solano ME et al who found importance of androgenic steroids in DOR. For the last decade role of DHEA has been studied, be it their secretion getting reduced with age from the adrenal gland or importance of improving results in in vitro fertilization in cases of DOR. It has a proinflammatory effect on immune function which opposes cortisol. It acts as an immune regulator in various tissues including ovaries. In mammals it also improves Th1 immune response. Treatment with DHEA selectively increases T lymphocytes, upregulating CD8+T lymphocyte population in ovaries while down regulating CD4+T lymphocyte population and thus balance of the CD4+/CD8+ population is maintained. Further Gleicher et al studied regarding the disturbed autoimmune status along with studying triple CGG repeats in fragile X mental retardation 1(FXMR1) gene besides role of autoimmune antibodies in case of premature ovarian insufficiency (POI). Still very few studies are available and more work needs to be done regarding the mechanism by which DHEAS is efficacious in improving IVF results in DOR or POI.

Keywords: DHEA; DHEAS; immunefunction; HPA axis;cortisol; Th1/Th2 cytokines; CD4+/CDB+ Balance in ovaries; FXMR1.

INTRODUCTION

Ovarian reserve by definition is thequantity and quality of the ovarian primordial follicular pool. Diminished ovarian reserve (DOR) is a decrease in thequantity of ovarian poolin women of reproductive age group, a reduction in the dynamic reserve of small antral follicles and deteriorating oocyte quality. These changes are seen once there is ageing of women. Increasing use of in vitro fertilization (IVF) has unmasked the occurrence of DOR. Roughly 10% of women undergoing IVF will show a poor response to gonadotropin stimulation [1-3]. Various genetic mutations and disorders of the endocrine system can increase or modulate the rate at which ovarian reserve gets exhausted and leads to premature ovarian insufficiency (POI) [4]. Abnormal immune function might be an important cause of DOR [5]. Thus for these patients becoming infertile and showing poor response to in vitro fertilization (IVF) different protocols to solve this problem have been proposed though none has proved to be ideal for these patients [6-8]. Dehydroepiandrosteronesulphate (DHEA) is a C19 androgenic steroid which has been found to be effective in many areas. Years of studies both in clinical setups and in animals have shown that levels of DHEA (S) are inversely correlated with cardiovascular risk, morbidity as wellas mortality [9]. DHEA protected against atherosclerosis in ovariectomized rabbits

since it alleviates inflammation in endothelial cells [10]. It can crossthe blood brain barrier (BBB), it has neuro active characteristics and positively influences mood, emotions and behavior in man [11, 12]. DHEA supplementation was used to improve the response to ovarian supplementation by Casson et al to begin with [13]. Since then a lot of workers have paid attention to improve DOR using DHEA. A meta-analysis was conducted by Narwechian et al, which revealed that DHEA administration caused a significant increase in the number of oocytes retrieved in women with DOR, based on some clinical trials. But they concluded that more trials were needed for verifying these results [14]. Also in ovariectomized sheep DHEA addition was found to be effective in increasing ovarian folliculogenesis [15]. Similarly in Wistar rats used as an in vivo model the responses were found [16]. Still the basic mechanism by which DHEA improves this ovarian reserve remains unclear. It has been found that DHEA can regulate immune function [17], and that it might regulate immune cellfunction of many tissues in mammals. Utililizing a human subcutaneous preadipocyte cell line, Chub -67, it was found by McNeils et al that DHEA inhibits the amplification of (11β-HSD1) [18]. Also DHEA served as a neurosteroid directly by directly interacting with nerve growth factor (NGF) as shown by Lazardis etalin an in vitro study for preventing neuronal apoptosis [19]. DHEA also improves oxidative stress induced endothelial dysfunction in ovariectomized rats [20]. Also DHEA treatment may also modulate the lymphocyte responses based on both human and animal trials [21, 22]. Thus DHEA may have an important rolein regulating the immune response in ovaries, which is the main aim of this review.

DHEAS IN DOR

Structure and Biochemistry

Structurally DHEAS is 5-androsten- 3β -ol-17-one which is a C19 androgenic steroid, which is mainly secreted by the adrenal zonareticularis. It gets synthesized by the steroidogenic enzyme P450c 17 and also partly by ovary (fig1) [23]. Its secretion has a diurnal rhythm, which is similar to that of cortisol [24, 25]. With increasing age both DHEA and its sulphate gradually declines, which is unlike that of other androgenic steroids. DHEA concentrations peak during the 3^{rd} decade of life, having a clear sex difference

because adult women have lower concentrations as compared to men [25-27]. Because of this age related fall supplementation of DHEA might be of helpin improving age related damage in human beings.



Figure 1. Chemical structure of DHEA

DHEA Supplementation & DOR

No set criteria for DOR exist. Following the consensus meeting of ESHRE working group on DOR held in 2011 [28], Bologna criteria were introduced, which recommends at least 2 of the following 3 feautures be present for labeling POR or DOR

i) Advanced maternal age (>=40years) or any other risk factor for POR

ii) A previous DOR (<=300cytes with a conventional stimulation protocol).

iii) An abnormalovarian response test (ORT) (i.e antral follicle count (AFC), 5-7 follicles, or antimullerian hormone (AMH), 0.5-1.1ng/ml).

Although despite these guidelines different researchers have used different criteria besides these Bologna criteria.

Many studies have examined the effect of DHEA supplementation on ovarian hormones. Analyzing 12 of those studies Zhang et al analyzed that DHEA supplementation is usually given orally t.i.d, or 75-90mg for the full day over 6-24 weeks [29]. It was shown in a previous case controlled studythat DHEA improved ovarian reserve (OR), and it significantly increased AFC and AMH levels [30-32], numbers of fertilized oocytes, normal day3 embryos, embryo transferred as well as the average embryoscore per oocyte [33]. Also they significantly decreased day3 follicle stimulating hormone (FSH) [31], fertilized aneuploid embryos [34], and miscarriage rates [35] (withallp values<0.05). Estradiol (E2) levels increased three times as per 2 studies [13, 36], but decreased according to another

one [32]. A lot of randomized prospective controlled studies investigated the effects regarding DHEA supplementation but with inconsistent results. As per some same results as described before [32, 36-38], but total oocytes retrieved and the fertilization rate was not consistent. Also IVF outcomes were not improved by DHEA in others [38, 39]. One can not compare these studies statistically however as there was no set criteria regarding defining DOR. DOR was defined as abnormally increased age-specific baseline FSH levels and/or abnormally low AMH levels [40-42], increased inhibin levels and AFC<4-5 [31, 32, 36, 39].

IMMUNE FUNCTIONS OF DHEA

DHEA exerts an effect on human neuroendocrine cells along with playing an important role in immune regulation, mainly by balancing proinflammatoryand anti-inflammatory signals. Inflammation develops in humans with age, that involves up regulation of some proinflammatory cytokines like interleukin (IL)-1, IL-6, tumour necrosis factor (TNF)-α, IL-12, interferon (IFN)- α and IFN- β . These cytokines negatively impact in old age on various systems of the body [43-45]. Hence an imbalanced relationship develops between proinflammatory cytokines and anti-inflammatory cytokines like IL-4, IL-6, IL-13 and IL-10 [46]. This is a part of hypothalamo-pituitary-adrenal (HPA)-axis. If HPA Axis is activated, both cortisol along with DHEA get released. Although cortisol has an anti-inflammatory effect, DHEA seems to have an opposite effect. With decreasing levels of DHEA with age there is an increase of cortisol: DHEA ratio which might interact with a weakened immune function [47, 48]. Increased cortisol: DHEAS ratio was seen in bereaved elderly participants, who showed decreased production of neutrophil reactive species [11]. A positive effect on immunity was seen with DHEA supplementation in the elderly. 20mg/day DHEA treatment for 8weeks reversed antioxidant parameters like decreased superoxide dismutase activityin the brain and heart, reduced inducible nitric oxide synthase mRNA levels and increased haemoxygenasem RNA levels in aged rats [49]. Cortisol acted in a dose related manner, both in vitro and vivo as shown by Buoso et al on human guanine nucleotide binding protein and the beta polypeptide 2 like1 (GNB2L1) promoter repressor, that decreased receptor for activated C kinase (RACK1)m RNA and protein expression. Using prolonged DHEA exposure counteracted the effect

of cortisol and restored RACK1 levels and cytokine production as assessed by lipopolydaccharide (LPS) Induced TNF- α release, which most likely occurred due to interfering with glucocorticoid receptor binding to the glucorticoid response element (GRE) sequence [50]. Other diseases and improved organ function and survival has been seen by use of DHEA supplementation. Still there mechanisms are not clearly understood. In past decades reasearchers have tried to study these effects in nonhumanmammals. In aged baboons, they found an increased C- Re active protein and increased cytokine release from unstimulated peripheral blood mononuclear cells (PBMC). Outcomes in amurine polymicrobial sepsis and trauma model by restoring TNF α in the liver and lungs occurred 48h after supplementation with DHEA and attenuated it in the liver after 96h, much like atime and organ dependent modulator [51]. Also restoration of splenocyte proliferation, decrease in rate of cellular apoptosis of splenocytes,

besides an attenuation of increased IL-6 levels is caused by DHEA addition [52]. DHEA also reduced PBMC's function and increased the ability of human PBMC patients with depressed response to release proinflammatory cytokines-IL-1 β , IL-6 and TNF- α , following abdominal surgery [53].

DHEA-Immune Response in Ovaries

Level of ovarian response in the ovaries changes with age along with ovarian reserve. Lot of interaction between immune systemand the ovaries occurs as the immune cells are associated with regulation of every step of the H-P-O Axis by regulating growth and regression of both follicles and corpus luteum (CL) [54-56]. In an adult ovary activated myeloid dendritic cells (MDC) also play a role in follicular development and atresia along with differentiation of the corpus luteum. There is massive infiltration of MDC and Tcells in the corpus luteum, which results in parenchymal and vascular regression of the CL [57]. OR of women with age related DOR is improved by DHEA in POI, even though POI is more closely related toovarian immune disorders. Young women having a history or family history of autoimmunity are at risk for POI [58, 59]. Many types of autoimmune ovarian autoimmune insufficiency abnormalities exist. Three different types of ovarian autoimmune insufficiency, autoimmune ovarian insufficiency associated with adrenal autoimmunity,

autoimmune ovarian insufficiency associated with nonadrenal autoimmunity and isolated idiopathic POI [60]. Most prevalent is adrenal autoimmunity [4, 61]. Testing for triple CGG repeats on both alleles of the fragile X mental retardation1 (FMR1) gene and assessed autoimmune status including an antiphospholipid antibody panel, an antinuclear antibody panel, total immunoglobulin levels, the thyroid antibodiesand antiadrenal antibodies then these researchers (group from lab of Gleicher et al) found that abnormal autoimmune function ,including expansion of triple CGG repeats on the FMR1 gene increased the risk of POI [62, 63].

Though the effects of DHEA supplementation in women with DOR are clearly evident the mechanism behind these effects still are not clear. With the compleximmune function of DHEA in women with DOR, some researchers have started studying the immune function of DHEA in women with DOR. Still limited research reports on this topic are available till date. Summary of the these limited studies is presented.

Conversion to other Steroids

There are no specific receptors for DHEA found till date, some theories suggest that DHEA might functiononce steroidogenic enzymes convert it to other downstream steroids, specially sex steroidslike estrogen and androgens (fig2) Small structural changes in androgens =>markedly different biological

effects. There are tissue specific expressions of steroidogenicenymes, and hence DHEA may have a specific function. These steroids interact critically with immune function. Estrone can shift the female immune system to a Th2-type response in the luteal phase, while postmenopausal women exhibit increased Th1 cytokines [64]. Also DHEA can restore PBMC function and increase the ability of human PBMC's torelease proinflammtory cytokines after surgery via the estrogen receptor; thus this immunomodulatory effect of DHEA appears to be connected to estrogen receptors - [53]. Though total androgen concentrations are not associated with pregnancy during DHEA supplementation with POI, interaction between DHEA and total and free testosterone also significantly affect pregnancy rate at the start of an IVF cycle [65]. There is significantly lower total testosterone in women with POI or anormal FMR1 genotypes [66]. Efficiency of androgen conversion from DHEA to testosterone(T) and the amplitude of Tgain are related to pregnancy rates. The conversion Is more pronounced in young women and women with selected FMR1 genotypes /subgenotypes [67]. DHEA and T also suppress concavalin A(Con A)-induced proliferation of thymocytesin vitro and DHEA is less potent than T, which means that the balance between the 2 steroids can alter immune homeostasis [68]. T and E2 vary widely following DHEA administration, and T:E2 ratio increased significantly in 7 nonobesepostmenopausal women [69].



Figure 2. The regulatory pathway by which DHEA Conversion occurs to other downstream steroids courtesy ref 38.

Abbreviations

DHEA, dehydroepiandrosterone; DHEAS, dehydro epiandrosterone sulfate; HSD 17β, 17β-hydroxy

steroid dehyrogenase isoenzymes; CYP19A1, P450 aromatase; SRD 5A, 5α reductase isoenzymes; AKR1C, 3α -hydroxy steroid dehyrogenaseisoenzymes.

Once steroidogenic enzymes convert DHEA to other steroids, these enzymes have a substantial effect on the immune response. Though women with POI have a poor ovarian follicle pool as compared to healthy volunteer fertile controls, women with POI and steroidogenic cell autoimmunity (SCA-POI) that involves autoantibodies directed against steroidogenic enzymes like 21 – α -hydroxylase, 17- β hydroxylase, and side chain cleavage enzyme (P450 sec Ab) [70-74], have a better OR than women with iPOI and postmenopausal women. Steroidsulphatase is controlled by an X linked gene. Women have twice the amount of steroid sulphatase in macrophages. The macrophages enter peripheral lymphoid organs through afferent lymphatic drainage [75,76]. IL-4 which is a typical Th2 cytokine increases the expression of 3-beta-hydroxy steroid dehydeogenase (HSD3B2) m RNA and thus might lead to the increased production of estrogen from DHEA [77].

Control of Th1 and Th2 Response

Normally cytokines play important role by influencing communication between T cells, macrophages and other immune cells. It has been deduced from studies in mice along with humans that there is presence of helper T (Th) cells which is based on the profile of cytokine secretion. The Th1-type immune response is believed to be associated with IgG2 a production that is driven by cytokines like IFN- γ , IL-2, TNF- α , and IL-12, while Th2-type immune response involves IgE production which is driven by specific cytokines like IL-4, IL-5, IL-10 and IL-13 [78, 79]. Restoration of cell mediated immune response of proinflammatory cytokines like IL-1, IL-6, TNF- α and IL-18 is brought about by DHEA. Treatment of mice with DHEA showed elevated production of cytokines like serum TNF –α, IL-6, IL-12 p70 and IFN-γ [80, 831]. It also raised vascular cell adhesion molecules (VCAM1) and intercellular celladhesion molecule 1(ICAM-1) in the granulosa cell layer cysts and the theca cell layer of all follicles and cysts when DHEA and rogenization causes the cyst formation [81]. There might be improvement of ovarian reserve by DHEA in patients having DOR by stimulating antiapoptotic processes in cumulus cells. Upregulation of genes related to extracellular matrix (ECM) formation is probably involved in these processes along with down regulation of genes which are related to cell development, differentiation, and apoptosis [82]. Also DHEA supplementation has been shown to have an effect on Th1/Th2 response.

Asthmatic female mice that are ovalbumin –sensitized show inhibition of Th2-associated cytokines and chemokines following administration of DHEA which possibly led to hyporesponsiveness [83]. Release of anti-inflammatory cytokines like IL-2 and IL-10 was also decreased by DHEA, that are Th2 –associated cytokines, along with reduction in the expression of activation marker CD69 on CD4+T cells [21]. Summing up these effects might be associated with an increased Th1 response along with weakening of Th2 response.

Effect on Interaction between CD4+/CD8+T Cells

Further DHEA improves immune function by regulating the multiplication of and balance between various kinds of lymphocytes. While culturing T Lymphocytes from BALB c mice in vitro, there was no change in viability of T Lymphocytes, although DHEA did increase the oxidative stress by decreasing antioxidant molecules, like glutathione (GSH) [84]. Orally when DHEA was given to young pigs DHEAS increased in vitro proliferation following immunization and it increased the in vivo response of immunization against keyhole limpet haemocyanin (KLH), thus increasing the neutrophil : lymphocyte ratio and increasing the concentration of IgG as shown by Burdick et al [22]. Effects of DHEAS are just opposite to that of cortisol : hencecortisol : DHEA ratiomight influence the differentiation of Tcells. There is a positive correlation of extrathymic (DP)CD4+/ CD8+ Tcells with circulating levels of TNF- α and with cortisol/DHEA ratio [85]. Flow cytometry showed that DHEA treatment in mice markedly increased the CD4+Lymphocytepopulation while reducing CD8+ lymphocyte population and hence modulating the CD4+/CD8+ lymphocyte balance in both ovarian tissue as wellas retroperitoneal lymph nodes [80]. Probably it is related to the selective T Lymphocyte infiltration of the ovarian tissue [86].

CONCLUSIONS

Thus DHEAS has been found to be effective in patients of DOR, helping getting bettet outcomes in IVF cycles. It might be regulating ovarian immunity via its conversion to other downstream steroids.ii) by balancing Th1/Th2 immune responses or kulvinder modulating types and balance of lymphocytes. Still the mechanism underlying immune effects of DHEA on ovarian tissue requires further clarification along with more number of studies.

References

- Garcia JE, JonesGS, Acosta AA, Wright G. Human menopausal gonadotropin/human chorionic gonadotropin follicular maturation for oocyte aspiration: Phase II, 1981. Fertil Steril 1983; 39:174-9.
- [2] Pellicer A, Lightman A, Diamond MP, RussellJ B, De Cherney AH. Outcome of in vitro fertilization in women with low response to ovarian stimulation. FertilSteril 1987; 47:812-5.
- [3] Keay SD, Liversedge NH, Mathur RS, Jenkins JM. Assisted conception following poor ovarian response to gonadotropin stimulation. Br J Obstet Gynecol 1997;104:521-7.
- [4] Moniaux D, Clement F, Dalbias –Tran R, Estienne A, Fabre S, Mansanet C, Monget P. The ovarian reserve of primordial follicles and the dynamic reserve of antral growing follicles. What is the link?BiolRepr 2014; 90:85.
- [5] Silva CA, Yamakami LY, Aikawa NE, Araujo DB, Carvalho JF, Bonfa E. Autoimmune primary ovarian insufficiency. Autoimmun Rev 2014;13:427-430.
- [6] Karande VC,. Managing and predicting low response to standard in vitro fertilization therapy: A review of the options. Treat Endocrinol 2003;2:257-272.
- [7] Loutradis D, Vomvolaki E, Drakakis P. Poor responder protocols for in vitro fertilizationtherapy. Options and results. Curr Opin Obstet Gynecol 2008; 20:374-378.
- [8] Caglar Ayte P, Kiliedag EB, Haydardedeoglu B, Simsek E, Cok T, Parlakgumus HA. Can calcium ionophore "use" in patients with diminished ovarian reserve increase fertilization and pregnancy rates?A randomized controlled study. FertilSteril 2015;104:1168-1174.
- [9] Mannie T, Viguie J, Rossier MF. In vivo and in vitro evidences of dehydroepiandrosterone protective roleon the cardiovascular system. Int J EndocrinolMetab 2015;13:e24660.
- [10] Wang L, Hao Q, Wang YD, Wang WJ, Li D, DJ. Peotective effects of dehydroepiandrosterone on atherosclerosis in ovariectomized rabbits via all eviating inflammatory injury in endothelial cells. Atherosclerosis 2011; 214:47-57.

- [11] Vitlie A, Khanfer R, Lord JM, Carrol D, Phillips AC. Bereavement reduces neutrophil oxidative burst in older adults:Role of the HPA axis and immunesenescence.Immun Ageing 2014;11:13.
- [12] Starka L O Duskova M, Hill M. Dehydroepiandro sterone:Aneuroactivesteroid. J Steroid Biochem Med Biol 2015; 145:254-260.
- [13] Casson PR, Lindsay MS, Pisarka MD, Carson SA, Buster JE. Dehydroepiandrosterone supplementation augments ovarian stimulation in poor responders: A case series Human Reprod 2000;15:2129-2132.
- [14] Narkwichean A, Maalouf W, Campbell RK, Jayaprakasan K. Efficacy of Dehydro epiandrosterone to improve ovarian reserve in women with diminished ovarian reserve : A metaanalysis.ReprodBiolEndocrinl 2013 ;11:44.
- [15] Narkwichean A, Jayaprakasan K, Maalouf W, Hernandez – Medrano JH, Pincott Allen C, Campbell RK. Effects of Dehydroepiandrosterone on in vitro ovine follicular development. Hum Reprod 2014;29:146-154.
- [16] Hassa H, Aydin Y, Ozatik O, Erol K, Qzatik Y, Effects of. Dehydroepiandrosterone (DHEA) on the follicular dynamics in a siminished ovarian reserve in vivo model. SystBiolReprod Med 2015; 61:117-121.
- [17] HazeldineJ,AritW,LordJM.Dehydroepiandrosterone as a regulator of immune cell function.J Steroid BiochemMolBiol 2010 ;120:127-136.
- [18] McNeils JC, Manolopous KN, Gathercole LL, Bajalska IJN, Stewart PM, Tumlinson JW Arlt W. Dehydroepiandrosterone exerts antiglucocorticoid action on human pre adipocytepro life ration, differentiation and glucose uptake. Am J Physiol Endocrinol Metab 2013;305: E1134-1144.
- [19] Lazardis J, Charalampopolous I, Alexis VI, Avlonitis N, Pediaditakis I, Efstathoupoulos P, Calogeropoulou T, Castanas E, Gravanis E. Neurosteroid Dehydroepiandrosterone interacts with nerve growth factor (NGF) receptors preventing neuronal apoptosis. PLoS Biol 2011;9:e1001051.
- [20] Camporez JP, Akamine EH, Davel AP, Franci CR, Rossoni LV, Carvalho CR. Dehydroepiandrosterone

protects against oxidative stress – induced endothelial dysfunction in ovariectomizedrats. JPhysiol 2011;589:2585-2596.

- [21] Pratschke S, Von Dossow-Hanfsting IV, Dietz J, Schneider CP, Tufman A, Albertsmeier M, Winter H, Angele MK. Dehydroepiandrosterone modulates T-cell response after major abdominal surgery. JSurg Res 2014; 189:117-125.
- [22] Burdick NC, Dominguez JA, Welsh TH Jr, Laurenz JC. Oral administration of Dehydroepiandrosterone sulfate (DHEAS) increases in vitrolymphocyte function and improves in vivo response of pigs to immunization against keyhole limpet hemocyanin (KLH) and ovalbumin. IntImmunopharmacol 2009;9:1342-1346.
- [23] Burger HG. Androgen production in women. FertilSteril 2002;77 (Suppl4) :S3-S5.
- [24] Liu CH, Laughlin JA, Fischer UG, Yen SS. Marked attenuation of ultradian and circadian rhythms of Dehydroepiandrosterone in postmenopausal women:evidence for a reduced 17, 20-desmolase enzymatic activity. J ClinEndocrinol Metab 1990;71:900-906.
- [25] Orentreich N, Brind JL, Rizer RL, Vongelman JH. Age changes and sex differencesin serum Dehydroepiandrosterone sulfate concentrations throughout adulthood. J Clin Endocrinol Metab 1984;59:551-555.
- [26] Palmert MR, Hayden DL, Mansfield MH, Crigler JF. Jr, Crowley WF Jr, Chandler DW, Boepple PA. The longitudinal study of adrenal maturation du ring gonadal suppression.Evidence that adrenarche is a gradual process. J Clin Endocrinol Metab2001;86:4536-4542.
- [27] Reiter EO, Fuldauer VG, Root AW. Secretion of the adrenal androgen, dehydroepiandrosterone sulfate during normal infancy, childhood, adolescence in sick infants and in children with endocrinologic abnormalities. J Pediatr 1977;99:766-770.
- [28] Ferrareti AP, La Marca A, Fauser BC, Tartlatzis B, Nargund G, Gianaroli L ESHRE Working Group on Poor Ovarian Response Definition. Eshre consensus and the definition of 'poor response'to ovarian stimulation for in vitro fertilization: The Bologna Criteria. Hum Reprod 2011; 26:1616-1624.

- [29] Zhang J, Qiu X, Gui Y, Xu Y, Li D, Wang L. Dehydroepiandrosterone improves the ovarian reserve of women with diminished ovarian reserve and is a potential regulator of the immune response in the ovaries. Biosciene Trends 2015;9(6):350-359.
- [30] Gleicher N, Weghofer A, Barad DH. Improvement in diminished ovarian reserve after dehydro epiandrosterone supplementation. Reprod Biomed Online 2010 ;21:360-365.
- [31] Yilmaz N, Uygur D, Inal H, Goekem U, Creek N, Mollamahmutoglu L. Dehydroepiandrosterone supplementation improves predictive markers for diminished ovarian reserve: Serum AMH, inhibin B, andantral follicle count. Eur J Obstet Gynecol Reprod Biol 2013;169:257-260.
- [32] Tsui KH, Lin LT, Chang R, Huang BS, Cneng JT, Wang PH. Effects of dehydroepiandrosterone supplementation on women with diminished ovarian reserve: A preliminary report and review. Taiwan J ObstetGynecol 2015; 54:131-136.
- [33] Barad D, Gleicher N. Effect of dehydro epiandrosterone on oocyte and embryo yields, embryo grade ande cell number in IVF. HumReprod 2006; 21:2845-2849.
- [34] Gleicher N, Weghofer A, Barad DH. Dehydro epiandrosterone (DHEA) reduces embryo aneuploidy: direct evidence from preimplantation genetic screening (PGS). Reprod Biol Endocrinol 2010; 8:140.
- [35] Gleicher N, Ryan E, Weghofer A, Blanco-Mejia S, Barad DH. Miscarriage rates after Dehydro epiandrosterone (DHEA) supplementation in women with diminished ovarian reserve:A case controlled study.). Reprod Biol Endocrinol 2009;7:108.
- [36] Zhang HH, Xu PY, Wu J, Zou WW, Xu XM, Cao XY, Wei LZ. Dehydroepiandrosteroneimproves follicular fluid bone morphogenetic protein-15 and accumulatedembryo score of infertility patients with diminished ovarian reserve undergoing invitrofertilization: Arandomized controlled trial.J Ovarian Research 2014;7:93.
- [37] Tartagni H, Cicinelli MV, Baldini D, Tartagni MV, Alrasheed H, DeSalvia MA, Loverro G, Montagnani

H. Dehydroepiandrosterone decreases the age related decline of the in vitro fertilization outcome in women younger than 40 years old. Reprod Biol Endocrinol 2015;13:18.

- [38] Poli E, Manfe S, Capuzzo D, Gava S, Vigano F, Coronella ML, Gangeni M. DHEA pretreated patients, poor responders to a first IVF (ICSI) cycle: clinicalresults. Clin Exp Obstet Gynecol 2014;41:5-9.
- [39] Kara M, Aydin T, Aran T, Turtekin N, Ozdemir B. Does Dehydroepiandrosterone supplementation really affect IVF-ICSI-in women with poor ovarian reserve?. Eur J ObstetGynecolReprodBiol 2014;173:63-65.
- [40] Barad DH Weghofer A, Gleicher N. Age specific levels for basal FSH assessment of ovarian function. Obstet Gynecol 2007; 109:1404-1410.
- [41] Barad DH Weghofer A, Gleicher N. Utility of age specific serumanti-mullerianhormoneconcentrations. Reprod BiolEndocrinol 2011; 22:284-291.
- [42] Fang T, Su Z, Wang L, Yuan P, Li R, Quyang N, ZhengL, Wang W. Predictive value of age -specific FSH levels for IVF-ET outcome in women with normal ovarian function. Reprod Biol Endocrinol2015;13:63.
- [43] McFarlane D, Wolf RF, Mc Daniel KA, White GI, Age associated alteration in innate immune response in captive baboons. J Gerontol A BiolSci Med Sci 2011;66:1309-17.
- [44] Fagiolo U, Cossarizza A, Scala E, Fanales-Belasio E, Ortolani C, Cozzi E, Monti D, Franceshi C, Paganelli R. Increased cytokine production in mononuclear cells of healthy elderly people. Eur J Immunol 1993;23:2375-2378.
- [45] Francheschi C, Monti D, Sansoni P, Cossarizza A. The immunology of exceptional individuals. The lessons of centenarians. Immunol Today 1995;16:12-16.
- [46] Lio D, Scola T, Crivello A, Colonna-Romano G, Candore G, Bonafe M, Cavallonel, Marchegiani F, Olivieri F, Francheschi C, Caruso C. Inflammation, genetics, and longevity. Further studies on the protective effects in men of IL-10-1082 promoter SNP.J Med Genet 2003;40:296-299.

- [47] Giunta S. Exploring the complex relation between. Inflammation and ageing (inflammpaging). Anti-imflamm-aging remodeling of inflammaging, from robustness to frailty. Inflamm Res 2008;57:558-563.
- [48] Butcher SK, Killampalli V, Lascelles D, Wang K, Alpar EK, Lord JM. Raisedcortisol "DHEAS ratios I the elderly after injury: Potential impact upon neutrophil function and immunity. Aging Cell 2005;4:319-324.
- [49] Yin FJ, KangJ, Han NN, Ma HT, Effect of Dehydroepiandrosterone treatment on hormone levels and antioxidant parameters on aged rats. GenetMol Res 2015;14:11300-11311.
- [50] Buoso E, Lanni C, Molteni E, Rousset F, Corsini E, Racchi M. Opposing effects of cortisol and Dehydroepiandrosterone on the expression of the receptor for Activated Kinase C Kinase 1:Implications in immunosenescence. Exp Gerontol 2011;46:877-883.
- [51] Barkhausen T, Hildebrand F, Krettek C, Van –Griensven M. DHEA dependent and organspecific regulation of TNF- α -MRNA expression in a murine polymicrobial sepsis and trauma model. Critical Care 2009 ;13:R114.
- [52] Schmitz D, KobbeP, Wegner A, Hammes F, Oberbeck RDehydroepiandrosterone during sepsis. Does the timing of administration influence the effectiveness .J Surg Res 2010;163:e73-77.
- [53] Frantz MC, Prix NJ, Wichman MW, Van den Engel NK, Hernandez –Richter T, FaistE, Chaudry IH, Jaunch KW, Angele MK. Dehydroepiandrosterone restores depressed peripheral blood mononuclear cell function following major abdominal surgery via the estrogen receptors. Critical Care Medicine 2005;33:1779-1786.
- [54] Vinatier D, Dufour P, Totdjeman-Rizzi N, Prolongeau JF, Depret-Moser S, Monnier JC. Immunological aspects of ovarian function. Role of the cytokines. Eur J ObstetGynecolReprodBiol 1995; 63:155-168.
- [55] Chrysdilopolous A. The relationship between the immune and endocrine system. Ann NY AcadSci 1997;816:83-93.
- [56] Patel JL. Involvement of immune cells in regulation of ovarian function. JR eprod Fertil Suppl1995; 49:365-377.

- [57] Bukovsky A, Caudle MR, Carson RJ, Gaytan F, Huleihel M, Kruse A, Schatten H, Telleria CM. Immune physiology in tissue regeneration and aging, tumor growth, and regenerative medicine. Aging (Albany NY) 2009; 1:157-181.
- [58] Gleicher N, Weghofer A, Barad DH. Cuuting edge assessment of the impact of autoimmunityin female reproductive system. JAutoimmun 2012; 38:374-380.
- [59] Cervera R, Balasch J. Bidirectional effects on autoimmunity and reproduction. Hum Reorod Update 2008; 14:359-366.
- [60] Carp HJ, Selmi C, Shoenfeld Y. The autoimmune bases of infertility and pregnancy loss. J Auto immun 2012; 38:J266-274.
- [61] Gleicher N, Weghofer A, Kushnir VA, Shohat-Tal A, Lazzaroni E, Lee HJ, Barad DH. Is androgen production with immune system activation potential evidence for existence of functional adrenal/ovarian autoimmune system in women ? Reprod Biol Endocrinol 2013;11:58.
- [62] Gleicher N, Weghofer A, Barad DH. Is the immunological noise of abnormal autoimmunity an independent risk factor premature ovarian aging ? Menopause 2009;16:760-764.
- [63] Gleicher N, Weghofer A, Barad DH. A pilot study of premature ovarian senescence II Different genotype and phenotype for genetic and autoimmune etiologies. Fertil Steril 2009;91:1707-1711.
- [64] Giron-Gonzalez JA, Moral FJ, Elvira J, Garcia-Gil D, Garrero F, Gavilan I, Escobar L. Consistent production of higher TH1:TH2 cytokine ratio by stimulated Tcells in men comared with women .Eur J Endocrinol 2000;143:31-36.
- [65] WeghoferA, KimH, BaradDH, GleicherN. The impact of androgen production d FMR1 genotypes on pregnancy potentialin women with dehydroepiandrosterone (DHEA) supplementation. Hum Reprod 2012; 27:3287-3293.
- [66] Gleicher N, Kim A, Weghofer A, Kushnir VA, Shohat-Tal A, Lazzaroni E, Lee HJ, Barad DH. Hypoandrogenism in association with diminished functional ovarian reserve Hum Reprod 2013;;28:1084-1091.

- [67] Gleicher N, Kim A, Weghofer A, Kushnir VA, Shohat-Tal A, LazzaroniE, Lee HJ, Barad DH. Starting and resulting testosterone levels after and rogensupplementation determines at all ages in vitro fertilization (IVF) pregnancy rates in women with diminished ovarian reserve .J Assist Reprod Genet 2013;30:49-62.
- [68] Yan G, Shang XJ,. Acomparison of modulation of proliferation of thymocyte by testosterone, dehydroepiandrosterone and rostenedione in vitro. ArchAndrol. 2005;51:257-265.
- [69] Caufriez A, Leprouli R, L'Hermite-Baleriaux M, Kerkhofs M, Copinschi G. Effects of a 3weeks dehydroepiandrosterone administration on sleep, sex steroids and multiple 24h hormonal profiles in postmenopausal women. Apilotstudy. ClinEndocrinol(Oxf)2013 ;79:716-724.
- [70] Hoek A, Schoemaker J, Drexhage HA. Premature ovarian failure and ovarian autoimmunity. Endocrin Rev 1997; 18:107-134.
- [71] Bakalov VK, Anasti JN, Calis KA, Vanderhoof VH, Premkumar A, Chen S, Furmaniak J, Smith BR, Merino MJ, Nelson LM. Autoimmuneoophoritis as a mechanism of follicular dysfunction in women with 46XX spontaneous premature ovarian failure. FertilSteril 2005;84:958-965.
- [72] Chen S, Sawicka J, Betterle C, Powell M, Prentice L, Volpato M, Rees Smith B. Furamaniak J. Autoantibodies to steroidogenic enzymes in autoimmunepolyglandular syndrome, Addisons disease and premature ovarian failure.J ClinEndocrinolMetab 1996; 81:1871-1876.
- [73] Betterle C, Dal Pra C, Mantero F, Zanchetta R. Autoimmuneadrenal insufficiency and autoimmune polyendocrine syndromes: Autoantibodies, autoantigensand their applicability in diagnosis and disease prediction. Endocr Rev 2002; 23:327-364.
- [74] Falorni A, Laurcti S, C andelero P, Perrino S, Coronella C, Bizzaro A, Bellastella A, Santeustanio F, De Bellis A. Steroid cell autoantiantibodies are preferentially expressed in women with premature ovarian failure who have adrenal autoimmunity. FertilSteril 2002;78:270-279.
- [75] Daynes RA, Aranci BA, Dowell FA, Huang K, Dudley D, Regulation of murine lymphokine production in vivo III. The lymphoid tissue microenvironment

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exerts reguloatory influences over T helper cell function. JExp Med 1990;171:979-996.

- [76] Namazi MR. Hypothesis:paradoxical absence of cellular immune-deficiency in X linked recessive ichthyosis and its explanation. JDermatolSci 2003;32:166-167.
- [77] Urata Y, LOsuga Y, Akiyamal, Nagai M, Izumi G, Takamura M, Hasegawa A, Harada M, Hirata T, Hirota Y, Yoshino O, Koga K, Kozuma S. Interleukin 4 and prostaglandin E2 synrrgistically upregulate 3beta-hydtoxy steroid dehydrogenase type2 in endometrioma stromal cells. Clin Endocrinol Metab 2013;98:1583-1590.
- [78] Belardeli F. Role of interferons and other cytokines in the regulation of the immune response. APMIS 1995;103:161-179.
- [79] Kasakura S. Arole for T helper type1 and type2 cytokines in the pathogenesis of various humndiseases. Rinsho Byori 1998;46:915-921.
- [80] Sander V, Luchetti CG, Solano ME, Elia E, Di Girolamo O, Gonzalez C. Motta AB. Role of the N, N' simrethylbiguanide metformin in the treatment of female prepubertal BALB/c mice hyperandrogenized with dehydroepiandrosterone. Reproduction 2006;131:591-602.
- [81] Solano ME, Sander VA, Ho H, Motta AH, Arek PC. Systemic inflammation, cellular influx and

upregulation of of ovarian VCAM -1 expression in a mouse model of poly cuystic syndrome. (PCOS). J ReprodImmunol 2011;92:33-44.

- [82] Tsui KH, Lin LT, Horng HC, Chang R, Huang BS, ChengJ T, Wang PH. Gen e expression of cumulus cells in women with poor ovarian reserve after dehydroepiandrosterone supplementation. Taiwan J ObstetGynecol 2014; 53:559-565.
- [83] Liou CJ, Huang WC, Dehydroepiandrosterone suppresses eosinophil infiltration and airway hyperresponsiveness via modulation of chemokines and Th2 cytokines in ovalbumin –sensitized mice. J ClinImmunol 2011; 31:656-665.
- [84] Solano ME, Sander VA, Wald MR, Motta AH. Dehydroepiandrosterone and metform in regulate proliferation of murine T lymphocyres. ClinExpImmunol2008;153:289-296.
- [85] Perez AR, Morrot A, Berberi LR., Terra-Granado E, Savino W,. Extrathymic CD4+CD8+ lymphocytes in Chagasdisease: Possible relationship with an immunoendocrine imbalance. Ann NY Acad Sci2012;1262:27-36.
- [86] Luchetti CG, Solano ME, SanderVA, Arcos MI, Gonzalez C.Di Girolamo O, Chiocchio S, Cremaschi G, Motta AB. Effects of Dehydroepiandrosterone on ovarian cystgenesis and immune function. JReprodImmunol 2004; 64:59-74.

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