

Review: Antimicrobial Property of Cow milk Protein

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ABSTRACT

The aims of this review were to know antimicrobial property of milk protein extracted from colostrum of cow milk. The heterogeneous mixture of cow milk can be divided into two basic groups, i.e. caseins and whey proteins, include of approximately 20% whey and 80% casein. Caseins are the most plentiful proteins of cow's milk. Their content varies inside the range of 2.6%–2.8%, which represents approximately 80% of general milk proteins. Therefore, casein epitopes is probably responsible for scientific signs. There are five casein fractions in cow's milk: $\alpha 1$, $\alpha 2$, β , κ , and γ , which represent 30%, 9%, 28%, 10% and more than 1% of the whole quantity of casein, respectively. Whey proteins constitute approximately 20% of milk composition. Whey proteins represent an important group with high nutritional and functional properties that have a positive effect on human body. Whey proteins include β -lactoglobulin, α -lactalbumin, lactoferrin, lactoperoxidase, lysozyme, bovine serum albumin, immunoglobulins, transferrin, proteose-peptones. The foremost antimicrobial proteins of milk whey protein are immunoglobulins, lactoferrin (Lf), lactoperoxidase (LP), and the lysozyme. Increase and acid production with the aid of starter cultures may be inhibited by bacteria and viruses bacteriophages, or brought substances including antibiotics.

Keywords: Immunoglobulins, Lactoferrin, Lactoperoxidase, lysozyme, Colostrum, bovine and milk.

INTRODUCTION

Milk is a longtime and healthful food source of power and it incorporate proteins nutrients and minerals. Further to its rate as a nutrient supply interest has arisen inside the capacity of milk to kill micro organism. Milk whey is a mixture of a variety of proteins. The mixture displays a wide range of chemical, physical and functional properties (C.A. Barth 1988). Whey proteins have been adequately separated into different fractions; the isolation of the major and minor proteins (L. Pedersen et al, 2003 and C.J. Fee 2006); however, the efficient purification of high-value minor proteins of similar molecular weights such as Lactoperoxidase and lactoferrin still remains as a challenge. A number of proteins observed in milk under various conditions exhibit antimicrobial property (Palmer, D. J *et al.*, 2006). For instance protein consisting of immunoglobulin (antibodies), lactoferrin, Lactoperoxidase and lysozyme are protecting proteins which can be crucial in the switch of passive immunity from the mother to the neonate. The immunoglobulin protects the neonate from infection till it'll expand their own immune device. Immunoglobulin's are a factor of the natural protection mechanism (Zasloff,

M. 2002). Immunoglobulins are located in high fixations in Colostrum the number one milk, and in low focuses in milk. However the immunoglobulin's, unique proteins located in milk are thought to have antimicrobial sporting events (Stelwagen, k., and D. J. Ormrod. 1998). Immune factors in colostrum and milk also play an important role in the host defense of the mammary gland itself protecting it from pathogenic organisms (Sordillo *et al.*, 1997; Oviedo-Boyso et al., 2007).

Lactoferrin and lactoperoxidase are a protein of the innate immune system and its natural or synthetic N-terminus derivatives have been proved to have antimicrobial, anticancer and antiendotoxin activities directly or through their modulatory effect on innate immunity with a decisive role in the inflammation (Pulido D et,al 2012). As a part of this special issue entitled "Antimicrobial peptides as mediators of innate immunity", the present contribution provides an overview of the basic structure-function features of Lactoferrin and its peptides as antimicrobial compounds. In addition, the modulatory role of Lactoferrin on some components of nonspecific immunity is discussed, with special emphasis on experimental infections,

cancer and sepsis in models of the endotoxemia (Sharma S ET, al.2013).

Lactoferrin an iron-binding glycoprotein was first isolated from cow's milk and subsequently from human milk. Lactoferrin is present in large quantities in mammalian secretions such as milk, tears, saliva, and seminal fluid, as well as in some white blood cells. Is also one of the minor proteins naturally occurring in cow milk at an average concentration of about 0.2 grams/liter? In Colostrum the lactoferrin content can be as high as 0.5 to 1 grams/liter. Lactoferrin inhibits microorganisms by binding iron and making this essential component unavailable to microorganisms (Tomita *et.,al* 2002 and Walzem *et.,al* 2002).

Lactoperoxidase has an approximate molecular weight of 77.5 kDa (W.A. Rombauts et al 1967) with an isoelectric point approaching 9.5. Molecular weight of Lactoferrin is 78.0 kDa with an isoelectric point around 8.7. In addition, proteins exhibit susceptible structure that alters their functionality, thus these macromolecules should be processed as quickly as possible and in as few steps as possible.

In biotechnology industry, yield and bioactivity are directly associated to efficient processing (M.R. Ladisch, 2001 and H.G. Harrison, et al 2003). Despite of the significant effort that has been applied toward developing downstream processes, there are still issues that need further considerations before an industrial application will be viable. Some of these challenges include labor-intensive experimental work rule of thumbs and consequently optimization of downstream processes can cover up to 50–80% of total production costs (P. Knight 1989 and F.D .A.U .S.2004)

Therefore, the objective of this review is to provide valuable information on antimicrobial property of milk protein extracted from cow milk and its public health importance.

LITERATURE REVIEW

Characteristics of Cow's Milk Proteins

Similar to human milk, the main component of bovine milk that determines its nutritional value is protein. Cow's milk proteins are a heterogeneous mixture and can be divided into two basic groups, i.e. caseins and whey proteins (Miciński J, et al 2013). Caseins are the most plentiful proteins of cow's milk. Their content varies inside the range of 2.6%–2.8%, which represents approximately 79% of general milk

proteins. Casein fractions range in awareness, contents of phosphorus, amino acid composition, molecular weight and isoelectric point. Seventeen Casein from bovine milk may also result in inflammatory reactions in mucous membrane of patients with celiac illness, which can be identified by using the presence of anti-prolamin antibodies.

Therefore, casein epitopes is probably responsible for scientific signs. There are five casein fractions in cow's milk: α s1, α s2, β , κ , and γ , which represent 30%, 9%, 28%, 10% and more than 1% of the whole quantity of casein, respectively (Cabrera-Chávez F 2009 and Farell HM et al 2004).

Whey proteins constitute approximately 0.6% of milk composition. Whey proteins represent an important group with high nutritional and functional properties that have a positive effect on human body. Whey proteins include β -lactoglobulin, α -lactalbumin, lactoferrin, lactoperoxidase, lysozyme, bovine serum albumin, immune globulins, transferrin, proteose-peptones (J Król J, et al 2008 and Michalski MC et al 2006).

Innate Immune Response of Milk Protein

The innate immune system is the primary line of defense shielding the body from infectious pathogens before the adaptive immune system comes into play. It represents a complicated interaction of cellular and molecular techniques geared toward detecting and finally casting off harmful pathogens. The innate immune device of the bovine mammary gland has developed into a fantastically powerful host defense mechanism (Rainard and Riollot, 2006).

The innate immune system consists of physical-chemical barriers soluble factors and cells which together function to recognize a foreign agent. This type of immunity includes barriers such as the mucosal epithelia and skin, fluid phase elements such as complement, pentraxin, ficolins and collectins, and innate cells such as neutrophils, eosinophils, basophils, mast cells, Dendritic Cells, macrophages, Natural Killer cells and innate lymphoid cells (Medzhitov R et al 2000). Unlike the adaptive immune system, the elements of the innate system respond quickly to insults because their receptors do not require rearrangement and do not generate memory. The system is often referred to as a nonspecific response. The mechanisms by which the components of the innate immunity function in host defense have been extensively

studied (Levy O 2007 and Eliassen E et al, 2017).

The bovine mammary gland performs a lively function in regulating the awareness of the unique immunoglobulins found in colostrum and milk, despite the fact that the mammary epithelium itself does not synthesize immunoglobulins. While a small amount of the immunoglobulins might also enter colostrum and milk from the blood serum through the paracellular direction due to “leaky” intercellular tight junctions (Lacy-Hulbert et al., 1999), the sizeable majority of immunoglobulins enter via a selective receptor-mediated intracellular path.

Microorganisms or toxins that correctly enter an organism come across the cells and mechanisms of the innate immune system. The innate reaction is typically caused while microbes are recognized through pattern recognition receptors, which understand additives which can be conserved amongst large groups of microorganisms, (Medzhitov R 2007) or when broken, injured or confused cells send out alarm alerts a lot of which (however not all) are identified via the same receptors as those that recognize pathogens (Matzinger P 2002). Innate immune defenses are non-particular meaning these structures reply to pathogens in a regularly occurring way (Alberts B et al 2002). This system does not confer long-lasting immunity in opposition to a pathogen. The innate immune system is the dominant machine of host defense in maximum organisms (Litman GW et al 2005).

The principal components of milk whey of nutraceutical ability includes beta-lactoglobulin, alpha-lactalbumin, bovine serum albumin (BSA), lactoferrin (Lf), immunoglobulins (Igs), lactoperoxidase (Lp) enzymes, glycomacropetides (GMP), lactose and minerals. The composition of the whey liquid additionally relies upon upon the supply of milk e.g., whey derived from buttermilk consists of extra lipid sphingomyelin compared to the ones derived from cheese. Whey is a popular dietary protein complement that is widely known to possess antimicrobial activity, immune modulation, advanced muscle power and body composition. moreover, whey is understood to prevent cardiovascular ailment and osteoporosis (Sultan, S. et al 2016). The important proteins present in bovine Milk like whey come from the mammary gland that secretes b-lactoglobulin (b-LG), 7 a-lactalbumin (a-lactalbumin), and glycomacropetide (GMP), and from serum, like IgG1 and IgG2, IgA, IgE,

and IgM and albumin. Except their use in useful foods whey protein merchandise and extra in particular whey protein-derived products, had been shown to be efficient in certain pathologies (Mehraban, M.H 2013).

The human immune device is an exceedingly complicated collection of systems and procedures that protects the body towards damage via pathogens. Immunity operates at some of stages. The skin and mucosal membranes are the primary line of protection appearing as a bodily and chemical barrier to invaders.

The second one level of defense is the innate immune system in which macrophages and neutrophils of the innate immune system offer a strong, but non-unique defense against pathogens, such as the cytokine-mediated inflammatory reaction. Ultimately, the adaptive immune system provides a focused reaction to particular pathogens. It is divided into humoral immunity, mediated through B lymphocytes and their antibodies, and mobile-mediated immunity, mediated by using T lymphocytes (hall, 2015).

Immune System Modulation

Bovine Colostrum and milk are rich assets of immune additives that are contributed via both the acquired and innate immune structures. Those immune factors play a role in conveying passive immunity to the offspring and shielding host immunity of the mammary gland itself. Variability in immune additives in Colostrum and milk are due to animal elements and control elements. More and more, immune additives from Colostrum and milk are being exploited commercially as antimicrobial sellers. Furthermore, vaccination processes to reinforce the herbal concentrations of immune components provide incredible capability within the improvement of hyper immune milk-derived merchandise for prophylactic or therapeutic use in humans (okay. Stelwagen et al 2008).

It is now nicely set up that youngsters growing up on a farm less regularly increase allergic reactions and allergies (Mutius E, Vercelli D.2010). Of the exceptional environmental factors investigated in those epidemiological studies, touch with cattle, endotoxin tiers in residence dirt, and the consumption of farm milk (i.e., cow’s milk with an unknown heating status) showed the most powerful affiliation with the safety of childhood asthma and hypersensitivity (Mutius E, Vercelli D.2010 and Riedler J, et al 2001). The intake of farm milk turned into related to higher Treg numbers in

blood, which have been negatively related to bronchial asthma and serum IgE ranges (Lluis A, et al 2014). Moreover, expanded demethylation of the FOXP3 gene and improved FoxP3+ T cellular numbers were detected in PBMC cultures of kids who have been uncovered to farm milk, suggesting that farm milk intake induces an immune regulatory phenotype.

Whey proteins suppressed in vitro lymphocyte mitogenesis and alloantigen-brought about proliferation, when blanketed in mature murine lymphocytes answers (Barta, et al, 1991) changed whey protein awareness also can suppress the mitogen-stimulated secretion of c-interferon, as well as the surface expression of interleukin-2 receptor, while introduced to T and B lymphocyte cultures (pass & Gill, 1999). Then again, Mercier, Gauthier, and Fliss (2004) claimed that addition of whey proteins from micro filtered-WPI to cellular tradition media, at a awareness of a hundred Ig/ml, stimulates in vitro proliferation of murine spleen lymphocytes.

Antimicrobial Effect of Milk Proteins

Milk whey additionally called cheese whey (Lactoserum) carries numerous precise additives that own a few critical antimicrobial and antiviral properties. Those are immunoglobulins, lactoferrin, lactoperoxidase, glycomacropeptide and sphingolipids. Some of these compounds are able to continue to exist after their passage through stomach and small intestine and exert their biological results within the massive gut. The crucial whey components that offer antimicrobial action within the intestinal tract are the immunoglobulins like IgG, IgM and IgA. IgG binds to toxin produced by means of *Clostridium difficile*, thereby decreasing diarrhea, dehydration and muscle aches.

Milk is a highest quality source of properly-balanced nutrients and indicates a numerous variety of organic function. Biological features of milk are especially due to milk peptides and proteins. Milk protein include of approximately 20% whey and 80% casein. Whey contains five predominant proteins, such as α -lactalbumin, glycomacropeptide, β lactoglobulin, proteose peptone three, immunoglobulins, and serum albumin, which together make up 85% of whey protein. Casein carries α s1-casein, α s2- casein, β -casein and κ -casein (okay Murakami & M Lagard 1998 and M Yamada et al 2002). The foremost antimicrobial proteins of milk are immunoglobulins, lactoferrin (Lf), lactoperoxidase (LP), and the lysozyme. Increase and acid production with the aid of starter cultures may

be inhibited by bacterial viruses, bacteriophages, or brought substances including antibiotics, sterilant and detergent residues, or free fatty acids produced via or as a result of the boom of microorganisms and natural often referred to as indigenous antimicrobial proteins.

Immunoglobulin

Immunoglobulins (Igs) in milk are an important defense own family of proteins for the newborn evidently protective the gut mucosa against pathogenic microorganisms. They inactivate micro organism via binding to specific sites on the bacterial floor their function is to confer passive immunity to the new newborn while its immune system is developing (Gapper et al. 2007). Three classes of immunoglobulins are typically located in milk. immunoglobulin G (IgG) the primary immunoglobulin in equine Colostrum (IgA) the main form in equine milk (Uniacke-Lowe et al. 2010) and M (IgM); IgG is often subdivided into two subclasses, IgG1 and IgG2 (Hurley 2003). Immunoglobulins (Igs) constitutes a complicated group, the factors of which are produced via B-lymphocytes; they make a significant contribution to the whey protein content material except exerting an essential immunological feature (mainly in colostrums).

These proteins are present the serum and physiological fluids of all mammals some of them attach to surfaces, where they behave as receptors, whereas others feature as antibodies, which are released inside the blood and lymph. IG are challenge to postnatal transfer via Colostrum as the placenta does now not allow passage of macromolecules (Butler, 1994).

The structure and fashionable feature of bovine Ig have been reviewed via Korhonen, Marnila, and Gill (2000). The range 25,000 kDa and two heavy chains (with molecular weight of 50,000–70,000 kDa) (Mulvihill & Donovan, 1987). The nomenclature of the elements of this family is based totally on their immunological cross-reaction with reference proteins, ideally of human beginning, as proposed via WHO (Butler, 1971). There are, but, three basic instructions of Ig: IgG, IgA and IgM, although IgG is regularly sub-divided into two subclasses IgG1 and IgG2. up to 80% (w/w) of all Ig in milk or whey is accounted for by IgG (de Wit, 1989) but qualitatively, the family of Ig found in bovine whey and colostrum include IgA and secretory IgA, IgG1, IgG2 and IgG fragments, IgM, IgE, J-chain or component, and free secretor aspect.

Structural and Property of Immunoglobulin

The immunoglobulins (Ig) seek advice from a heterogeneous circle of relatives of glycoproteins ranging in size from 15 to 1000 Kd that proportion commonplace antibody activity. One Ig consists of four instructions: IgG1 and IgG2, IgA, IgM, and IgE. These were diagnosed in milk, and enter it from blood serum. One those proteins are monomers of 20 Kd polypeptide chains and 50 to 70 Kd polypeptide chains which can be go linked with the aid of disulfide bonds. Cow milk incorporates 06 to 1.0 g L⁻¹ of Ig, 80% of that's IgG. (Eigel, W et al 1984 and de Wit, J. N 1989). Those proteins exhibit a higher denaturation (unfolding) temperature than a-la and P-Lg, but in the presence of other whey proteins they are extremely thermo labile, which may be due to interplay with P-Lg and BSA via disulfide bond formation. IgM has the capacity to engage with and agglutinate MFG in bloodless milk and additionally bind bacteria and fatty substances in desalted whey (Eigel, W et al 1984).

Antibacterial Activity of Immunoglobulin

Antibody performs a critical role in immunologic defenses against many micro organism, and patients who have a deficiency of immunoglobulin are at elevated risk to developing extreme bacterial infections (Winkelstein, J et al 1974). Currently, human immunoglobulin preparations suitable for intravenous administration were evolved. It's far important to ensure that this immunoglobulin keeps practical antimicrobial hobby and is able to enhancing host immunity.

Those four chains are joined together with disulphide bonds. The complete Ig or 'antibody' molecule has a molecular weight of approximately one hundred eighty kDa. The two same antigen-binding web sites are formed through the N-terminal part of one heavy and one light chain. The bovine IgG molecule takes place predominantly in subclasses: IgG1 and IgG2. Monomeric IgM and IgA have a similar primary shape to IgG except for the addition of a C-terminal octapeptide to the heavy chains. IgA occurs as a monomer or dimer, the latter comprising IgA molecules joined together via a J-chain and a secretory element. This complicated is referred to as secretory IgA (SIgA) and has a molecular weight of approximately 380 kDa. Except for ruminant lacteal secretions, IgA is the dominating Ig in all outside secretions of the body. IgM includes five subunits, just like monomeric IgA, which might be connected together in a circular mode by way of disulphide

bonds and a Jchain; the molecular weight of pentameric IgM is approximately 900 kDa (Larson BL 1992). These days, industrial colostrum whey-derived immunoglobulin preparations are extensively available within the market that are advertised underneath the category of feed supplements and newborn farm animal substitutes (Scammell, A.W 2001).

Lactoferrin

Lactoferrin is an iron-chelating multifunctional glycoprotein of about 80 kDa devoted to the safety of mammals towards pathogenic aggressors. This cationic glycoprotein belongs to the transferrin own family of iron chelators, such as serum transferrin and avian ovotransferrin. Lactoferrin indicates the highest avidity for iron of the transferring and its three-D structure is very just like different transferring, but it's miles enormously undoubtedly charged inside the N-terminal region. Lf is almost free of iron (10-15% iron, apo-Lf) in colostrum, milk, and mucosal secretions which include the ones from the intestinal, breathing and reproductive tracts, in addition to in tears and saliva. It is produced by means of the acinar cells of glands (Legrand D 2016 and Flanagan JL, et al 2009). Lf is a chief protein in milk.

Lactoferrin, an iron binding protein present in milk whey, possesses antibacterial properties. It sequesters iron from bacteria (Troost et al., 2001). Seeing that pathogens mainly have excessive iron requirements for metabolism and growth, these belongings of lactoferrin makes it widely antimicrobial in nature. Lactobacilli can utilize lactoferrin-bound iron, as a result allowing lactoferrin to both inhibit pathogenic bacteria and assist the boom of lactobacilli (Troost, F.J. et al 2001). It also glycoprotein found in maximum organic fluids of mammals together with milk, saliva, tears, synovial fluid and the secondary granules of neutrophils and blood and mucous secretions and released from activated neutrophils within the inflammatory re-sponse. It has broad spectrum antimicrobial properties and is broadly considered to be an critical component of the host protection towards microbial infections (Lønnerdal 2003).

Iron-binding proteins exert many physiological functions in biological systems. Many of these proteins are involved in the transport of iron within the body and its storage in certain compartments. At the same time, they minimize the prooxidant effect of iron. Other iron-binding proteins are enzymes that require iron as a cofactor for optimal activity. Finally, some iron-

binding proteins have been shown to regulate gene transcription and mRNA stability (Leibold EA, Guo B. 1992)

Structure and Properties of Lactoferrin

Lactoferrin is a glycoprotein with a molecular weight of approximately 80 kDa, which suggests excessive affinity for iron. It's miles a easy polypeptide chain folded into symmetrical lobes (N and C lobes) which might be distinctly homologous with one another (33–41% homology). These two lobes are connected via a hinge place containing elements of a helix between amino acids 333 and 343 in human lactoferrin which provides extra flexibility to the molecule.

The polypeptide chain includes amino acids 1–332 for the N lobe and 344–703 for the C lobe and is made from helix and pleated sheet structures that create two domain names for every lobe (domain names I and II). Each lobe can bind a metallic atom in synergy with the carbonate ion. The metals that it binds are the Fe²⁺ or Fe³⁺ ions, but it has additionally been discovered certain to Cu²⁺, Zn²⁺ and Mn²⁺ ions (Shanbacher FL et al 1992 and van der Strate BWA et al 2001).

Antibacterial Activity of Lactoferrin

Lactoferrin plays a key role in maintaining cellular iron levels in the body mostly in milk, several functions have been attributed to lactoferrin. It is considered a key component in the host's first line of defense, as it has the ability to respond to a variety of physiological and environmental changes. The structural characteristics of LF provide functionality in addition to the Fe³⁺ homeostasis function common to all transferrins: strong antimicrobial activity against a broad spectrum of bacteria, fungi, yeasts, viruses and parasites. Anti-inflammatory and anticarcinogenic activities and several enzymatic functions (Kanyshkova TG et al 2003)

Lactoferrin is considered to be a part of the innate immune system. At the same time, lactoferrin also takes part in specific immune reactions, but in an indirect way (Legrand et al., 2005). The antibacterial activity of Lactoferrin has been widely documented both in vitro and in vivo for Gram-positive and Gram-negative bacteria and in some acid–alcohol-resistant bacteria. Lactoferrin's bacteriostatic function is due to its ability to take up the Fe³⁺ ion, limiting use of this nutrient by bacteria at the infection site and inhibiting the growth of these

microorganisms as well as the expression of their virulence factors. Lactoferrin's bactericidal function has been attributed to its direct interaction with bacterial surfaces. Its ability to bind free iron, which is one of the elements essential for the growth of bacteria, is responsible for the bacteriostatic effect of lactoferrin (Reyes RE et al 2005). Due to its strategic position on the mucosal surface lactoferrin represents one of the first defense systems against microbial agents invading the organism mostly via mucosal tissues. Lactoferrin affects the growth and proliferation of a variety of infectious agents including both Gram-positive and negative bacteria, viruses, protozoa, or fungi (Kirkpatrick et al., 1971).

Nevertheless, some bacteria are able to adapt to the new conditions and release siderophores (iron chelating compounds of bacterial origin) that compete with lactoferrin for Fe³⁺ ions. Some other types of bacteria, including Neisseriaceae family, adapt to new conditions by expressing specific receptors capable of binding lactoferrin, and to cause changes in the tertiary structure of the lactoferrin molecule leading to iron dissociation (Schryvers et al., 1998; Ekins et al., 2004)

Even a bactericidal effect of lactoferrin has been described. This bactericidal activity is not iron-dependent and may be mediated through more than one pathway. Receptors for the N-terminal region of lactoferrin have been discovered on the surface of some microorganisms. The binding of lactoferrin to these receptors induces cell-death in Gram-negative bacteria due to a disruption in the cell wall.

The subsequent release of lipopolysaccharide (LPS) leads to impaired permeability and a higher sensitivity to lysozyme and other antimicrobial agents. LPS can be disposed of even without the direct contact of lactoferrin with the cell surface (Rossi et al., 2002). Bactericidal activity affecting Gram-positive bacteria is mediated by electrostatic interactions between the negatively charged lipid layer and the positively charged lactoferrin surface that cause changes in the permeability of the membrane (Valenti and Antonini, 2005).

It has been discovered that lactoferricin, a cationic peptide generated by the pepsin digestion of lactoferrin, has more potent bactericidal activity than the native protein. There are two forms known at present: lactoferricin H (derived from human lactoferrin) and lactoferricin B (of bovine origin) (Bellamy et al., 1992). The

proteolytic activity of lactoferrin is considered to inhibit the growth of some bacteria such as *Shigella flexneri* or enteropathogenic *E. coli* through degrading proteins necessary for colonization. However, this can be disabled by serine protease inhibitors (Orsi, 2004; Ward et al., 2005)

Lactoperoxidase

Lactoperoxidase is an vital piece of the nicely developed creature normal shield framework, and is located in diverse emissions consisting of tears and spit, and in milk at a grouping of round 30 mg/l, speaking to more or less 1% w/w of the whey proteins. Lactoperoxidase is also an individual from the peroxidase circle of relatives and a meeting of chemical compounds broadly discovered in nature that use hydrogen peroxide to oxidize thiocyanates to hypothiocyanate atomic load of Lactoperoxidase is round 78 000 Da and it contains diverse starch bunches while lactoperoxidase, hydrogen peroxide and thiocyanate particle are to be had collectively, the lactoperoxidase-catalyzed response yields center individual oxidation results of thiocyanate, prominently hypothiocyanate, which display antimicrobial impacts against microscopic organisms, parasites and infections (Seifu et al., 2005; Madureira et al., 2007).

The mix of Lactoperoxidase, hydrogen peroxide and thiocyanate is named the 'lactoperoxidase framework'. In natural liquids, the thiocyanates are available in warm blooded animal discharge from the ingestion of glucosinolate and cyanogenic glycoside containing feeds and hydrogen peroxide is available because of generation by polymorphonuclear leucocytes amid phagocytosis, creation by different microorganisms and as a result of explicit oxidation responses (Madureira et al., 2007).

Lactoperoxidase may be secluded from milk or whey utilizing particle trade innovations. The most broadly perceived utilization of the lactoperoxidase framework is in the safety of crude milk amid capability and delivery.

The lactoperoxidase framework can likewise be utilized as an antimicrobial pre-treatment to permit low-temperature warm remedy of warmth sensitive nourishments and refreshments. The lactoperoxidase framework has moreover been accounted for to be gainful inside the annihilation of microorganisms from new meat surfaces, in the safeguarding of corrective objects, in lessening plaque series, gum disease and carious accidents (Seifu et al., 2005).

Structure and Properties of Lactoperoxidase

Bovine LP includes a single polypeptide chain containing 612 amino acid residues. Its amino acid series is understood and the molecular weight is approximately 78 kDa. Lactoperoxidase is a simple protein having an excessive isoelectric point of 9.6 contributes to the humoral immune defense against pathogens by oxidation of thiocyanate and iodide (Flemmig, J et al 2016). The amino acid composition has the same opinion closely with that pronounced via Carlstrom (1969). The latter author calculated a molecular mass of 78,030 Da, along with the heme institution and carbohydrate residues Dull et al. (1990). Deduced an amino acid collection for bLPO from cDNA libraries. The deduced sequence concurs carefully with the series suggested via Cals et al. (1991). The enzyme includes a single polypeptide chain containing 612 amino acid residues, with 15 half of-cystines and 4-5 potential N-glycosylation sites. The peculiar range of half of-cystines is constant with different proof suggesting a disulfide bond between the heme organization and the peptide chain. However, (Rae and Goff 1998) have proven that the heme prosthetic group of bLPO is the iron complicated of one,5-bis (hydroxyl ethyl)-three,8-dimethyl-2, four-divinylporph.

The lactoperoxidase molecule has a carbohydrate content of about 10 % and possesses five potential N-glycosylation sites. The observed chromatographic and electrophoretic heterogeneity of LP \pm at least ten fractions of lactoperoxidase were identified \pm is possibly due to changes during the isolation process. Some of the glycosidic fractions may be lost during the purification process and a partial deamidation may be also a cause of heterogeneity. Nevertheless there is no significant difference in enzymatic activity between the various lactoperoxidase fractions (Paul & Ohlsson, 1985).

The natural importance of lactoperoxidase is its inclusion in the characteristic host protection framework against attacking smaller scale life forms. In cow-like milk it is one of the indigenous antimicrobial specialists. Alongside that antiviral movement, corruption of different cancer-causing agents and insurance of creature cells against peroxidative impacts have been accounted for. The response items created by the reactant activity of lactoperoxidase in the supposed lactoperoxidase framework are innocuous to mammalian cells (Reiter and Haernulv, 1984; Reiter and Perraudin, 1991;

Wolfson and Sumner, 1993; de Wit and van Hooydonk, 1996).

Antimicrobial Activity of Lactoperoxidase

The lactoperoxidase system can kill or inhibit the growth and metabolism of many species of microorganisms. Many cellular systems (i.e., outer membrane, cell wall, cytoplasmic membrane, transport systems, glycolytic enzymes, and nucleic acids) can be altered by the lactoperoxidase system. According to Pruitt and Reiter (1985), for any particular microorganism, the antimicrobial effects depend upon the reaction conditions. When adequate concentrations of lactoperoxidase are provided, bactericidal effects are greater at low temperatures (0- 5°C), at low pH (5 or less), and in the absence of reducing agents (Pruitt, K. M., and B. Reiter. 1985).

Different groups of bacteria show a varying degree of resistance to the lactoperoxidase. Gram-negative, catalase positive organisms such as pseudomonads, coliforms, salmonellae and shigellae are not only inhibited by the lactoperoxidase but, depending on medium conditions (pH, temperature, incubation time, cell density etc.), may be killed, provided H₂O₂ is supplied exogenously. Gram-positive, catalasenegative bacteria like streptococci and lactobacilli are generally inhibited but not killed by the activated lactoperoxidase.

This difference in sensitivity to the lactoperoxidase can probably be explained by the differences in cell wall structure and their different barrier properties. Mammalian cells are not affected by oxidation products of SCN₂ and it is suggested that the LP-s is not only atoxic to human cells but may protect these cells against toxic effects of H₂O₂ (Reiter & HaÈrnulv, 1984; Reiter & Perraudin, 1991; de Wit & van Hooydonk, 1996). Distinctive gatherings of microbes demonstrate a shifting level of protection from the lactoperoxidase. Gram-negative, catalase positive living beings, for example, pseudomonades, coliforms, salmonellae and shigellae are repressed by the lactoperoxidase as well as, contingent upon medium conditions (pH, temperature, brooding time, cell thickness and so on.), might be killed, if H₂O₂ is provided exogenously. Gram-positive, catalasenegative microscopic organisms like streptococci and lactobacilli are commonly restrained yet not slaughtered by the enacted lactoperoxidase. This distinction in affectability to the lactoperoxidase can presumably be clarified by the distinctions in cell divider structure and their diverse

obstruction properties (Reiter and HaÈrnulv, 1984; Reiter and Perraudin, 1991; de Wit and van Hooydonk, 1996).

The antibacterial activity/mechanism of the lactoperoxidase system is well-documented. It is the major intermediary product of the lactoperoxidase system reaction, hypothiocyanite (OSCN), which oxidizes essential protein sulfhydryls, resulting in altered cellular system functions and which causes inhibition of growth and/or death of the microorganism (Pruitt, K et al 1982). The hypothiocyanite ion can be formed by mixing the components (lactoperoxidase, thiocyanate, and hydrogen peroxide) of the lactoperoxidase system together.

Lysozyme

Lysozyme is bounteous in emissions including tears, salivation, human milk, and bodily fluid. It is additionally present in cytoplasmic granules of the macrophages and the polymorphonuclear neutrophils (PMNs). A lot of lysozyme can be found in egg white. C-type lysozymes are firmly identified with alpha-lactalbumin in arrangement and structure, making them part of a similar family (Williams S, and Vocadlo D 2017). In people, the lysozyme chemical is encoded by the LYZ quality. Lysozyme activity is nearly undetectable in cow milk, but very high in human milk (0.12 grams/liter). The concentration of lysozyme is highest in human colostrum and pre-colostral milk. The limited lysozyme activity in cow milk increases due to mastitis and high somatic cell counts. Heating cow milk at 75°C for 15 minutes destroys 25 percent of the activity of this enzyme. However, human milk lysozyme is more heat stable than cow milk lysozyme. (Yoshimura K et al, .1988 and Peters CW et al., 1989)

Lysozyme is a singular polypeptide chain involving 129 amino acids, in which lysine is the N-end amino destructive and leucine is the C-end one. It is a globular essential protein depicted by nuclear heap of 14.3 kDa and cross-associated by four disulfide securities (Masschalck, B et al., 2002 and Cegielska, R.R et al., 2008). It is a basic antimicrobial administrator in milk, which disposes of organisms by separating the β-1,4-glycosidic bond between C-1 of N-acetyl muramic destructive and C-4 of N-acetyl glucosamine stores of the peptidoglycan in the bacterial cell divider. Lysozyme is among the minor milk proteins that has pulled in extended thought starting late in light of its solid antimicrobial activity against a wide extent of microorganisms and from this time forward

potential in sustenance defending and security. Lysozyme has antibacterial action against various microscopic organisms.

This compound ordinarily works in relationship with lactoferrin or immunoglobulin A. Lysozyme is compelling against *Escherichia coli* working together with immunoglobulin A. It causes lysis of certain types of salmonellae in relationship with ascorbate and peroxide, the two of which are available in low focuses in milk.

Microwave illumination can diminish the movement of lysozyme against *Escherichia coli*. What's more, lysozyme can constrain the movement of neutrophils into harmed tissue and may work as a calming operator. (Zhao J and Li, D et al., 2011).

Structure and Properties of Lysozyme

Lysozyme represents one of the most appreciably investi-gated models for expertise the mechanism of protein stability, folding and denaturation 1-14. Like most proteins, denatured lysozyme may include a combination of conformation-al isomers that exist in a state of thermodynamic equilibrium (Dill, k.A. and Shortle, D. 1991 & Tanford, C. 1968). Further understanding of the molecular mechanism of two-stage unfolding of lysozyme will require structural analysis of denatured lysozyme and fractionation of diverse populations of conformational isomers that constitute the denatured lysozyme.

In this report, the mechanism of denaturation and un-folding of lysozyme has been analyzed by the technique of disulfide scrambling. In the presence of denaturant and a thiol initiator, the native lysozyme denatures by using shuffling its local disulfide bonds and converts to a aggregate of fully oxidized scrambled isomers that are trapped by way of non-native disulfide bonds. Lysozyme includes four disulfide bond and may also undertake 104 viable scrambled isomers. The technique of disulfide scrambling affords foremost advantages for characterization of the denaturation of lysozyme. (Chang, J.-Y. 2001).

Antimicrobial Activity of Lysozyme

Lysozyme antimicrobial activity relies at the potential of hydrolyzing the b-1, four linkages between N-acetyl muramic acid and N-acetyl glucosamine found in peptidoglycan. Gram-advantageous bacteria are very susceptible to lysozyme, due to the fact their cell wall is made up for 90% of peptidoglycan; in Gram-

negatives, peptidoglycan counts best for 5-10% of the cell wall and lies beneath the outer membrane of the cell envelope (Losso et al 2000). Nevertheless, lysozyme can be effective against Gram-negative bacteria in the presence of membrane destabilizing agents, such as detergents and chelators (Gill & Holley, 2000). This makes lysozyme by alone de facto ineffective within the safety of meals products where in the spoilage is due to Gram-terrible micro organism like Enterobacteriaceae and Pseudomonadaceae which might be surprisingly common contaminants of meat-based merchandise (Coma, 2008).

Gram-positive micro organisms are typically sensitive to lysozyme because of its enzymic action at the cell wall peptidoglycan. but, within the final 2 a long time evidence has amassed for a killing mechanism of Gram-positive micro organism unbiased of enzymic activity, wherein the cationic residences of lysozyme play an essential function and enzymic and lytic pastime are now not necessarily linked. Despite these multiple mechanisms to kill, some Gram-positive bacteria are resistant (Chris W. Michiels and Barbara Masschalck 2003).

Chromatography

The purpose of chromatography is to separate a complicated mixture into character element exploiting the partition effect which distribute the molecules into the distinctive phases. Chromatographic analysis of phosphate, in addition to not unusual inorganic anions, has traditionally been done using ion-exchange columns with alkaline eluents (carbonate/hydrogen carbonate answers or alkaline hydroxides) and conductivity detection (B. L'opez-Ruiz 2000 and J. Weiss, 2003).

Those systems had been installed because the technique of desire for recurring packages, and a big range of regulatory and official methods contain the use of this chromatography configuration (J.D. Pfaff 1993 and D.P. Hautman et al 1997). A mechanistic model is used to describe the physical phenomena based on a set of mathematical equations.

Two types of physical phenomena dominate chromatography: movement of solutes through the packed bed of porous particles via mass transfer mechanisms, and adsorption based on the fundamental thermodynamic interactions between migrating solutes and the stationary phase. The general system of equations used to describe the mass transfer phenomena consist of

two sets of partial differential mass conservation equations. The general rate model for a chromatographic process includes convective and diffusive flows through porous particles on the column level and imitates mass transfer resistances and surface interactions on particle level. In Ion Exchange Chromatography (IEC), an external film surrounding adsorbent particles is commonly presumed to model the movement of components from column to particle level; the sorption of protein on the particle surface can be described by the steric mass-action (SMA) model, developed by and generally used for the modeling of salt gradient elution in Ion Exchange Chromatography (IEC), (Iyer et al., 1999).

Recently lactoperoxidase (LPOase) and lactoferrin (Lf), presents preparation great difficulties because of its fragility, which tends to increase with increasing purity. Lactoperoxidase has been isolated and purified using cation-exchange chromatography, such as carboxymethyl (CM) cellulose at pH 5.1 & pH 5.7 (Langbakk, B., and T. Flatmark. 1989).

Previously Lf has been separated by DEAE anion-exchange chromatography. It has also isolated using CM-Sephadex at pH 7.0 (Johansson, B. 1969), cellulose phosphate, heparin Sepharose and metal chelate interaction chromatography.

The presence of LPOase in the OD fraction was estimated by absorbance of yellowish color at 412 nm (Yoshida, S. 1988). When the OD fraction eluted from the BTP-650M matrix, 350 ml of the deeply colored fractions were collected from 3.3 L of 1.8 M AS whey. The OW fraction was used as the starting material for CM cation exchange chromatography. This fraction (protein concentration 1.60%) was passed through the column of CM-TP at pH 7.7. Most of the whey proteins eluted as unbound proteins. After the calculation of protein concentration from absorbance at 280 nm Lactoperoxidase, Lactoferrin-a and Lactoferrin - b were estimated at 28.6, 7.2 and 14.9 mg respectively. Lactoperoxidase and Lactoferrin distributed not only in the OD fraction but also in the amino acid fraction (Yoshida, S. 1989); therefore, these proteins contents did not agree with the data for untreated acid whey described later.

CONCLUSION AND RECOMMENDATION

In the light of these first findings, the antimicrobial properties of Cow milk, whey proteins against

bacteria, both Gram positive and Gram negative, seem very promising. In many cases the role of the single whey protein (Lactoferrin, lysozyme, Lactoperoxidase and Immunoglobulin) as well as its mechanism of action remain to be fully clarified and Gram-positive micro organisms are typically sensitive to lysozyme because of its enzymic action at the cell wall peptidoglycan. Nonetheless, these arguments reinforce the claim of Cow milk as functional food with diverse bioactivities.

Today, the purification of high-value proteins extracted from cow milk has attracted a great attention. Due to the high number of potential applications of protein isolates, a few chromatographic processes have been developed to isolate high-purity protein fractions.

Ion exchange chromatography (IEC) is one of the most powerful techniques to overcome bio molecules purifying challenges and is commonly applied in downstream processes (B. Ekstrand 1989). As a recommendation research is needed to determine the amount of milk protein that has optimal antimicrobial effective and cost effectiveness in promoting growth and to determine if there are any advantages of using whey instead of dried skimmed milk in the treatment of children with moderate malnutrition.

REFERENCES

- [1] Alberts B, Johnson A, Lewis J, Raff M, Roberts K, Walters P (2002). *Molecular Biology of the Cell* (Fourth ed.). New York and London: Garland Science. ISBN 978-0-8153-3218-3.
- [2] Amercham Biosciences. *Ion Exchange chromatography, Principles and methods*, Amercham Pharmacia. Biotech SE. 2002; 751.
- [3] B. Ekstrand, Antimicrobial factors in milk a review, *Food Biotechnol.* 3 (1989) 105 - - 126. Doi: 10.1080/08905438909549703.
- [4] B. L'opez-Ruiz, J. *Chromatogr. A* 881 (2000) 607.
- [5] Barta, O., Barta, V. D., Crisman, M. V., & Akers, R. M. (1991). Inhibition of lymphocyte blastogenesis by whey. *American Journal of Veterinary Research*, 52, 247-253.
- [6] Bellamy W., Takase M., Yamauchi K., Wakabayashi H., Kawase K., Tomita M. (1992): Identification of the bactericidal domain of lactoferrin. *Biochimica et Biophysica Acta*, 1121, 130-136.
- [7] Butler, J. E. (1971). *Physicochemical and immunochemical studies on bovine IgA and*

- glycoprotein-a. *Biochimica et Biophysica Acta*, 251, 435–449.
- [8] Butler, J. E. (1994). Passive immunity and immunoglobulin diversity. In *Indigenous Antimicrobial Agents of Milk – Recent Developments*. IDF Special Issue, 94044, 14–50.
- [9] C.A. Barth, S. E., Milk proteins: nutritional, clinical, functional and technological aspects, Darmstadt, 1988.
- [10] C.J. Fee, A. Chand, Capture of lactoferrin and lactoperoxidase from raw whole milk by cation exchange chromatography, *Sep. Purif. Technol.* 48 (2006) 143–149. doi:10.1016/j.seppur.2005.07.011.
- [11] Cabrera-Chávez F, Calderón de la Barca AM. Bovine milk intolerance in celiac disease is related to IgA reactivity to α and β -caseins. *Nutrition*. 2009; 25(6):715–716.
- [12] Cals, M.-M., Mailliart, P., Brignon, G., Anglade, P. and Dumas, B. (1991) Primary structure of bovine lactoperoxidase, a fourth member of a mammalian heme peroxidase family. *Eur. J. Biochem.*, **198**, 733–9.
- [13] Carlström, A. (1969) Physical and compositional investigations of the sub fractions of lacto peroxidase. *Acta. Chim. Scand.*, 23, 185–202.
- [14] Cegielska, R.R.; Lesnierowski, G.; Kijowski, J. Properties and application of egg white lysozyme and its modified γ preparations—A review. *Polish Journal of Food and Nutrition Sciences* 2008, 58 (1), 5–10.
- [15] Chang, J.-Y. And Li, L. (2001) *J. Biol. Chem.* 276, 9705–9712.
- [16] Chris W. Michiels and Barbara Masschalck (2003). Laboratory of Food Microbiology, Katholieke Universiteit Leuven, Kasteelpark Arenberg 22, B-3001 Leuven, Belgium.
- [17] Chromatographic and electrophoretic methods analysis of milk proteins *Journal of Chromatography*, 624 (1992) 81-102.
- [18] Clinical and Laboratory Standards Institute. 2006. Approved standard: M2-A9. Performance standards
- [19] Coma, V. (2008). Bioactive packaging technologies for extended shelf life of meat based products. *Meat Science*, 78, 90e103.
- [20] Cross, M. L., & Gill, H. S. (1999). Modulation of immune function by a modified bovine whey protein concentrates *Immunology and Cell Biology*, 77, 345–350.
- [21] D.P. Hautman, D. Munch, J.D. Pfaff, US EPA Method 300.1, Determination of Inorganic Anions in Drinking Water by Ion Chromatography, EPA/600/R-98/118, NIST PB-98-169196INZ, 1997.
- [22] De Wit JN & van Hooydonk ACM (1996) Structure, functions and applications of lactoperoxidase in natural antimicrobial systems. *Netherlands Milk & Dairy Journal* 50, 227±244.
- [23] De Wit JN & van Hooydonk ACM (1996) Structure, functions and applications of lactoperoxidase in natural antimicrobial systems. *Netherlands Milk & Dairy Journal* 50, 227±244.
- [24] De Wit, J. N. (1998). Nutritional and functional characteristics of whey proteins in food products. *Journal of Dairy Science*, 81, 597–602.
- [25] de Wit, J. N., Functional properties of whey proteins, in *Developments in Dairy Chemistry*, Vol. 4, Fox, P. F., Ed., Elsevier Applied Science, New York, 1989, 285.
- [26] Dill, K.A. and Shortle, D. (1991) *Annu. Rev. Biochem.* 60, 795–825.
- [27] Dupas, c., adt, i., cottaz, a., boutrou, r., molle, d., jardin, j., jouvet, t. & degraeve, p. (2009): A chromatographic procedure for semi-quantitative evaluation of casein phosphopeptides in cheese. *Dairy Sci. Technol.*, 89, 519–529.
- [28] Eigel, W. N., Butler, J. E., Ernstrom, C. A., Farrell, H. M., Jr., Harwalkar, V. R., Jenness, R., and Whitney, R. McL., Nomenclature of proteins of cow's milk: fifth revision, *J. Dairy Sci.*, 67, 1599, 1984.
- [29] Ekins A., Khan a.g., shouldice s.r., schryvers a.b. (2004): Lactoferrin receptors in gram-negative bacteria: insights into the iron acquisition process. *Biometals*, 17, 235–243.
- [30] Eliassen E, Di Luca D, Rizzo R, Barao I. The Interplay between Natural Killer Cells and Human Herpesvirus-6. *Viruses* 2017; 9(12): 367.
- [31] F.D.A.U.S. Department of Health and Human Services, Guidance for Industry PAT–A Framework for Innovative Pharmaceutical Manufacturing and Quality Assurance. (2004).
- [32] Farell HM, Jimenez-Flores R, Bleck GT, Brown EM, Butler JE, Creamer LK, et al. Nomenclature of the proteins of cow's milk – sixth revision. *J Dairy Sci.* 2004; 87(6) : 1641–1674.
- [33] Flanagan JL, Willcox MDP. Role of lactoferrin in the tear film. *Biochimie* 2009; 91(1): 35-43.
- [34] Flemmig, J.; Gau, J.; Schlorke, D.; Arnhold, J. Lactoperoxidase as a potential drug target. *Expert Opin. Ther. Targets* 2016, 20, 447–461. [CrossRef] [PubMed] for antimicrobial disk susceptibility tests, 9th ed. CLSI, Wayne, Pa.

- [35] Gapper LW, Copestake DEJ, Otter D, Indyk HE (2007) Analysis of bovine immunoglobulin G in milk, colostrum and dietary supplements: a review. *Anal Bioanal Chem* 389:93–109.
- [36] Gill, A. O., & Holley, R. A. (2000). Inhibition of bacterial growth on ham and bologna by lysozyme, nisin and EDTA. *Food Research International*, 33, 83e90.
- [37] H.G. Harrison, P. Todd, S.R. Rudge, P.P. Demetri, *Bioseparations and Engineering*, Oxford University Press, Oxford, 2003.
- [38] Hall, J. E. (2015). *Guyton and Hall textbook of medical physiology*. Elsevier Health Sciences
- [39] Hurley WL (2003) Immunoglobulins in mammary secretions. In: Fox PF, Mc Sweeney PLK (eds) *Advanced dairy chemistry: proteins*, vol 1, 3rd edn. Kluwer Academic/Plenum Publishers, New York
- [40] INPPAZ. Taller Internacional sobre Vigilancia de Salmonella y la resistencia antimicrobiana en patógenos transmitidos por los alimentos. 2000. Buenos Aires, Argentina.
- [41] Iyer, H.; Tapper, S.; Lester, P.; Wolk, B.; van Reis, R. Use of the steric mass action model in ion-exchange chromatographic process development. *J. Chromatogr. A* 1999, 832, 1–9. [CrossRef].
- [42] J. Weiss, D. Jensen, *Anal. Bioanal. Chem.* 375 (2003) 81.
- [43] J.D. Pfaff, US EPA Method 300.0, Methods for the Determination of Inorganic Substances in Environmental Samples, EPA -600/R-93- 100, NIST PB-94-121811, August 1993.
- [44] Johansson, B. 1969. Isolation of crystalline lactoferrin from human milk. *Acta Chem. Scand.* 23:683.
- [45] K Murakami; M Lagarde; Y Yuki. *Electrophoresis*, 1998, 19: 2521–2527.
- [46] K. Stelwagen, E. Carpenter, B. Haigh, A. Hodgkinson and T. T. Wheeler. doi: 10. 2527 /jas.2008-1377 originally published online October 24, 2008. Immune components of bovine colostrum and milk. *Journal of animal science research*.
- [47] Kanyshkova TG, Babina SE, Semenov DV, Isaeva N, Valssov AV, Neustroev KN, et al. Multiple enzymatic activities of human milk lactoferrin. *Eur J Biochem* 2003; 270:3353–61.
- [48] Karlsson E, Ryden L, Brewer J Protein purification. Principles, High Resolution Methods, and Applications. Ion exchange chromatography. 2nd ed. New York: Wiley; 1998.
- [49] Kirkpatrick C.H., Green I., Rich R.R., Schade A.L. (1971): Inhibition of growth of *Candida albicans* by iron-unsaturated lactoferrin: relation to host-defense mechanisms in chronic mucocutaneous candidiasis. *The Journal of Infectious Diseases*, 124, 539–544.
- [50] Korhonen, H., Marnila, P., & Gill, H. (2000). Milk immunoglobulins and complement factors: A review. *British Journal of Nutrition*, 84, S1–S7.
- [51] Król J, Litwiń czuk A, Zarajczyk A, Litwiń czuk Z . Alfalaktoalbumina i beta - laktoalbumina jako związki biologiczne, czynne frakcji białkowej mleka [Alpha-lactalbumin and betalactalbumin as biologically active compounds of milk protein fraction]. *Med Weter.* 2008; 64 (12) :1375–1378.
- [52] L. Pedersen, J. Mollerup, E. Hansen, A. Jungbauer, Whey proteins as a model system for chromatographic separation of proteins, *J. Chromatogr. B.* 790 (2003) 161–173. doi:10.1016/S1570-0232(03)00127-2.
- [53] Lacy-Hulbert, S. J., M. W. Woolford, G. Nicholas, C. G. Prosser, and K. Stelwagen. 1999. Effect of milking frequency and pasture intake on milk yield and composition of late lactation cows. *J. Dairy Sci.* 82:1232–1239.
- [54] Laemmli, U. K., 1970. Cleavage of structural proteins during the assembly of the head bacteriophage T4. *Nature* 227:680.
- [55] Langbakk, B., and T. Flatmark. 1989. Lactoperoxidase from human colostrum. *Biochem. J.* 259:627.
- [56] Larson BL (1992) Immunoglobulins of the mammary secretions. In *Advanced Dairy Chemistry 1-Proteins*, pp. 231±254 [PF Fox, editors]. London: Elsevier Science Publishers
- [57] Legrand D. Overview of lactoferrin as a natural immune modulator. *J Pediatr* 2016; 173(Suppl.): S10-5. Elizabeth D. Strange *, Edyth L. Malin, Diane L. Van Hekken and Jay J. Basch.
- [58] Legrand D., Ellass E., Carpentier M., Mazurier J. (2005): Lactoferrin: a modulator of immune and inflammatory responses. *Cellular and Molecular Life Sciences*, 62, 2549–2559.
- [59] Leibold EA. Guo B. 1992. Iron-dependent regulation of ferritin and transferrin receptor expression by the iron-responsive element binding protein. *Annu. Rev. Nutr.* 3 12:345-8.
- [60] Levy O. Innate immunity of the newborn: basic mechanisms and clinical correlates. *Nat Rev Immunol* 2007; 7(5): 379-90.
- [61] Li, D.; Zhang, T.; Xu, C.; Ji, B. Effect of pH on the interaction of baicalein with lysozyme by spectroscopic approaches. *Journal of Photochemistry and Photobiology. B: Biology* 2011, 104, 414–424.

- [62] Lluís A, Depner M, Gaugler B, Saas P, Casaca VI, Raedler D, et al. Increased regulatory T-cell numbers are associated with farm milk exposure and lower atopic sensitization and asthma in childhood. *J Allergy Clin Immunol* (2014) 133:551–9. doi:10.1016/j.jaci.2013.06.034
- [63] Lönnerdal B. Infant formula and infant nutrition: bioactive proteins of human milk and implications for composition of infant formulas. *Am J Clin Nutr* 2014; 99(3): 712S–7S.
- [64] Losso, J. N., Nakai, S., & Charter, E. A. (2000). Lysozyme. In A. S. Naidu (Ed.), *Natural food antimicrobial systems* (pp. 185e210). Boca Raton (Florida): CRC Press LLC.
- [65] M Yamada; K Murakami; JC Wallingford; Y Yuki. *Electrophoresis*, 2002, 23: 1153–1160.
- [66] M.R. Ladisch, *Bioseparations Engineering: Principles, Practice, and Economics*, John Wiley & Sons, New York, 2001.
- [67] Madureira, a.r., pereira, c.i., gomes, a.m.p., pintado, m.e., and malcata, f.x. (2007) Bovine whey proteins – overview on their main biological properties. *Food Res. Int.* 40, 1197–1211.
- [68] Masschalck, B.; Deckers, D. Michiels. Lytic and nonlytic mechanism of inactivation of gram-positive bacteria by lysozyme under atmospheric and hydrostatic pressure. *Journal of Food Protection* 2002, 65, 916–923.
- [69] Matzinger P (Apr 2002). "The danger model: a renewed sense of self" (PDF) *Science* 296 (5566): 301–05 .Bibcode:2002Sci...296..301M. doi:10.1126/science.1071059. PMID 11951032.
- [70] Medzhitov R (Oct 2007). "Recognition of microorganisms and activation of the immune response". *Nature*.449 (7164):8196.Bibcode: 2007Natur.449..819M. doi: 10.1038 /nature 06246. PMID 17943118.
- [71] Medzhitov R, Janeway C Jr. Innate immunity. *N Engl J Med* (2000) 343(5): 338–44. doi:10.1056/NEJM200008033430506
- [72] Mehraban, M.H.; Yousefi, R.; Taheri-Kafrani, A.; Panahi, F.; Khalafi-Nezhad, A. Binding study of novel anti-diabetic pyrimidine fused heterocycles to β -lactoglobulin as a carrier protein. *Coll. Surf. B Biointerfaces* 2013, 112, 374–379. [CrossRef] [PubMed].
- [73] Mercier, A., Gauthier, S. F., & Fliss, I. (2004). Immuno modulating effects of whey proteins and their enzymatic digests. *International Dairy Journal*, 14, 175–183.
- [74] Michalski MC, Januel C. Does homogenization affect the human health properties of cow's milk? *Trends Food Sci Technol.* 2006; 17(8):423–437.
- [75] Miciński J, Zwierzchowski G, Kowalski I, Szarek J. Health promoting properties of selected milk components. *J Elementol.* 2013;18(1):165–186. http://dx.doi.org/10.5601/jelem.2013.18.1.14.
- [76] Murray, Baron, Jorgensen, Landry and Pfaller (ed.). 2007. *Manual of clinical microbiology*, 9th ed. American Society for Microbiology, Washington, DC.
- [77] Orsi N. (2004): The antimicrobial activity of lactofer-rin: current status and perspectives. *Biometals*, 17, 189–196.
- [78] P. Knight, *Chromatography: 1989 Report, Bio/Technology.* 7 (1989) 243–249.
- [79] Paul KG & Ohlsson PI (1985) The chemical structure of lactoperoxidase. In *The Lactoperoxidase System, Chemistry and Biological Significance*, pp. 15±29 [KM Pruitt and JO Tenovuo, editors]. New York: Marcel Dekker.
- [80] Pruitt, K. M., and B. Reiter. 1985. Biochemistry of peroxidase system: antimicrobial effect. In K. M. Pruitt and J. Tenovuo (ed.), *the lactoperoxidase system: Chemistry and biological significance*. Marcel Dekker, New York.
- [81] Pruitt, K. M., and J. Tenovuo. 1982. Kinetics of hypothiocyanite production during peroxidase-catalyzed oxidation of thiocyanate. *Biochem. Biophys. Acta* 704:204-214.
- [82] Pulido D, Nogués MV, Boix E, Torrent M. Lipopolysaccharide neutralization by antimicrobial peptides: A gambit in the innate host defense strategy. *J Innate Immun* 2012; 4(4): 327-36.
- [83] Rae, T. D. and Goff, H.M. (1998) the heme prosthetic group of lactoperoxidase. *J. Biol. Chem.*, 272, 27968–77.
- [84] Rainard, P., and C. Riollot. 2006. Innate immunity of the bovine mammary gland. *Vet. Res.* 37:369–400.
- [85] Reiter B & HaÈrnulv G (1984) Lacto peroxidase antibacterial system: natural occurrence, biological functions and practical applications. *Journal of Food Protection* 47, 724±732.
- [86] Reiter B & HaÈrnulv G (1984) Lactoperoxidase antibacterial system: natural occurrence, biological functions and practical applications. *Journal of Food Protection* 47, 724±732.
- [87] Reiter B & Perraudin JP (1991) Lactoperoxidase: biological functions. In *Peroxydases in Chemistry and Biology*, pp. 143± 180. Boca Raton: CRC Press.
- [88] Reiter B & Perraudin JP (1991) Lactoperoxidase: biological functions. In *Peroxydases in Chemistry and Biology*, pp. 143± 180. Boca Raton: CRC Press.

- [89] Reyes RE, Manjarrez HA, Drago ME. El hierro and la virulencia bacteriana. *Enf Inf Microbiol* 2005; 25:104–7.
- [90] Riedler J, Braun-Fahrlander C, Eder W, Schreuer M, Waser M, Maisch S, et al. Exposure to farming in early life and development of asthma and allergy: a cross-sectional survey. *Lancet* (2001) 358:1129–33. doi:10.1016/S0140-6736 (01)06252-3 PubMed Abstract | CrossRef Full Text | Google Scholar.
- [91] Rosa L, Cutone A, Lepanto MS, Paesano R, Valenti P. Lactoferrin: A natural glycoprotein involved in iron and inflammatory homeostasis. *Int J Mol Sci* 2017; 18(9): 1985.
- [92] Rossi P., Giansanti F., Boffi A., Ajello M., Valenti P., Chiancone E., Antonini G. (2002): Ca²⁺ binding to bovine lactoferrin enhances protein stability and influences the release of bacterial lipopolysaccharide. *Biochemistry and Cell Biology*, 80, 41–48.
- [93] Scammell, A.W. Production and uses of colostrum. *Aust. J. Dairy Technol.* 2001, 56, 74–82.
- [94] schryvers a.b., bonnah r., yu r.h., Wong H., Retzer M. (1998): Bacterial lactoferrin receptors. *Advances in Experimental Medicine and Biology*, 443, 123–133.
- [95] Seifu, e., buys, e.m., and donkin, e.f. (2005) Significance of the lactoperoxidase system in the dairy industry and its potential applications: a review. *Trends Food Sci. Technol.* 16, 137–154.
- [96] Shagghi N, Palombo EA, Clayton AHA, Bhave M. Archetypal tryptophan-rich antimicrobial peptides: Properties and applications. *World J Microbiol Biotechnology* 2016; 32(2): 31.
- [97] Shanbacher FL, Goodman RE, Talhouk RS. Bovine mammary lactoferrin: implications from messenger ribonucleic acid (mRNA) sequence and regulation contrary to other milk proteins. *J Dairy Sci* 199276:3812–31.
- [98] Sharma S, Sinha M, Kaushik S, Kaur P, Singh TP. C-lobe of lactoferrin: the whole story of the half-molecule. *Biochem Res Int* 2013; 2013: 271641.
- [99] Sparsh Ganju and Parag R. Gogate, A review on approaches for efficient recovery of whey proteins from dairy industry effluents, *Journal of Food Engineering*, 215, (84), (2017).
- [100] Sultan, S.; Huma, N.; Butt, M.S.; Aleem, M.; Abbas, M. Therapeutic potential of dairy bioactive peptides: A Contemporary Perspectives. *Crit. Rev. Food Sci. Nutr.* 2016. [CrossRef] [PubMed]. Tanford, C. (1968) *Adv. Protein Chem.* 23, 121^282.
- [101] Troost, F.J.; Steijns, J.; Saris, W.H.; Brummer, R.J. Gastric digestion of bovine lactoferrin in vivo in adults. *J. Nutr.* 2001, 131, 2101–2104. [PubMed].
- [102] Uniacke-Lowe T, Huppertz T, Fox PF (2010) Equine milk proteins: chemistry, structure and nutritional significance. *Int Dairy J* 20:609–629
- [103] Valenti P., Antonini G. (2005): Lactoferrin: an important host defense against microbial and viral attack. *Cellular and Molecular Life Sciences*, 62, 2576–2587.
- [104] Van der Strate BWA, Belijaars L, Molema G, Harmsen MC, Meijer DK. Antiviral activities of lactoferrin. *Antiviral Res* 2001 52:225–39.
- [105] Von Mutius E, Vercelli D. Farm living: effects on childhood asthma and allergy. *Nat Rev Immunol* (2010) 10:861–8. doi:10.1038/nri2871 PubMed Abstract | Cross Ref Full Text | Google Scholar.
- [106] W.A. Rombauts, W.A. Schroeder, M. Morrison, Bovine Lactoperoxidase. Partial Characterization of the Further Purified Protein, *Biochemistry.* 6 (1967) 2965–2977. Doi: 10.1021/bi00862a002.
- [107] Ward p.p., Paz E., Conneely o.m. (2005): Multifunctional roles of lactoferrin: a critical overview. *Cellular and Molecular Life Sciences*, 62, 2540–2548.
- [108] Williams S, Vocadlo D. "Glycoside hydrolase family 22". *Cazypedia*. Retrieved 11 April 2017.
- [109] Winkelstein, J.; Drachman, R.: Phagocytosis: the normal process and its clinically significant abnormalities. *Pediat. Clins N. Am.* 21: 551-569 (1974).
- [110] Wolfson LM & Sumner SS (1993) Antimicrobial activity of the lacto peroxidase system. A review. *Journal of Food Protection* 56, 887±892.
- [111] Yoshida, S. 1988. Isolation of lactoperoxidase of 89,000 daltons and a globulin of 81,000 daltons from milk acid whey. *J. Dairy Sci.* 71:2021.
- [112] Yoshida, S. 1989. Preparation of lactoferrin by hydrophobic interaction chromatography from milt. *Acid whey. J. Dairy Sci.* 72:1446.
- [113] Zhang X, Zhivaki D, Lo-Man R. Unique aspects of the perinatal immune system. *Nat Rev Immunol* 2017; 17(8): 495-507.
- [114] Zhao, L.; Sun, J.-S.; Sun, L. The g-type lysozyme of *scophthalmus maximus* has a broad substrate spectrum and is involved in immune response against bacterial infection. *Fish and Shellfish Immunology* 2011, 30, 630–637.

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